

PREVALENCE AND ANTIBIOGRAM OF ESCHERICHIA COLI ISOLATED FROM URINARY TRACT INFECTIONS IN A TERTIARY CARE HOSPITAL

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ABSTRACT

One of the most common bacterial infections seen in clinical practice particularly in developing countries are the Urinary tract infection (UTIs). Most common organism responsible for UTI is E.coli. Despite the wide availability of antimicrobial therapy, UTI still remains a major health hazard. This was a retrospective study conducted in a tertiary care hospital to know the prevalence and antibiogram of E. coli. Samples received include mid-stream clean catch urine, suprapubic aspirate and from Foley's catheter. All the plates were inspected for growth and the isolates were identified by observing colony morphology, Gram-stain characteristics and relevant biochemical tests. The isolates were tested for their antimicrobial

susceptibility and the results were interpreted by modified Kirby Bauer disc diffusion method, according to the guidelines of Clinical and Laboratory Standards Institute. Out of 704 samples tested, 438 (62.21%) were positive for E.coli. E.coli was highly resistant to Ampicillin (91.7%) followed by Ofloxacin (66.6%) and least resistance was seen with Imipenem (2.73%) followed by Piperacillin-tazobactam (21.6%) and Amikacin (21.9%). Knowledge of prevalence and antimicrobial susceptibility pattern of E. coli will help in selecting an appropriate antibiotic for empirical therapy. Formulation of hospital infection control committee and strict adherence to the guidelines of the committee will help in preventing the emergence of drug resistance.

Key words: Urinary tract infection, Escherichia coli, antimicrobial susceptibility, bacteriuria.

INTRODUCTION

One of the commonest bacterial infections are the Urinary tract infections (UTIs), and the most common organism accounted for UTI is Escherichia coli (E.coli).^[1, 2] Despite the widespread availability of antibiotics, UTI is one of the most important causes of morbidity in general population and also the common cause of nosocomial infection among hospitalized patients.^[3] Treatment of UTIs can be difficult because of recurrences and asymptomatic infections. The susceptibility pattern of uropathogens have been changing over the past years, both in community and nosocomial infections.^[4,5]

Antibiotics are invariably used for the treatment of UTIs, though resistance to antibiotics has been reported all over the world, particularly in developing countries.^[6] Treatment of UTIs is a challenge due to the increasing level of antimicrobial resistance.^[7] There is an increased emergence of antimicrobial resistance in the uropathogens, probably due to the empirical administration of anti bacterial therapy, even before the availability of the urine culture results, is a matter of concern worldwide.^[8] The prevalence of antimicrobial resistance in patients with UTI is increasing and can vary according to geographical and regional location.^[9]

For treatment of UTIs and prevention of antimicrobial resistance, knowledge of the common organisms responsible for UTIs and their antibiotic susceptibility patterns in specific geographical locations will help physicians in choosing an appropriate empirical treatment. There are only few studies of prevalence of UTI and antibiogram of E.coli in this part, hence the present study was undertaken to find out the prevalence and antibiogram of UTI due to E.coli

MATERIAL AND METHODS

This was a retrospective study conducted in a tertiary care hospital in South India. Data was collected from March 2010 to March 2011 using the microbiological records of consecutive urine samples received in the laboratory during the study. All positive samples reports from both community acquired and nosocomial UTI was included from various specialities.

Urine samples were cultured using a 0.01 ml calibrated loop onto blood agar and MacConkey agar plates, incubated at 37°C for 18-24 hours and the number of colonies was counted. All specimens were inoculated on cystine lactose electrolyte-deficient medium, (Hi Media,

Mumbai) by semi quantitative method. The specimen yielding more than or equal to 10^5 organisms/ml of urine was interpreted as significant. Samples received include mid-stream clean catch urine, suprapubic aspirate and from Foley's catheter. All the plates were inspected for growth and the isolates were identified by observing colony morphology, Gram-stain characteristics and relevant biochemical tests.^[10]

The isolates were tested for their antimicrobial susceptibility and the results were interpreted by modified Kirby Bauer disc diffusion method, according to the guidelines of Clinical and Laboratory Standards Institute.^[11] The antibiotics tested were Ampicillin (10 µg), Gentamicin (10µg), Co-trimoxazole (1.25/23.75 µg), Ofloxacin (10 µg), Nalidixic acid (30 µg), Piperacillin-tazobactam (100µg/10 µg), Ceftriaxone (30 µg), Ceftazidime (30 µg), Amikacin (30 µg), Nitrofurantoin (300 µg), Imipenem (10 µg) (Hi Media, Mumbai, India). ATCC E. coli 25922 was inoculated was used as control strain.

RESULTS

The total specimens were 704, out of which 438 (62.21%) were positive for E. coli. The age and sex distribution of E. coli is shown in table 1.

Table 1: Age and sex distribution of E. coli from isolated cases

Age group (years)	Male	Female	Total
0-10	9	15	24
10-20	9	6	15
20-50	105	141	246
>50	45	108	153
Total	168	270	438

Highest number of cases were in the age group of 20-50 years, and maximum cases were from females

The distribution of E. coli from different specialities is shown in table 2.

Table 2: Distribution of E. coli from different specialities

Specialty	Number	Percentage
General medicine	146	33.3
Gynaecology	95	21.6
General surgery	54	12.3
Pediatric	24	5.4
Urology	119	27.1

Maximum cases were from General medicine and urology departments.

The antibiogram of *E. coli* is shown in table 3

Table 3: Antibiotic Resistance Pattern of *E. coli*.

Antibiotic	Number	Percentage
Ampicillin	402	91.7
Gentamicin	246	56.1
Co-trimoxazole	234	53.4
Ofloxacin	292	66.6
Nalidixic acid	252	57.5
Piperacillin-tazobactam	95	21.6
Ceftriaxone	248	56.6
Ceftazidime	240	54.7
Amikacin	96	21.9
Nitrofurantoin	138	31.5
Imipenem	12	2.73

Highest resistance was seen with Ampicillin and least resistance to Imipenem.

DISCUSSION

Knowledge on local prevalence and antimicrobial resistance trends among urinary isolates is important in guiding clinicians appropriate for appropriate empirical treatment of UTI. *E. coli* is the most prevalent pathogen contributing to these infection, but resistance is seen nearly 70-80% of the strains to the commonly used antibiotics.^[12] In the present study *E. coli* accounted for 62.21% of all the positive cases. This is in accordance with the recent studies where the predominant organism isolated was *E. coli*.^[13-15]

In the present study highest resistance was seen with Ampicillin (91.7%), followed by Ofloxacin (66.6%) and least resistance to Imipenem (2.73%). This is in accordance with other studies where a high percentage of *E. coli* isolates were resistant to Ampicillin and Ofloxacin.^[14-16] Fluoroquinolones are considered highly effective in treatment of UTI because of concentrating ability in urine and high renal clearance.^[17] However due to wide spread use of Fluoroquinolones, there have been reports of evolving bacterial resistance to Fluoroquinolones.^[18-19]

E. coli showed varied resistance among Aminoglycosides (56% to Gentamicin and 21.9% to Amikacin) The finding differs from the other study where resistance to Gentamicin and Amikacin was high,^[19] but another study showed 40% resistance to Gentamicin and resistance to Amikacin was only 9%.^[20] Resistance to Ceftriaxone was 56.6%, Ceftazidime

was 54.7% and to Piperacillin-tazobactam was 21.6%. The findings are similar with the other studies.^[21-23] Tazobactam in combination with piperacillin has an excellent clinical efficacy in various infections.^[24] Tazobactam seems to be the most promising beta lactamase inhibitor, which has unlike clavulanic acid and sulbactam, its own antibiotic activity.^[25] Resistance to Nitrofurantoin was 31.5%. In some studies Nitrofurantoin was found to be the most effective drug for UTI.^[26-28] Antimicrobial resistance pattern varies with time which might increase or decrease.^[29] Antibiotic susceptibility studies will help in early detection of development of antibiotic resistance and preserve powerful antibiotics like Imipenem for the treatment of life threatening infections.

Limitations of the study

This was a retrospective study and sample size was small. Future studies should be prospective with large sample size.

CONCLUSION

In the present study, *E. coli* showed resistance to commonly used antibiotics. Antimicrobial susceptibility patterns vary with time and region. Prevalence and antibiotic susceptibility studies need to be conducted regularly, which will help in developing guidelines for treatment of UTI. The present study emphasizes the need for constant monitoring of susceptibility of uropathogens in different regions in order to rationalize the antibiotic use.

Conflict of interest: None

REFERENCES

- 1) Sharma S. Current understanding of Pathogenic Mechanisms in UTIs. *Ann Natl Acad Med Sci* 1997;33:31-8.
- 2) Stamm WE. Urinary tract infections and pyelonephritis, Chapter 269. In: Harrison's Principles of Internal Medicine. 16th ed. Kasper DL, Braunwald E, Fauci AS, Hauser SL, Longo DL, Jameson JL, Eds. (McGraw-Hill, New York) 2005. p. 1715-21.
- 3) Ronald AR, Pattulo MS. The natural history of urinary infection in adults. *Med Clin North Am* 1991;75:299-312.
- 4) Manges AR, Natarajan P, Solberg OD, Dietrich PS, Riley LW. The changing prevalence of drug-resistant *E coli* clonal groups in a community: Evidence for community outbreaks of urinary tract infections. *Epidemiol Infect* 2006;134:425-31.

- 5) Kahan NR, Chinitz DP, Waitman DA, Dushnitzky D, Kahan E, Shapiro M. Empiric treatment of uncomplicated urinary tract infection with fluoroquinolones in older women in Israel: Another lost treatment option? *Ann Pharmacother* 2006;40:2223-7.
- 6) Lamikanra A, Okeke IN. A study of the effect of the urban/rural divide on the incidence of antibiotic resistance in *Escherichia coli*. *Biomed Lett* 1997;55:91-7.
- 7) Belet N, İşlek I, Belet U, Sunter AT, Kuçukoduk S. Comparison of trimethoprim-sulfamethoxazole, cephadroxil and cefprozil as prophylaxis for recurrent urinary tract infections in children. *J Chemother* 2004;16:77-81.
- 8) Oladeinde BH, Omoregie R, Olley M, Anunibe JA. Urinary tract infections in a rural community of Nigeria. *N Am J Med Sci* 2011;3(2):75-7.
- 9) Karlowsky JA, Kelly LJ, Thornsberry C, Jones ME, Sahm DF. Trends in antimicrobial resistance among urinary tract infection isolates of *Escherichia coli* from female outpatients in the United States. *Antimicrob Agents Chemother* 2002;46:2540-5.
- 10) Crichton PB. Enterobacteriaceae: *Escherichia*, *Klebsiella*, *Proteus* and other genera. Chapter 20. In: Mackie and McCartney Practical Medical Microbiology. 14th ed. Collee JG, In: Fraser AG, Marmion BP, Simmons A, Eds. Churchill Livingstone, New York, 1996. p. 361-84.
- 11) Clinical Laboratories Standards Institute (CLSI). Performance of standards for antimicrobial disk susceptibility tests; approved standards. 10th ed. M02-A10. vol 29. Wayne, PA: CLSI;2009.
- 12) Kapil A. The challenge of antibiotic resistance: Need to contemplate. *Indian J Med Res* 2005;121:83-91.
- 13) Mandal J, Acharya NS, Buddhapriya D, Parija SC. Antibiotic resistance pattern among common bacterial uropathogens with a special reference to ciprofloxacin resistant *Escherichia coli*. *Indian J Med Res* 2012;136(5):842-9.
- 14) Manjunath G, Prakash R, Vamseedhar Annam KS. The changing trends in the spectrum of the antimicrobial drug resistance pattern of the uropathogens which were isolated from hospitals and community patients with urinary tract infections in Tumkur and Bangalore. *Int J Biol Med Res*. 2011; 2(2):504-07.
- 15) Dogra V, Sharma A, Mishra B, Thakur A, Loomba PS. Drug-resistant Gram-negative bacilli in urinary tract infection: A need for strict antibiotic prescription policy. *Int J Health Allied Sci* 2012;1:204-6.

- 16) Gupta N, Kundra S, Sharma A, Gautam V, Arora DR. Antimicrobial susceptibility of uropathogens in India. *J Infect Dis Antimicrob Agents* 2007;24:13-8.
- 17) Piddock LJV, Wise R. Mechanism of resistance to quinolones and clinical perspectives. *J Antimicrobial Chemother* 1989;23:427-83.
- 18) Al- Tawfiq JA. Increasing antibiotic resistance among isolates of *Escherichia coli* recovered from inpatient and outpatients in a Saudi Arabian hospital. *Infect Control Hosp Epidemiol* 2006; 27:748-53.
- 19) Mohanty S, Kapil A, Das BK, Dhawan B. Antimicrobial resistance profile of nosocomial Antibiotic Resistance Pattern of UTI uropathogens in a tertiary care hospital. *Indian J Med Sci* 2003;57:148-54.
- 20) Gupta V, Yadav A, Joshi RM. Antibiotic resistance pattern in uropathogen. *Indian J Med Microbiol* 2002;20:96-8.
- 21) Anuradha K, Sailaja VV, Umabala P, Satheesh T, Lakshmi V. Sensitivity pattern of gram negative bacilli to three β - lactam/ β - lactamase inhibitor combination using the automated API system. *Indian J Med Microbiol* 2007;25:203-8.
- 22) Eshwarappa M, Dosegowda R, Vrithmani I et al. Clinico-microbiological profile of urinary tract infection in South India. *Indian J Nephrol.* 2011;21(1):30-6.
- 23) Akram M, Shahid M, Khan AU. Etiology and antibiotic resistant patterns of community acquired urinary tract infections in JNMC Hospital Aligarh, India. *Ann Clin Microbiol Antimicrob* 2007;6:4.
- 24) Niki Y. Fundamental and clinical studies on beta- lactamase inhibitors. *Nippon Rinsho* 2001;59:771-6.
- 25) Blahova J, Hupkova M, Kremery V Sr. The effectiveness of so called potentiated penicillins (augmentin and tazobactam) in vitro. *Cas Lek cesk* 1995;134:558-61.
- 26) Sahm DF, Thornsberry C, Mayfield DC, Jones ME, Karlowsky JA. Multidrug- resistant urinary isolates of *Escherichia coli*: prevalence and patient demographics in the United States in 2000. *Antimicrob Agents Chemother* 2001;45:1402-6.
- 27) Al-Tawfiq JA. Increasing antibiotic resistance among isolates of *Escherichia coli* recovered from inpatient and outpatients in a Saudi Arabian hospital. *Infect Control Hosp Epidemiol* 2006; 27:748-53.
- 28) Sunil Kumar Biradar, Srikanth, Praveen Kumar Doddamani. Prevalence and anti-biogram of uropathogens in a tertiary care hospital. *World Journal of Pharmaceutical research* 2013;2(5):1534-43.

- 29) Dyer IE, Sankary IM, Dawson JO. Antibiotic Resistance in Bacterial Urinary Tract Infections, 1991 to 1997. WJM 1998;169(5):265-8.