

# WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 5.045

Volume 3, Issue 9, 1282-1296.

Research Article

ISSN 2277-7105

# A NOOTROPIC EFFECT OF BENINCASA HISPIDA ON ACH AND CHAT ACTIVITY IN COLCHICINE INDUCED EXPERIMENTAL RAT MODEL OF ALZHEIMER'S DISEASE: POSSIBLE INVOLVEMENT OF ANTIOXIDANTS

# <sup>1</sup>Chandan Roy\*, <sup>2</sup>Debasis Mazumdar

<sup>1</sup>Dept. of Physiology, Berhampore Girls' College, University of Kalyani, Berhampore, West Bengal, India.

<sup>2</sup>Dept. of Agricultural Statistics, Bidhan Chandra Krishi Viswavidyalaya (State Agriculture University), Mohanpur, Nadia, West Bengal, India.

Article Received on 25 August 2014,

Revised on 18 Sept 2014, Accepted on 14 Oct 2014

\*Correspondence for Author

Dr. Chandan Roy

Dept. of Physiology, Berhampore Girls' College, University of Kalyani, Berhampore, West Bengal, India.

iamchandan@rediffmail.com

#### **ABSTRACT**

The fruit *Benincasa hispida* (BH) is an important ingredient of 'Kusmanda lehyam' (Ayurvedic medicine), which is widely used, in nervous disorders. The present study has been designed to evaluate the cognition facilitating effect of BH pulp extract in colchicine induced experimental rat model of AD and to investigate the role of central cholinergic system in the nootropic effect of BH pulp extract with the possible involvement of antioxidant enzymes. The behavior study, Acetylcholine concentration, cholineacetyl transferase activity, antioxidant level such as, superoxide dismutase (SOD), catalase (CAT), reduced glutathione (GSH) and lipid peroxidation (LPO) level were studied in different parts of the brain such as frontal cortex (FC) and hippocampus (HPC) in colchicine induced experimental

Alzheimer rat model before and after treatment with BH. BH (400 mg/kg p.o.) induced statistically significant reversal of colchicine induced cognitive deficits. BH (400 mg/kg p.o.) markedly induced frontal cortical and hippocampal concentrations of Ach and ChAt activity, the effects being statistically significant on days 7, 14 and 21 respectively. Moreover, BH significantly increased SOD, CAT, GSH activities and significantly decreased LPO level on day 7, 14 and 21 respectively. The aqueous pulp extract of BH (400 mg/kg body weight) containing vit- A, C, E results significant protection in the level of antioxidant status in frontal cortex and hippocampas after a certain period of co administration on colchicine

induced oxidative stress without causing any general and metabolic toxicity and possibly thereby induced frontal cortical and hippocampal concentration of Ach and ChAT activity.

**KEY WORDS:** *Benincasa hispida*, Colchicine, Alzheimer's disease, Acetylcholine, Cholineacetyltransferase, Antioxidant.

# INTRODUCTION

The fruit of *Benincasa hispida* (Thunb) Cogn, commonly called as ash guard, belonging to cucurbitaceae family is employed as a main ingredient in kusmanda lehyam, in Ayurvedic system of medicine. The fruit *Benincasa hispida* (BH) is an important ingredient of 'Kusmanda lehyam' (Ayurvedic medicine), which is widely used, in nervous disorders. The major constituents of this fruits are triterpenoids, flavanoids, glycosides, saccharides, carotenes, vitamins, β sitosterin, and uronic acid (Nadkarni, 1976; Wollen et al., 1991; Yashizumi et al., 1998). Some of the important isolated compounds of BH reported were triterpenes, sterols and glycosides (Yashizumi et al., 1998) and volatile oils.

Alzheimer's disease (AD) is a complete neurodegenerative disorders characterized by the loss of learning, memory and other cognitive functions. AD is characterized by degeneration of neurons, especially pyramidal neurons in the hippocampus, entorhinal cortex, and neocortical areas and cholinergic neurons in the median forebrain.

Colchicine, as a microtubule-disrupting agent (James & Dennis, 1981) produces marked destruction of hippocampal granule cells, mossy fibers and septohippocampal pathways (SHC, a cholinergic link between medial septum and vertical limb of diagonal band). In induces neurofibrillary degeneration by binding to tubulin, the principal structural protein of microtubule (McClure, 1972; Wilson & Fried-Kin, 1966; Walsh et al., 1986). This event is associated with loss of cholinergic neurons and decrease in acetylcholine transferase, thereby resulting in impairment of learning and memory (Kevin et al., 1989; Dwaine & Thomas, 1990). Oxidative stress due to increase in free radical generation of impaired endogenous antioxidant mechanism is an important factor that has been implicated in neuronal damage and in AD, and cognitive defects seen in elderly (Pratico & Delanty, 2000; Cantuli et al., 2000). A number of in vitro studies have shown that antioxidants, both endogenous and dietary, can protect nervous tissue from damage by oxidative stress. Vitamin C has been described to be a major hydrophilic antioxidant in human plasma (Frei et al., 1989), CSF (Spector & Lorenzo, 1973; Lonnrot et al., 1996) and the central nervous system (Rice, 2000).

The same study showed that vitamin E prevented neuronal damage from reactive nitrogen species (Tagami et. al., 1998). Both vitamin E and  $\beta$  carotene were found to protect rat neurons against oxidative stress from exposure to ethanol (Mitchel et al., 1999). BH is rich in vitamin C, E and beta- carotene (Roy et al., 2007).

Thus the present study was undertaken to determine the cognition facilitating effect of BH pulp extract in colchicine induced experimental rat model of AD and to investigate the role of central cholinergic system in the nootropic effect of BH pulp extract with the possible involvement of antioxidant enzymes.

#### MATERIALS AND METHODS

# **Subjects**

Male Holtzman strain adult albino rats approximately 120 days old and weighing 250-300gm were used in the following studies. The animals were individually housed and maintained under standard laboratory conditions with natural dark and light cycle (approximately 12-h light/10-h dark cycle) and room temperature (27±1°C) and constant humidity (60%) in accordance with 'Institutional Ethical Committee' rules and regulations. Food and water were freely available except during testing. Drinking water was supplied ad libitum. Five days prior to behavioral training, animals were reduced to 85% of their free feeding weight by limiting their daily ration of food. Food deprivation was maintained throughout testing except for 3 days immediately prior to, and following surgery. Body weights of the rats were recorded every day and maintained in the laboratory throughout the experimental period. The behavioral procedure was carried out between 12:00 and 14:00 h.

# Collection and Preparation of Water Extract from the Pulp of BH

The fruit of BH were purchased from the local market and the identity of the plant was authenticated by the Botanical Survey of India, Howrah and kept in S.N.Pradhan Centre for Neurosciences, University of Calcutta. The pulp of BH fruit was used throughout the experimental study. The fruit of BH were cut into pieces, sun-dried under shady region because sun-dry may results in loss of chemicals and ground with the help of an electrical grinder to get a free flowing powder. This powder was subjected to extraction with water (1:3) by simple decoction method at room temperature for 48 hours. The extract obtained was filtered through Whatman filter paper and vaccum dried at  $40^{\circ} - 50^{\circ}$  C to get a dry powder, which was dissolved in double distilled water for final use (Roy et al., 2007).

#### **Treatment**

The control animal was treated with artificial cerebrospinal fluid or ACSF. The BH pulp extract was given orally through orogastric cannula at the standard dose of 400 mg/kg p.o. for seven, fourteen and twenty one consecutive days respectively (between 10:00 and 11:00 hrs). The dose was standardized in the laboratory.

The animals were sacrificed by cervical dislocation and the different parts of the brain like Frontal cortex (FC), Hippocampus (HPC), Cerebral cortex (CC), Cerebellum (CB), Caudate nucleus (CN), Pons & Medulla (PM) and Midbrain (MB), were isolated for biochemical estimation after seven, fourteen and twenty one days respectively.

# **Grouping of Animal:** The animals were divided into four groups:

- 1. Control (ACSF) rats
- 2. Colchicine induced Alzheimer's rat model
- 3. Control rats treated with BH pulp extract
- 4. Colchicine induced Alzheimer's rat model treated with BH pulp extract.

# **Behavioural Study**

The apparatus used consisted of a shuttle box with two identical compartments, separated by a hurdle. During training, each rat was placed in one compartment and after 5 sec a buzzer, situated in the ceiling of the shuttle box, was sounded (2.8 kHz, 70 dB) (conditioned stimulus, CS) for 3 sec, followed by electric shock (1.5 mA, 2 sec) (unconditioned stimulus, UCS) through the grid floor. If the rat crossed to the unelectrified safe compartment during presentation of CS, an avoidance response was recorded, otherwise UCS was applied. Each rat was given 20 trials for 5 days, with an intertrial interval of 30 sec, before lesioning, until it reached the criterion of 100% active avoidance response. Rats not reaching this criterion were discarded from the study (Jaiswal & Bhattacharya, 1992). Retention of the acquired active avoidance response, in the different treatment groups, was assessed on days 7, 14 and 21, following lesioning with colchicine or ibotenic acid, by noting the number of trials required to criterion of 100% active avoidance response.

# Preparation of Experimental Alzheimer's Rat Model by Colchicine

Prior to surgery, all the animals were subjected to overnight fasting though drinking water was not withdrawn. During procedures, the animals were anaesthetized with sodium pentobarbital (50mg/kg b.w.) and re-strained in a stereotaxic apparatus (INCO, INDIA Ltd)

equipped with a custom-made ear bar, which prevents the damage of the tympanic membrane. Head was fixed in such a position that lambda and bregma sutures were in the same horizontal plane by introducing the incisor bar properly attached to the mouth. For aseptic surgery, absolute alcohol or rectified spirit was applied. The scalp was incised and retracted. An incision was made in the scalp and two holes were drilled in the skull for placement of the injection cannula into the lateral cerebral ventricles. The stereotaxic coordinates for intracerebroventricular injection were: 0.8 mm posterior to bregma, 1.8 mm lateral to the sagittal suture and 3.6 mm below the cortical surface (Veerendrakumar & Gupta, 2002). Subjects were infused through a 10 μl Hamilton syringe with 15 μgm of colchicine (Wako chemicals) in 5 μl of artificial cerebrospinal fluid (ACSF; in Mm: 147 Nacl, 2.9 Kcl, 1.6 Mgcl<sub>2</sub>, 2.2 Dextrose and 1.7 Cacl<sub>2</sub>) in lateral cerebral ventricle bilaterally. A total volume of 10 μl was delivered to the injection site and the injection cannula was left in place for 2-3 min following infusion.

*Postoperative care:* After surgery, all aseptic measures and care were taken for feeding until recovery from surgical stress. Penicillin was given post operatively to all animals for 3 consecutive days by intramuscular route. After 3 days of surgery, experiment was started and continued routinely until sacrificed. Similar procedure was repeated thrice, each at an interval of two days.

# **Biochemical Estimation**

# **Tissue Preparation**

Rats were sacrificed by cervical dislocation on day 7, 14 and 21 immediately after behavior study. The Frontal cortex (FC) and Hippocampus (HPC) were dissected out. The tissues were weighed and homogenized in ice-cold phosphate buffer and prepared for biochemical estimation.

# **Estimation of Ach and ChAT Activity**

Rats of the colchicine group were killed by decapitation at the predetermined time intervals and the frontal cortex and hippocampus were dissected out (Glowinski & Iversen, 1966). The tissues were homogenised in 10 volumes (w/v) of ice-cold Tris-HCl buffer (pH 7.6) and divided into aliquots for estimation of acetylcholine (Ach) levels by a fluorimetric technique (Speeg, 1974), choline acetyltransferase (ChAT) activity by a radiometric method (Haba et al., 1988).

# Estimation of SOD, CAT, GSH activity and LPO level

Catalase (CAT) activity was estimated by the method of Cohen et al., (1970); Roy et al., (2007), Superoxide Dismutase (SOD) was estimated by the method of Mishra & Fridovich (1972); Roy et al., (2007), Reduced glutathione (GSH) level was measured according to the method of Ellman (1959) and Lipid Peroxidation (LPO) was estimated by the method of Bhattacharya et al. (2001); Roy et al., (2007).

# **Statistical Analysis**

The data were expressed as MEAN  $\pm$  S.E.M. and were analyzed statistically using one way analysis of variance (one way ANOVA) followed by multiple comparison 't' test. In addition to this, two-tailed Student't' test was performed to determine the level of significance between the means. Difference below the probability level 0.05 was considered statistically significant.

# **RESULTS**

# **Results of Behavioural Parameter**

Colchicine, injected i.c.v., induced marked deficits of the learned active avoidance task, as compared to their ACSF treated counterparts, after 7, 14 and 21 days following administration of the neurotoxins. The retention deficit was evident by day 7 and increased progressively on days 14 and 21. BH (400 mg/kg p.o.) induced dose-related statistically significant reversal of colchicine induced cognitive deficits, when assessed on day 7, remained statistically non-significant (Table 1).

# **Results of Parameters of Cholinergic System**

Colchicine, administered i.c.v., markedly reduced frontal cortical and hippocampal concentrations of Ach and ChAT activity, as compared to the ACSF administration control group. The effects were discernible by day 7, and increased progressively, thereafter, on days 14 and 21. BH (400 mg/kg p.o.) tended to reverse the deleterious effects of colchicine on all these biochemical parameters, the effects being statistically significant on days 14 and 21, but not on day 7 (Tables 2, 3 and 4).

# **Results of Parameters of Oxidative Stress**

Colchicine, administered i.c.v., markedly reduced SOD, CAT and GSH activity and markedly increased LPO level in different mentioned brain parts respectively, as compared to the ACSF administration control group. The effects were discernible by day 7, and increased

progressively, thereafter, on days 14 and 21. BH (400 mg/kg p.o) tended to reverse the deleterious effects of colchicine on all these biochemical parameters, the effects being statistically significant on days 7, 14 and 21 respectively (Table 5, 6. 7 and 8).

Table 1: Effect of BH on the retention of an active avoidance learning acquisition in cognitive deficits induced by colchicine (15  $\mu$ g, i.c.v.) in rats (values are Mean  $\pm$  SEM)

| Treatments    |   |                        | Number of trials required to achieve 100% avoidance response |                    |  |  |  |  |
|---------------|---|------------------------|--|--------------------|--|--|--|--|
| (mg/kg)       | N | Colchicine group       |  |                    |  |  |  |  |
|               |   | Day 7                  | <b>Day 14</b>  | Day 21             |  |  |  |  |
| ACSF          | 6 | $4.9 \pm 0.02$         | 3.3±0.02   | 2.7±0.04           |  |  |  |  |
| ВН            | 6 | 2.9±0.03*              | $2.1\pm0.02^*$   | 1.3±0.04*          |  |  |  |  |
| COLCHICINE    | 6 | 7.7±0.03**             | 8.2±0.03**   | 9.3±0.04**         |  |  |  |  |
| BH+COLCHICINE | 6 | 5.42±0.03 <sup>#</sup> | $3.64\pm0.02^{\#}$   | $2.24\pm0.04^{\#}$ |  |  |  |  |

Values are mean  $\pm$  SEM, n = 6; \*p < 0.001, \*\*p < 0.001 when compared with ACSF group. \*p < 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student't' – test.

Table 2: Effect of BH on acetylcholine concentrations of frontal cortex and hippocampus in colchicine (15  $\mu$ g i.c.v.) administered rats (values are Mean  $\pm$  SEM)

| Treatments (mg/kg) |   | Acetylcholine concentrations (nmol/g) |                         |                         |                         |                         |                         |  |
|--------------------|---|---------------------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|--|
|                    | N | Frontal cortex                        |                         |                         | Hippocampus             |                         |                         |  |
|                    |   | Day 7                                 | Day 14                  | Day 21                  | Day 7                   | Day 14                  | <b>Day 21</b>           |  |
| ACSF               | 6 | 24.64±0.02                            | 26.30±0.03              | 23.52±0.04              | 30.54±0.03              | 28.42±0.02              | 31.62±0.04              |  |
| ВН                 | 6 | 28.22±0.04*                           | 34.66±0.06*             | 30.02±0.04*             | 33.84±0.03*             | 35.24±0.08*             | 38.22±0.04*             |  |
| COLCHICINE         | 6 | 18.22±0.02**                          | 15.44±0.04**            | 11.52±0.03**            | 23.42±0.04**            | 19.14±0.09**            | 14.44±0.06**            |  |
| BH+COLCHICINE      | 6 | 23.56±0.04 <sup>#</sup>               | 30.66±0.04 <sup>#</sup> | 24.86±0.05 <sup>#</sup> | 28.96±0.04 <sup>#</sup> | 33.12±0.03 <sup>#</sup> | 31.48±0.04 <sup>#</sup> |  |

Values are mean  $\pm$  SEM, n = 6; \*p < 0.001, \*\*p < 0.001 when compared with ACSF group. \*p

< 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student't' – test.

Table 3: Effect of BH on choline acetyltransferase activity of frontal cortex and hippocampus in colchicine (15  $\mu$ g, i.c.v.) administrated rats (Values are Mean  $\pm$  SEM)

| Treatments    | n | Choline acetyltransferase activity (nmol/mg protein/h) |                         |                         |                         |                         |                         |  |
|---------------|---|--|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|--|
| (mg/kg)       |   |  | <b>Frontal cortex</b>   |                         |                         | Hippocampus             |                         |  |
|               |   | Day 7  | <b>Day 14</b>           | Day 21                  | Day 7                   | Day 14                  | Day 21                  |  |
| ACSF          | 6 | 21.22±0.04   | 20.44±0.02              | 19.96±0.07              | 19.22±0.04              | 18.54±0.06              | 19.44±0.04              |  |
| ВН            | 6 | 21.44±0.02*  | 24.88±0.03*             | 22.56±0.02*             | 22.42±0.05*             | 22.54±0.04*             | 23.46±0.01*             |  |
| COLCHICINE    | 6 | 16.44±0.04**   | 14.24±0.04**            | 11.96±0.04**            | 14.66±0.04**            | 12.22±0.02**            | $9.88 \pm 0.08^{**}$    |  |
| BH+COLCHICINE | 6 | 18.46±0.02 <sup>#</sup>                                | 21.34±0.06 <sup>#</sup> | 23.34±0.02 <sup>#</sup> | 17.62±0.02 <sup>#</sup> | 19.66±0.04 <sup>#</sup> | 16.68±0.02 <sup>#</sup> |  |

Values are mean  $\pm$  SEM, n = 6; \*p < 0.001, \*\*p < 0.001 when compared with ACSF group. \*p < 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student 't' – test.

Table 4: Effect of BH on muscarinic cholinergic receptors in frontal cortex and hippocampus in colchicine (15  $\mu$ g, i.c.v.) administered rats (Values are Mean  $\pm$  SEM)

| Treatments    | N | (3H) QNB binding (pmoles/mg protein) |                        |                        |                        |                        |                        |  |  |
|---------------|---|--------------------------------------|------------------------|------------------------|------------------------|------------------------|------------------------|--|--|
| (mg/kg)       |   | ]                                    | Frontal cortex         |                        |                        | Hippocampus            |                        |  |  |
|               |   | Day 7                                | Day 14                 | Day 21                 | Day 7                  | Day 14                 | Day 21                 |  |  |
| ACSF          | 6 | 1.54±0.01                            | 1.71±0.02              | 1.56±0.02              | 1.37±0.03              | 1.44±0.02              | 1.42±0.02              |  |  |
| ВН            | 6 | 1.56±0.02*                           | 1.72±0.02*             | 1.96±0.01*             | 1.59±0.01*             | 1.66±0.01*             | 1.86±0.01*             |  |  |
| COLCHICINE    | 6 | 0.83±0.02**                          | 0.56±0.02**            | 0.46±0.01**            | 0.71±0.02**            | 0.53±0.02**            | $0.42\pm0.02^{**}$     |  |  |
| BH+COLCHICINE | 6 | 1.26±0.01 <sup>#</sup>               | 1.33±0.01 <sup>#</sup> | 1.48±0.02 <sup>#</sup> | 1.08±0.04 <sup>#</sup> | 1.41±0.02 <sup>#</sup> | 1.58±0.01 <sup>#</sup> |  |  |

Values are mean  $\pm$  SEM, n = 6; \*p < 0.001, \*\*p < 0.001 when compared with ACSF group. \*p < 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student 't' – test.

Table 5: Effect of BH on SOD activity in frontal cortex and hippocampus in colchicine (15  $\mu$ g, i.c.v.) administered rats (Values are Mean  $\pm$  SEM)

| Treatments (mg/kg) |   |                         | SOD                     | (% inhibition           | unit)                   |                         |                         |  |
|--------------------|---|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|--|
|                    | N | Frontal cortex          |                         |                         | Hippocampus             |                         |                         |  |
|                    |   | Day 7                   | Day 14                  | <b>Day 21</b>           | Day 7                   | <b>Day 14</b>           | <b>Day 21</b>           |  |
| ACSF               | 6 | 13.30±0.02              | 11.28±0.04              | 11.31±0.04              | 11.12±0.03              | 12.47±0.07              | 12.42±0.04              |  |
| ВН                 | 6 | 10.31±0.03*             | 9.82±0.02 *             | 9.96±0.04*              | $9.79\pm0.05^*$         | 10.27±0.04*             | 10.81±0.02*             |  |
| COLCHICINE         | 6 | 20.66±0.06**            | 19.11±0.05**            | 19.60±0.06**            | 18.58±0.06**            | 20.85±0.03**            | 21.40±0.02**            |  |
| BH+COLCHICINE      | 6 | 15.38±0.03 <sup>#</sup> | 14.19±0.06 <sup>#</sup> | 14.31±0.07 <sup>#</sup> | 14.46±0.05 <sup>#</sup> | 14.43±0.02 <sup>#</sup> | 15.43±0.02 <sup>#</sup> |  |

Values are mean  $\pm$  SEM, n = 6; \*p < 0.001, \*\*p < 0.001 when compared with ACSF group. \*p < 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student't' – test.

Table 6: Effect of BH on CAT activity in frontal cortex and hippocampus in colchicine (15  $\mu$ g, i.c.v.) administered rats (Values are Mean  $\pm$  SEM)

| Treatments (mg/kg) |   | CAT (% inhibition unit) |               |                       |              |                         |                         |  |
|--------------------|---|-------------------------|---------------|-----------------------|--------------|-------------------------|-------------------------|--|
|                    | N | Frontal cortex          |               |                       | Hippocampus  |                         |                         |  |
|                    |   | Day 7                   | <b>Day 14</b> | Day 21                | Day 7        | Day 14                  | <b>Day 21</b>           |  |
| ACSF               | 6 | 13.98±0.03              | 12.30±0.09    | 12.54±0.04            | 13.19±0.04   | 12.16±0.04              | 12.46±0.02              |  |
| ВН                 | 6 | $12.27\pm0.04^*$        | 10.81±0.04*   | 10.66±0.03*           | 11.79±0.05*  | 10.55±0.03*             | 10.22±0.02*             |  |
| COLCHICINE         | 6 | 21.65±0.03**            | 20.35±0.04**  | 19.69±0.05**          | 20.77±0.06** | 20.13±0.04**            | 22.36±0.04**            |  |
| BH+COLCHICINE      | 6 | $15.72\pm0.04^{\#}$     | 14.23± 0.03#  | $14.82 \pm 0.06^{\#}$ | 14.40± 0.06# | 13.97±0.07 <sup>#</sup> | 15.72±0.06 <sup>#</sup> |  |

Values are mean  $\pm$  SEM, n = 6; \*p < 0.001, \*\*p < 0.001 when compared with ACSF group.

Table 7: Effect of BH on LPO level in frontal cortex and hippocampus in colchicine (15  $\mu$ g, i.c.v.) administered rats (Values are Mean  $\pm$  SEM)

| Treatments    | N | LPO (nmol of TBARS / gm mol of tissue) |                        |                        |                        |                        |                        |  |
|---------------|---|--|------------------------|------------------------|------------------------|------------------------|------------------------|--|
| (mg/kg)       |   | ]                                      | Frontal cortex         |                        |                        | Hippocampus            | 5                      |  |
|               |   | Day 7                                  | Day 14                 | Day 21                 | Day 7                  | Day 14                 | Day 21                 |  |
| ACSF          | 6 | 4.03±0.02                              | 3.99±0.04              | 3.46±0.04              | $3.89\pm0.02$          | 3.72±0.04              | 3.98±0.02              |  |
| ВН            | 6 | 2.93±0.01*                             |                        | 3.19±0.02*             | 2.89±0.03*             | 3.35±0.02*             | 2.89±0.02*             |  |
| COLCHICINE    | 6 | 8.52±0.02**                            | 7.46±0.04**            | 7.59±0.03**            | 7.70±0.04**            | 7.19±0.02**            | 8.52±0.04**            |  |
| BH+COLCHICINE | 6 | 5.29±0.03 <sup>#</sup>                 | 4.63±0.05 <sup>#</sup> | 4.20±0.04 <sup>#</sup> | 4.68±0.03 <sup>#</sup> | 4.60±0.04 <sup>#</sup> | 5.02±0.02 <sup>#</sup> |  |

Values are mean  $\pm$  SEM, n = 6; \*p < 0.001, \*\*p < 0.001 when compared with ACSF group. \*p < 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student't' – test.

Table 8: Effect of BH on GSH level in frontal cortex and hippocampus in colchicine (15  $\mu$ g, i.c.v.) administered rats (Values are Mean  $\pm$  SEM)

| Treatments    | N | Reduced glutathione (μg/g of tissue) |                         |                         |                         |                         |                         |
|---------------|---|--------------------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|
| (mg/kg)       |   | Frontal cortex                       |                         |                         | Hippocampus             |                         |                         |
|               |   | Day 7                                | Day 14                  | Day 21                  | Day 7                   | <b>Day 14</b>           | <b>Day 21</b>           |
| ACSF          | 6 | 30.54±0.06                           | 29.62±0.11              | 29.61±0.04              | 23.9±0.06               | 26.65±0.04              | 29.42±0.02              |
| ВН            | 6 | 32.21±0.07*                          | 31.2±0.08 *             | 28.08±0.08*             | 26.71±0.04*             | 27.68±0.02*             | 32.26±0.04*             |
| COLCHICINE    | 6 | 19.96±0.05**                         | 18.78±0.04**            | 15.26±0.08**            | 16.15±0.06**            | 17.17±0.04**            | 15.56±0.02**            |
| BH+COLCHICINE | 6 | 26.8±0.04 <sup>#</sup>               | 26.81±0.03 <sup>#</sup> | 24.87±0.07 <sup>#</sup> | 25.33±0.07 <sup>#</sup> | 25.17±0.03 <sup>#</sup> | 24.44±0.02 <sup>#</sup> |

Values are mean  $\pm$  SEM, n = 6; \*\*p < 0.001 when compared with ACSF group. \*p < 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student't' – test.

<sup>\*</sup>p < 0.001 when compared with colchicine treated group. Data were analyzed statistically using one-way ANOVA Test followed by student't' – test.

# **DISCUSSION**

The present study evaluates the nootropic effect of BH on cognition facilitating effect and central cholinergic system in colchicine induced experimental rat model of AD with the possible involvement of antioxidant enzymes. It is evident from the results of the present investigation that intracerebroventricular administration of colchicine induced marked deficits of the learned active avoidance task, as compared to their ACSF treated counterparts after 7, 14 and 21 days in colchicines treated experimental AD group. But, treatment with aqueous pulp extract of BH significantly decreased marked deficits of the learned active avoidance task in BH cotreated colchicines treated experimental AD group in comparision with only colchicines treated experimental AD group. These findings can be explained by alterations of the parameters of oxidative stress namely lipid peroxidation level (LPO), SOD, CAT and GSH activity along with alterations of AchE and ChAT activity respectively. Thus, the marked deficit in the retention of the learned active avoidance task, in rats induced by colchicine, noted in this study, is consonant with earlier report (Emerich & Walsh, 1990). Treatment with aqueous pulp extract of BH was able to reverse cognitive deficits induced by colchicine, the effects being evident after 2 weeks of treatment. The reversal of cognitive deficits, induced by colchicine, was accompanied by attenuation of its cholinotoxic effects, indicating that the drug was capable of promoting cholinergic recovery. The findings support clinical (Koti, 1991) and experimental (Verma & Kulkarni, 1991) observations that BH can improve memory in states of cognitive deficits. From our present investigation, i.c.v. administration of colchicines markedly reduced frontal cortical and hippocampal concentrations of Ach, ChAt activity, as compared to the ACSF administration control group. The effects were discernible by day 7, and increased progressively, thereafter, on days 14 and 21. BH (400 mg/kg p.o.) tended to reverse the deleterious effects of colchicine on all these biochemical parameters, the effects being statistically significant on days 14 and 21, but not on day 7. It was reported that the i.c.v. injection of colchicines significantly decreased the number of cholinergic neurons in the medial septum /vertical limb of the diagonal band, which project to the hippocampus and synapse on granule cells, pyramidal cells and interneurons. Therefore, it was expected that the intracerebroventricularly administered colchicines would preferentially act on the cholinergic neurons (Dwaine & Thomas, 1991). Nootropic agents, like piracetam, which have been shown to facilitate central cholinergic mechanisms (Chouinard et al., 1983; Moos et al., 1988), are known to improve memory only in the presence of cognitive deficits.

Intracerebroventricular infusion of colchicine causes it to bind with tubulin which is the structural and functional protein of microtubule and thereby generates more and more reactive oxygen species (ROS) leading to neurodegeneration and ultimately produces a condition akin to AD or produces experimental AD model which is histopathologically characterized by the extracellular deposition of senile plaques and the intracellular deposition of neurofibrillary tangles. Free radicals play a crucial role in the pathogenesis of AD. Lipid peroxidation can be used as an index for measuring the damage that occurs in membranes of tissue as a result of free radical generation (Dianzani, 1985; Husain & Somani, 1997). In our present study, ICV infusion of colchicine, it significantly increased the LPO level. The results of significant elevation of LPO level in colchicine treated experimental Alzheimer's group is possibly due to the generation of free radicals via auto-oxidation or through metal ion or superoxide catalyzed oxidation process. This is possibly due to the generation of free radicals via autooxdidation or through metal ion or superoxide catalyzed oxidation process. In the present experiment, BH significantly decreased LPO level in a dose dependent manner compared to other groups. So, from the result of LPO levels it may be concluded that the protection by BH may be due to vitamin E and beta carotene which is present in BH pulp extract.

Endogenous antioxidant status in colchicines induced experimental Alzheimer's rat model was evaluated here by noting the activities of CAT, SOD and GSH as these are the important biomarkers for scavenging free radicals (Venkateswaran & Pari, 2003). Colchicine induced oxidative stress is further supported here by the study of antioxidant scavenger enzyme activities.

CAT that protects the tissues from highly reactive hydroxyl radical catalyzes the reduction of hydrogen peroxide. The primary role of CAT is to scavenge H<sub>2</sub>O<sub>2</sub> that has been generated by free radicals or by SOD in removal of superoxide anions and to convert it to water (Ribiere et al., 1992). The destruction of superoxide radicals is catalyzed by SOD, is an important defense system against oxidative damage. From our experimental results of the aforesaid antioxidant enzyme activities in brain tissues colchicine significantly decreased SOD, CAT, GSH activities in colchicine treated experimental Alzheimer's group rather than control, BH treated group and BH pretreated colchicine treated experimental Alzheimer's groups. BH containing vitamin E and beta-carotene significantly increased SOD, CAT, GSH, number of correct choices along with significantly decreased the latency time in a dose dependent

manner rather than other groups. Glutathione is an endogenous antioxidant, which is present majorly in the reduced form within the cells. It prevents the hydroxyl radical generation by interacting with free radicals. During this defensive process, reduced glutathione is converted to oxidized form under the influence of the enzyme glutathione peroxidase (GPX). The decreased level of reduced glutathione in colchicine treated experimental group seen in our study indicates that there was an increased generation of free radicals and the reduced glutathione was depleted during the process of combating oxidative stress (Reiter, 2000; Schulz et al, 2000). This has probably been possible either from the low level of ROS production or through a rapid dissolution of ROS that has further been strengthened from the elevated activities of important antioxidant defense enzymes CAT and SOD, studied in this experiment. Literature study has shown that the BH contains high level of vitamin E and beta-carotene which protects rat neurons against oxidative stress possibly through the presence of both vitamin E and beta-carotene. Because vitamin E (alpha tocopherol and other tocopherol) is the most potent antioxidant that can break the propagation of free radical chain reactions in the lipid part of biological membranes. It may be inferred from the present results BH protects rat neurons against oxidative stress as is evidenced from our results of LPO, CAT, SOD and GSH activities possibly by vitamin E and beta carotene which is present in BH. So, keep in this view, i.c.v. administration of colchicines may generate oxidative stress either by producing reactive oxygen species (ROS) or by hampering the endogenous antioxidant enzymes leading to cholinotoxicity. The aqueous pulp extract of this plant containing vit- A, C, E results significant protection in the level of antioxidant status in frontal cortex and hippocampas after a certain period of co administration on colchicine induced oxidative stress without causing any general and metabolic toxicity and possibly thereby induced frontal cortical and hippocampal concentration of Ach and ChAT activity.

# **CONCLUSION**

The present investigation, together with earlier reported clinical and experimental data, and the similarly of behavioural and biochemical effects on cholinergic markers and different antioxidant enzyme activity along with LPO level, permit the categorization of BH as a nootropic agent. However, it may be proposed that further research on this field is essential to find out other active ingredients present in the BH pulp extract and their specific role by which the therapeutic importance may be evaluated and the outcome of which can be utilized in the protection of AD.

#### **ACKNOELEDGMENTS**

We are highly indebted to B.Sc final year students, Berhampore Girls' College, University of Kalyani for their valuable co-operation. The authors have no conflicts of interest and there is no funding agency to support of this study and its publication.

# **REFERENCES**

- 1. Bhattacharya SK, Bhattacharya A, Das K, Muruganandam AV, Sairam K. (2001). Further investigations on the antioxidant activity of *Ocimam sanctum* using different paradigms of oxidative stress in rats. *J Natural Remedies*, 1(1): 6-16.
- 2. Cantuli CI, Shukitt-Hale B, Joseph JA. (2000). Neurobehavioral aspects of antioxidants in aging. *Int J Dev Neurosci*, 18(4-5): 367-81.
- 3. Chouinard G, Annable L, Ross-Coomard A, Oliver M, Fontaine F. (1983). Piracetam in elderly psychiatric patients with mild diffuse cerebral impairment. *Psychopharmacology*, 81(2):100-06.
- 4. Cohen G, Dembiec D, Marcus J. (1970). Measurement of Catalase activity in tissue extracts. *Annals Biochem*, 34:30-37.
- 5. Dianzani MU. (1985). Lipid peroxidation in ethanol poisoning: A critical reconsideration. *Alcohol*, 20: 161-73.
- 6. Dwaine FE, Thoams JW. (1990). Cholinergic cell loss and cognitive impairments following intracerebroventricular of intradentate injection of colchicine. *Brain Res*, 517: 157-67.
- 7. Ellman GL. (1959). Tissue sulfhydryl groups. *Arch Biochem Biophys*, 82: 70-77.
- 8. Emerich DF, Walsh TJ.(1990). Intracerebroventricular injection in rats. *Brain Res*, 517: 157-60.
- 9. Frei B, England L, Ames BN. (1989). Ascorbate is an outstanding antioxidant in Human blood plasma. *Proc Natl Acad Sci*, 86: 6377-381.
- 10. Glowinski J, Iversen LL. (1966). Regional studies of catecholamines in the rat brain. I. The disposition of [3H] norepinephrine, [3H] dopamine and [3H]dopa in various regions of the brain. *J Neurochem*, 13(8):655-69.
- 11. Haba K, Ogawa N, Kawata M, Mori A. (1988). A method for parallel determination of choline acetyltransferase and muscarinic cholinergic receptors: application in aged-rat brain. *Neurochem Res*, 13 (10):951-55.
- 12. Husain K, Somani SM. (1997). Interaction of exercise and ethanol on hepatic and plasma antioxidant system in rat. *Pathophysiology*, 4, 69-74.

- 13. Jaiswal AK, Bhattacharya SK. (1992). Effects of shilajit on memory, anxiety and brain monoamines in rats. *Indian J Pharmacol*, 24:12-17.
- 14. James FF, Dennis WL. (1981). Long term memory: disruption by inhibitors of protein synthesis and cytoplasmic flow. *Pharmacol Biochem Behav*, 15:289 96.
- 15. Kevin PN, William RM, Hugh AT. (1989). Colchicine induced alterations of reference memory in rats: role of spatial versus non-spatial task components. *Behav Brain Res*, 35: 45-53.
- 16. Koti ST. (1991). Effect of Mentat on school student's performance. *Probe*, 3: 250-52.
- 17. Lonnrot K, Metsa-Ketela T, Molnar G, Ahonen JP, Latvala M, Peltola J, Pietila T, Alho H. (1996). The effect of ascorbate and ubiquinone supplementation on plasma and CSF total antioxidant capacity. *Free Radic Biol Med*, 21:211-17.
- 18. McClure WO. (1972). Effects of drugs upon axoplasmic transport. *Adv Pharmacol Chemother*, 10:185-20.
- 19. Mishra HP, Fridovich I. (1972). The generation of radical during superoxide auto oxidation of hemoglobin. *J Biol Chem*, 34:30-37.
- 20. Mitchell JJ, Paiva M, Heaton MB. (1999). Vitamin E and beta-carotene protect against ethanol combined with ischemia in an embryonic rat hippocampal culture model of fetal alcohol syndrome. *Neurosci Lett*, 263(2-3):189-92.
- 21. Moos WH, Davis RE, Schwarz RD, Gamzu ER. (1988). Cognition activators. *Med Res Rev*, 8(3): 353–91.
- 22. Nadkarni AK. (1976). In: Indian Materia Medica. Papular Prakashan. Bombay, India. 185-86.
- 23. Pratico D, Delanty N. (2000). Oxidative injury in disease of the central nervous system: focus on Alzheimer's disease. *Am J Med*, 109: 577-85.
- 24. Reiter RJ. (2000). Melatonin: lowering the high price of free radicals. *News Physiol Sci*, 15: 246-50.
- 25. Ribiere C, Hininger I, Rouach H, Nordmann R. (1992). Effects of chronic ethanol administration on free radical defense in rat myocardium. *Biochem Pharmacol*, 44: 1495-500.
- 26. Rice ME. (2000). Ascorbate regulation and its neuroprotective role in the brain. *Trends Neurosci*, 23: 209-16.

- 27. Roy C, Ghosh TK, Guha D. (2007). The antioxidative role of *Benincasa hispida* on colchicine induced experimental rat model of Alzheimer's disease. *Biogenic Amines*, 21(1-2): 44-57.
- 28. Schulz JB, Lindenau J, Seyfried J, Dichgans J. (2000). Glutathione, oxidative stress and neurodegeneration. *Eur J Biochem*, 267(16): 4904-911.
- 29. Spector B, Lorenzo AV. (1973). Ascorbic acid homeostasis in the central nervous System. *Am J Physiol*, 225: 757-63.
- 30. Speeg Jr K V. (1974). In: "Choline and Acetylcholine: Handbook of Chemical Assay Methods", Hanin I. (Ed). New York: Raven Press, 129-33.
- 31. Tagami M, Yamagata K, Ikeda K, Nara Y, Fujino H, Kubota A, Numano F, Yamori Y. (1998). Vitamin E prevents apoptosis in cortical neurons during hypoxia and oxygen reperfusion. *Lab Invest*, 78(11): 1415-429.
- 32. Veerendra Kumar MH, Gupta YK. (2002). Intracerebroventricular administration of colchicine produces cognitive impairment associated with oxidative stress in rats. *Pharmacol Biochem Behav*, 73(3): 565 71.
- 33. Venkateswaran S, Pari L. (2003). Effect of Coccinia indica leaves on antioxidant status in streptozotocin-induced diabetic rats. *J Ethnopharmacol*, 84: 163-68.
- 34. Verma A, Kulkarni SK. (1991). Effect of a herbal psychotropic preparation, BR-16A (Mentat), on performance of mice on elevated plusmaze. Indian J Exp Biol, 29, 1120-123.
- 35. Walsh TJ, Schulz DW, Tilson HA, Schmechel DE. (1986). Colchicine-induced granule cell loss in rat hippocampus: selective behavioral and histological alterations. *Brain Res*, 398: 23-36.
- 36. Wilson L, Friedkin M. (1966). The biochemical events of microtubule: L Synthesis and properties of colchicine labeled with tritium in its acetyl moiety. *Biochemistry*, 51: 2463-468.
- 37. Wollen WE, Faure R, Gaydou EM. (1991). A rare triterpene as major constitute of the "wax" on fruits of *Benincasa hispida*. *Indian drugs*, 28(10):458-60.
- 38. Yashizumi S, Murakam T, Kadoya M, Matsuda H, Yamahara J, Yoshikava M. (1998). Histamine release inhibitors from was guard the fruits of *Benincasa hispida* (thunb). *Yakugaku Zassi*, 118(5): 188-92.