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ANTIBIOGRAM OF PSEUDOMONAS AERUGINOSA ISOLATED FROM VENTILATOR ASSOCIATED PNEUMONIA IN A TERTIARY CARE HOSPITAL

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ABSTRACT

VAP is the most common nosocomial infection in the intensive care unit (ICU) with an incidence ranging from 8% to 28% in mechanically ventilated patients. The mortality and the morbidity associated with VAP can be reduced by early identification of the pathogen and administration of appropriate antimicrobial therapy. Pseudomonas aeruginosa is a major nosocomial pathogen, which causes variety of infections such as pneumonia, urinary tract infections, surgical site infections and bacteremia. It accounts for 9-10% of hospital infections. Antibiotic resistance, a global concern, is particularly pressing in developing nations, including India. Drug resistance is due to many factors such as overuse and misuse of antibiotics, incorrect choice of an antibiotic, poor compliance etc. . The aim of the present study was to know the prevalence and antibiogram of pseudomonas aeruginosa isolated from VAP. This was a retrospective study conducted at a

tertiary care hospital in South India. The data was obtained from the Microbiology department from March 2009 to March 2011. Most common gram positive organism was coagulase negative staphylococci and most common gram negative bacilli was Acinetobacter baunannii followed by Pseudomonas aeruginosa. Pseudomonas aeruginosa was resistant to most commonly used antibiotics, highest resistance was seen with Cephalosporin group followed by Fluoroquinolones (table 3). Least resistance was seen with Aztreonam followed

by Imipenem. Increasing Cephalosporin and Fluoroquinolone resistance is a matter of concern. Local bacteriological data should be collected regularly and such information should be utilized in guiding the empirical antibiotic therapy and also to develop an effective antibiotic policy.

Key words: Ventilator associated pneumonia (VAP), antibiogram, resistance, intensive care unit

INTRODUCTION

Ventilator-associated pneumonia (VAP) is defined as pneumonia occurring more than 48 hours after the initiation of endotracheal intubation and mechanical ventilation including pneumonia developing even after extubation [1]. VAP is the most common nosocomial infection in the intensive care unit (ICU) with an incidence ranging from 8% to 28% in mechanically ventilated patients [1, 2]. The mortality and the morbidity associated with VAP can be reduced by early identification of the pathogen and administration of appropriate antimicrobial therapy.

The international study of infections in intensive care unit (ICU), which was conducted in 2007, demonstrated that the patients who had longer ICU stays had higher rates of infection, especially infections due to resistant Staphylococci, Acinetobacter, Pseudomonas species, Candida species [3]. Delay in initiating appropriate antimicrobial therapy, can increase the mortality associated with VAP, and thus therapy should not be postponed for the purpose of performing diagnostic studies [4]. Empirical therapy can be initiated based on the knowledge of local prevalence and sensitivity pattern of the pathogens at a particular region.

Pseudomonas aeruginosa is a major nosocomial pathogen, which causes variety of infections such as pneumonia, urinary tract infections, surgical site infections and bacteremia. It accounts for 9-10% of hospital infections [5]. Antibiotic resistance, a global concern, is particularly pressing in developing nations, including India [6]. Drug resistance is due to many factors such as overuse and misuse of antibiotics, incorrect choice of an antibiotic, poor compliance etc. The patterns of organisms causing infections and their antibiotic resistance pattern vary widely from one country to another, as well as from one hospital to other. The aim of the present study was to know the prevalence and antibiogram of pseudomonas aeruginosa isolated from VAP.

MATERIAL AND METHODS

This was a retrospective study conducted at a tertiary care hospital in South India. The data was obtained from the Microbiology department from March 2009 to March 2011.

Inclusion criteria

The diagnosis of VAP was established using clinical pulmonary infection score (CPIS), which was evaluated on a daily basis until the patient was on ventilator support. CPIS of greater than six was used as diagnostic criteria for VAP [7].

Exclusion criteria

Patients with clinical and radiological signs suggestive of pneumonia on admission.

Collection of the Endotracheal Aspirate

Endotracheal aspirate was preferred over protected specimen brush sampling and bronchoalveolar lavage, as these techniques are more invasive and studies have shown no mortality benefit of using these over endotracheal aspirate [8-10]. Only one sample was collected from each patient and was immediately taken to the laboratory for processing.

Microbiological processing

Gram stain preparations were made from all aspirate samples within the first hour. Samples were inoculated onto 5% blood agar, MacConkey agar, which were reconstituted according to the manufacturer's specifications, and sterilised at 121°C for 15 minutes. The plates were incubated at 37°C for 18-24 hours. positive Gram stain (> 10 polymorphonuclear cells/ low power field and > 1 bacteria/ oil immersion field with or without the presence of intracellular bacteria) and quantitative endotracheal aspirate culture showing > 105 colony forming units/ ml [11-13].Based on the clinical and microbiological criteria, 59 out of 153 patients admitted were diagnosed with VAP. The organisms isolated were identified by standard microbiological techniques [14].

The isolates were tested for their antimicrobial susceptibility and the results interpreted by modified Kirby Bauer disc diffusion method, according to the guidelines of Clinical and Laboratory Standards Institute[15]. The following anti-microbials were used, Piperacillin (100 mcg), Piperacillin+tazobactum (100/10mcg), Ciprofloxacin (5mcg), Ofloxacin (5mcg), Gentamicin (10mcg), Amikacin (30mcg), Cefepime (30mcg), Cefuroxime (30 mcg), Ceftriaxone (30mcg), Ceftazidime (30mcg), Aztreonam (30mcg), Imipenem (10mcg) (Hi

Media, Mumbai). Pseudomonas aeruginosa ATCC 27853 was used as the control strain. The information was recorded and analyzed using Microsoft Excel (2010 version) and the results are explained in frequency and percentage.

RESULTS

A total of 246 patients were on Ventilator, out of which 59 patients had VAP, incidence of 23.9%. The age and sex distribution of patients is shown in table 1.

Table 1 Age and sex distribution of the patients

Age group (years)	Male	Female	Total
0-10	5	6	11
10-20	3	3	6
20-40	14	11	25
40-60	7	6	13
>60	3	1	4
Total	32	27	59

Maximum number of cases were from age group of 20-40 yrs and maximum patients were male.

Table 2 Bacterial Isolated from VAP (n=59)

Organism	Number	Percentage			
Gram positive					
Staphylococcus aureus	7	11.8			
Streptococcus pneumoniae	5	8.4			
CONS	8	13.5			
Enterococci species	2	3.3			
Gra	m negative				
Pseudomonas aeruginosa	11	18.6			
Klebsiella pneumonia	7	11.8			
Acinetobacter baunannii	12	20.3			
Haemophilus influenzae	2	3.3			
Escherichia coli	3	5.08			
Citrobacter spp	2	3.3			

CONS = Coagulase negative staphylococci

Most common gram positive organism was CONS and most common gram negative bacilli was Acinetobacter baunannii followed by Pseudomonas aeruginosa

Table 3 Antibiogram of Pseudomonas aeruginosa (Resistance pattern). (n=11)

Antibiotic	Number	Percentage	
Piperacillin	6	54.5	
Piperacillin+tazobactum	4	36.3	
Ciprofloxacin	8	72.7	
Ofloxacin	7	63.6	
Gentamicin	3	27.2	
Amikacin	2	18.1	
Cefepime	9	81.8	
Cefuroxime	9	81.8	
Ceftriaxone	10	90.9	
Ceftazidime	8	72.7	
Aztreonam	0	0	
Imipenem	1	9.09	

Maximum resistance was seen with Cephalosporin group and least with Imipenem and Aztreonam.

DISCUSSION

Delay in initiating appropriate antibiotic therapy can increase the mortality associated with VAP, and thus therapy should not be postponed for the purpose of performing diagnostic studies [16,17]. One of the reasons for increased antibiotic resistance is inappropriate selection of empirical therapy [18]. Thus the empirical therapy should be based on the local bacteriological profile, patient demographics and local hospital conditions. In the present study, incidence of VAP was 23.9%. In other studies the incidence varied between 15.5-47% [19-22].

The majority of bacterial isolates in our study were gram negative bacilli (table 2). Acinetobacter baunannii was the most common organism followed by Pseudomonas aeruginosa. The organisms isolated were similar to other studies [19-22]. Pseudomonas aeruginosa was resistant to most commonly used antibiotics, highest resistance was seen with Cephalosporin group followed by Fluoroquinolones (table 3). Least resistance was seen with Aztreonam followed by Imipenem. Recent studies also report similar resistance pattern [23,24].

Despite advances in sanitation facilities and the introduction of a wide variety of antimicrobial agents with antipseudomonal activities, Pseudomonas aeruginosa is still responsible for life threatening infections. Nosocomial infections caused by Ps. aeruginosa are difficult to treat because of both intrinsic resistance of the species and its ability to

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acquire further resistance to multiple groups of antimicrobial agents including β-lactams, aminoglycosides and fluoroquinolones [25]. P. aeruginosa is an epitome of opportunistic nosocomial pathogen and despite therapy, the mortality to nosocomial pseudomonal pneumonia is approximately 70% in immunocompromised patients. Unfortunately, it demonstrates resistance to multiple antibacterial, thereby jeopardizing the selection of appropriate treatment [26].

The present study has certain limitations, this was a retrospective study and the sample size was small. We did not include multidrug resistant strains using molecular methods. Future studies should include the above parameters.

CONCLUSION

Pseudomonas aeruginosa was resistant to most commonly used anti-pseudomonals. Increasing Cephalosporin and Fluoroquinolone resistance is a matter of concern. Local bacteriological data should be collected regularly and such information should be utilized in guiding the empirical antibiotic therapy and also to develop an effective antibiotic policy. We suggest restricted and rational use Amikacin and Piperacillin+tazobactum in our hospital setting to prevent rapid emergence of resistance.

Conflict of interest: None

REFERENCES

- 1) Chastre J, Fagon JY. Ventilator-associated pneumonia. Am J Respir Crit Care Med 2002;165: 867-903.
- 2) Safdar N, Crnich CJ, Maki DG. The pathogenesis of ventilator associated pneumonia: Its relevance to developing effective strategies for prevention. Respir Care 2005;50:725-39
- 3) Radji M, Fauziah S, Aribinuko N. Antibiotic sensitivity pattern of bacterial pathogens in the intensive care unit of Fatmawati Hospital, Indonesia. Asian Pacific Journal of Tropical Biomedicine 2011;1:39-42.
- 4) Iregui M, Ward S, Sherman G, Fraser VJ, Kollef MH. Clinical importance of delays in the initiation of appropriate antibiotic treatment for ventilator-associated pneumonia. Chest 2002;122:262-8.
- 5) Hancock RE, Speert DP. Antibiotics for Pseudomonas and related infections. In: Dodge JA, Brock DJ, Widdicombe JH, editors. Cystic fibrosis-current topics. Vol. 3. United States: John Wiley and Sons Ltd; 1996. p. 245-66.

- 6) Ganguly NK, Arora NK, Chandy SJ, Fairoze MN, Gill JP, Gupta U, et al. Rationalizing Antibiotic use to limit antibiotic resistance in India. Indian J Med Res 2011;134:281-94.
- 7) Pugin J, Auckenthaler R, Mili N, Janssens JP, Lew PD, Suter PM. Diagnosis of ventilator-associated pneumonia by bacteriologic analysis of bronchoscopic and nonbronchoscopic "blind" bronchoalveolar lavage fluid. Am Rev Respir Dis 1991;143:1121-9
- 8) Sanchez-Nieto JM, Torres A, Garcia-Cordoba F, El-Ebiary M, Carrillo A, Ruiz J, et al. Impact of invasive and noninvasive quantitative culture sampling on outcome of ventilator-associated pneumonia: A pilot study. Am J Respir Crit Care Med 1998;157:371-6.
- 9) Ruiz M, Torres A, Ewig S, Marcos MA, Alcon A, Lledo R, et al. Noninvasive versus invasive microbial investigation in ventilator associated pneumonia: Evaluation of outcome. Am J Respir Crit Care Med 2000;162:119-25.
- 10) Sole Violan J, Fernandez JA, Benitez AB, Cardenosa Cendrero JA, Rodriguez de Castro F. Impact of quantitative invasive diagnostic techniques in the management and outcome of mechanically ventilated patients with suspected pneumonia. Crit Care Med 2000;28:2737-41
- 11) Porzecanski I, Bowton DL. Diagnosis and treatment of ventilator associated pneumonia. Chest 2006; 130: 597-604.
- 12) Wu CL, Yang DI, Wang NY, Kuo HT, Chen PZ. Quantitative culture of endotracheal aspirates in the diagnosis of ventilator-associated pneumonia in patients with treatment failure. Chest 2002; 122: 662-668.
- 13) Koenig SM, Truwit JD. Ventilator-associated pneumonia:diagnosis, treatment and prevention. Clin Microbiol Rev 2006; 19: 637-657.
- 14) Collee JG, Marr W. Culture of Bacteria. In: Collee JG, Fraser AG, Marmion BP, Simmons A, editors. Mackie and McCartney Practical Medical Microbiology. 14th ed. New York: Churchill Livingstone; 1996. pp. 113-29.
- 15) Clinical Laboratories Standards Institute (CLSI). Performance of standards for antimicrobial disk susceptibility tests; approved standards. 10th ed. Wayne, PA: CLSI;2009. Vol 29. M02-A10.
- 16) Iregui M, Ward S, Sherman G, Fraser VJ, Kollef MH. Clinical importance of delays in the initiation of appropriate antibiotic treatment for ventilator-associated pneumonia. Chest 2002;122:262-8.

- 17) Alvarez-Lerma F; ICU-acquired Pneumonia Study Group. Modification of empiric antibiotic treatment in patients with pneumonia acquired in the intensive care unit. Intensive Care Med 1996;22:387-94.
- 18) Kollef MH. Inadequate antimicrobial treatment: An important determinant of outcome for hospitalized patients. Clin Infect Dis 2000;31:S131-8.
- 19) Torres A, Puig de la Bellacasa J, Xaubet A, Gonzalez J, Rodriguez-Roisin R, Jimenez de Anta MT, et al. Diagnostic value of quantitative cultures of bronchoalveolar lavage and telescoping plugged catheters in mechanically ventilated patients with bacterial pneumonia. Am Rev Respir Dis 1989;140:306-10.
- 20) Kollef MH. Ventilator-associated pneumonia: A multivariate analysis. JAMA 1993;270:1965-70.
- 21) Fagon JY, Chastre J, Domart Y, Trouillet JL, Pierre J, Darne C, et al. Nosocomial pneumonia in patients receiving continuous mechanical ventilation: Prospective analysis of 52 episodes with use of protected specimen brush and quantitative culture techniques. Am Rev Respir Dis 1989;139:884.
- 22) Panwar R, Vidya SN, Alka KD. Incidence, clinical outcome and risk stratification of ventilator-associated pneumonia: A prospective cohort study. Indian J Crit Care Med 2005;9:211-6.
- 23) Goel V, Hogade SA, Karadesai SG. Ventilator associated pneumonia in a medical intensive care unit: Microbial aetiology, susceptibility patterns of isolated microorganisms and outcome. Indian J Anaesth 2012;56:558-62.
- 24) Gupta A, Agrawal A, Mehrotra S, Singh A, Malik S, Khanna A. Incidence, risk stratification, antibiogram of pathogens isolated and clinical outcome of ventilator associated pneumonia. Indian J Crit Care Med 2011;15:96-101.
- 25) Strateva T, Yordanov D. Pseudomonas aeruginosa a phenomenon of bacterial resistance. J of Medical Microbiology 2009;58:1133-48.
- 26) Javiya VA, Ghatak SB, Patel KR, Patel JA. Antibiotic susceptibility patterns of Pseudomonas aeruginosa at a tertiary care hospital in Gujarat, India. Indian J Pharmacol 2008;40(5):230-4.