

WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 5.045

Volume 4, Issue 2, 861-873.

Research Article

ISSN 2277-7105

EBOLA VIRUS-A PROTOTYPE PATHOGEN CAUSING HAEMORRHAGIC FEVER, AN EPIDEMIC HEALTH THREAT IN PUBLIC - WORLD'S NEXT AIDS- STATUS OF IT'S VACCINE AND RESEARCH DEVELOPMENT

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Article Received on 21 Nov 2014,

Revised on 16 Dec 2014, Accepted on 10 Jan 2015

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ABSTRACT

As the discovery of Ebola prototype pathogen as Ebola viral disease in 1976, the global spread of Ebola virus is reaching pandemic proportions, indicating a global developmental and threat in the community. The safety preventive measures, which can broadly and effectively control the spread of Ebola virus, can take it away from pandemic. In last few years, we observed that significant progress has been made in the areas like basic virology, pathogenesis of virus immunology and the development of various kinds of anti-retroviral drugs. However, the development of Ebola vaccine can be the measure to control its spread and treatment also. It is difficult to develop Ebola

virus vaccine because of abrupt onset of Ebola hemorrhagic fever having incubation period of 2-21 days, ban on conduct of clinical trial on the existing animal model and some logistical problems; it may also face the problems like high variability in the genetic mutation or genome of virus, lack of immune correlates of protection etc. The Ebola vaccine development needs more volunteers or candidates to be tested in phase I clinical trial. Manufactures of vaccines needs to produce protective immune reaction toward the surface protein, multiple vaccine concepts, subunit vaccines, including DNA vaccines, live recombinants. A number of candidate of EVD vaccines have been tested in animals, promising results are obtained in non-human primates. The article reviews, the art of development of Ebola virus therapy, considering Ebola vaccine, antiretroviral drugs and creating awareness of prevalence of the disease.

Keywords:Ebola virus, Ebola hemorrhagic fever (EHF), Antiretroviral drugs, Ebola Vaccines.

INTRODUCTION

The Ebola species reported in 1976 in the Democratic Republic of the Congo near Ebola, a small river. Since then, outbreaks have appeared sporadically.^[1] From 1976 (when it was first identified) through 2013, the WHO reported a total of 1,716 cases and since from the beginning the year, Ebola has killed nearly 3900 people in West Africa and can become world's next AIDS.^[2]

Ebola hemorrhagic fever remains a plague for the population of equatorial Africa, since 2000 with an increasing number of outbreaks. All human cases due to the emergence or remergence of Ebola species. Case fatality rate of African species in man are as high as 90%, as no prophylaxis or effective development of Ebola virus therapy is available. This infection is characterized by immune suppression and systemic inflammatory responses which causes impairment of vascular, coagulation and immune system that leads to multiorgan failure, resembling septic shock. [3]

Ebola hemorrhagic fever is caused by any of five genetically distinct members belonging to family Filoviridae: Zaire Ebola virus (EBOV); Sudan Ebola virus (SUDV); Tai forest Ebolavirus, (TAFV); Bundibugyo Ebolavirus (BDBV) and the fifth Reston Ebolavirus (RESTV). Reston Ebolavirus (RESTV) has only caused disease in nonhuman primates (NHP). Zaire, Sudan and Bundibugyo Ebolavirus are responsible for the most of the EHF outbreaks. EBOV constitutes serious threat in both human and NHP's in sub-Saharian Africa. Ebola hemorrhagic fever has been associated with large human outbreaks with case fatality rates as high as 90%. The case fatality rate of EBOV in NHP is unknown but some ecological data informs that it has contributed to declines of up to 98% of local great ape population in Gabon and the republic of Congo. [4]

Table 1: Species of genus Ebolavirus.

Species	Virus	Region	Fatality rate
Zaire Ebola virus	EBOV	Africa	60-90%
Sudan Ebola virus	SUDV	Africa	40-60%
Bundibugyo	BDBV	Africa	25%, based on one outbreak
Ebolavirus	TAFV	Africa	Unknown, only one known infection in Ivory
Tai forest Ebolavirus	RESTV	Asia	Coast
Reston Ebolavirus			Not known to cause lethal infection in humans.
			Lethal in non-human primates.

Ebola virus constitutes the family Filoviridae in the order of mononegavirales. Ebola virus particles have a uniform diameter of 80 nm but can vary in length, with lengths up to 14000 nm."Fig.1"

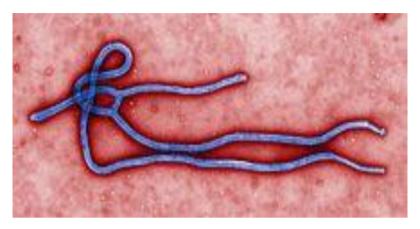


Figure 1: Ebola virus.

The genome consist of seven genes in order 3' leader, nucleoprotein(NP), virion protein (VP) 35, VP40, glycoprotein, VP30, VP24, RNA- dependent RNA polymerase(L)-5' trailer. In addition, EBOV expresses at least one nonstructural soluble GP (sGP) encoded by the GP gene. The GP, with variable contributions from other viral proteins such as NP, appears to be the key immunogenic protein in vaccine protection. With the exception of glycoprotein gene, all genes are monocistronic, encoding for one structural protein. An important distinction of Ebola virus from other mononegavirales is the production of a soluble glycoprotein, which is the primary product of the GP gene, and gets secreted to large quantities from infected cells.

Despite important achievements during the past two decades to unravel the molecular biology and pathogenesis of Ebola virus, we are still unclear about virulence factor and host responses, which seem to be partly detrimental to the host.^[6]

Ebola, a small river in Democratic Republic of the Congo, the first case recognized on 26th august 1976. Mabalo Lokela, a 44 year old school teacher suffered from hemorrhagic fever, The EVD emerged in the human population in the winter of 1976.^[7]

Now there is evidence that EVD is a zoonosis and its etiological agents, the Ebola virus, crossed the simian-human species barrier and arose among humans as a result of use of

unsafe injection and transfusion practices in postcolonial Africa. Today, EBOLA/EVD is the leading cause of death in sub-Saharan Africa and the fifth biggest killer in the world.^[8]

The virus may be acquired upon contact with blood or bodily fluids of infected animals. Spreading through the air has not been documented in the natural environment. Fruit bats are considered to be a most likely reservoir of the EBOV, carrier and may spread the virus without being affected. Three types of fruit bats (Hysignathus monstrosus, Epomops franqueti and Myonycteris torquata) have been identified as being in contact with EBOV. Antibodies against Ebola Zaire and Reston viruses have been found in fruit bats in Bangladesh, thus identifying potential virus hosts and signs of the Filoviruses in Asia. [9] Once human infection occurs, male survivors may be able to transmit the disease via semen for nearly two months. To confirm the diagnosis, blood samples are tested for viral antibodies, viral RNA or virus itself.

Proposed plan of action

Any virus has two key features-infectivity which refers to the speed with which the virus can spread, and virulence which indicate how fatal the infection can. Ebola is dangerous because it is highly virulent (it has fatality rate up to 90% as per the WHO), but its infectivity is not very high. Ebola is not a respiratory disease or air born infection. Comparing the situation with other viral infection carriers did not show any symptoms (H1N1), on the other hand, any one infected by the Ebola virus becomes contagious only when they begin to show symptoms. This makes the process of detection much easier. [10]

Although progress has been made in understanding virus biology, no licensed vaccines or treatments currently exist, as there is a desperate need for a vaccine that not only prevents it's transmission but also to control future incidences. Though inhalable Ebola vaccine passes test on animals with long-lasting protection after single inhaled dose. A next stage of vaccine research in human subjects entered in phase-I clinical trial.^[11]

Hoped to have vaccine will be available by Nov, 2014. Because DNA vaccines, adenovirus-based vaccine, and VSIV-based vaccines have entered clinical trials. Healthcare professionals would have to be alert and recognize the symptoms early.^[12]

Published studies have included expression of Ebola virus protein subunits in live vaccinia virus vectors, selected DNA plasmids, Ebola virus-like particles, replication-competent

vesicular stomatitis virus, Venezuela equine encephalitis virus replicons, recombinant parainfluenza virus type 3, replication-defective recombinant adenovirus (rAd), and DNA combined with rAd prime-boost strategie. While many of these approaches have been evaluated in a nonhuman primate model, only DNA/rAd-, rAd-, or vesicular stomatitis virus-based vaccines have shown efficacy in primates. [13]

This article emphasizes on the current status of Ebola outbreaks, its vaccines and research development. "Fig.2"

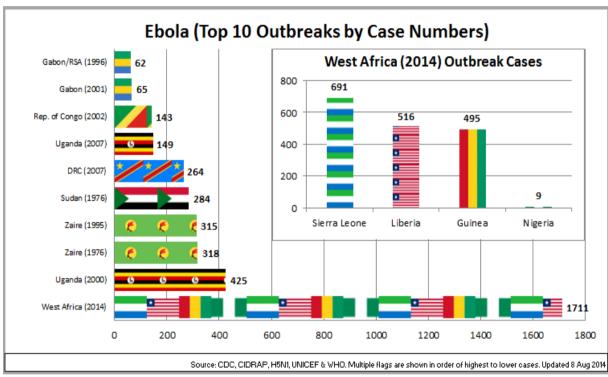


Figure 2: Ebola outbreaks.

Plan of action

Table 2:-Laboratory tests used in diagnosis include

Timeline of Infection	Diagnostic tests available		
Within a few days after symptoms begin	 Antigen-capture enzyme-linked immunosorbent assay (ELISA) testing IgM ELISA Polymerase chain reaction (PCR) 		
Later in disease course or after recovery	4. Virus isolation5. IgM and IgG antibodies		
Retrospectively in deceased patients	6. Immunohistochemistry testing7. PCR8. Virus isolation		

The rVSV platform has shown complete efficacy as a preventive single-shot vaccine in 3 relevant animal models, including some non-human primate models.

Table 2:-Overview of scientific information on unproven vaccines under development.

Type of vaccine What it does/state		Safety in humans	Availability	
	of research			
Chimpanzee adenovirus serotype 3 (ChAd3) vaccine	Uses a chimpanzee adenovirus that does not grow, containing the gene for EVD surface protein. In a study of animals given a lethal dose of EVD, all 16 were protected by a single dose of the vaccine.	More than 1300 people have received these vaccines for other diseases, including over 1000 people in Gambia, Senegal, Burkina Faso, and Kenya. These other vaccines seem safe so far, but as yet there is no safety information on an EVD vaccine in humans.	There is no information from human trials. An early trial of an EVD vaccine containing two EVD strains, Zaire and Sudan, is planned to start in September 2014 in the US. A vaccine against Zaire EVD may be evaluated in the UK, and then in one or two African countries in 2014. The earliest availability depends on results of trials and Manufacturing timelines. Approximately 15 000 doses might be available by the end of 2014.	
Recombinant Vesicular Stomatitis Virus (rVSV) vaccine	The rVSV vaccine aims to induce EVD-specific immune responses. The vaccine protected all 20 animals from a lethal dose of EVD. Animals with weakened immunity were not harmed by rVSV-EVD. The vaccine was safe when injected directly into the brain of animals.	The rVSV vaccine aims to induce EVD-specific immune responses. The vaccine protected all 20 animals from a lethal dose of EVD. Animals with weakened immunity were not harmed by rVSV-EVD. The vaccine was safe when injected directly into the brain of animals.	Safety, efficacy and duration of protection are unknown. An early trial is due to start soon in the US. Eight hundred doses are currently available.	

The above products were prioritized for consideration based on the availability of non-human primates (NHP) efficacy data with a filovirus challenge, and justification for a human dose based on the clinical data of the product.

The platform has shown partial to complete efficacy in post exposure treatment against homologous filovirus challenge. Knowledge gained from these data should advance the development of rVSV-based vaccines for human use.

The safety and immunogenicity of an Ebola virus vaccine in its first phase I human study. A three-plasmid DNA vaccine encoding the envelope glycoproteins (GP) from the Zaire and Sudan/Gulu species as well as the nucleoprotein was evaluated in a randomized, placebocontrolled, double-blinded, dose escalation study. Healthy adults, ages 18 to 44 years, were randomized to receive three injections of vaccine at 2 mg (n = 5), 4 mg (n = 8), or 8 mg (n =8) or placebo (n = 6). Immunogenicity was assessed by enzyme-linked immunosorbent assay (ELISA), immunoprecipitation-Western blotting, intracellular cytokine staining (ICS), and enzyme-linked immunospot assay. The vaccine was well-tolerated, with no significant adverse events or coagulation abnormalities. Specific antibody responses to at least one of the three antigens encoded by the vaccine as assessed by ELISA and CD4⁺ T-cell GP-specific responses as assessed by ICS were detected in 20/20 vaccines. CD8⁺ T-cell GP-specific responses were detected by ICS assay in 6/20 vaccines. This Ebola virus DNA vaccine was safe and immunogenic in humans. Further assessment of the DNA platform alone and in combination with replication-defective adenoviral vector vaccines, in concert with challenge and immune data from nonhuman primates, will facilitate evaluation and potential licensure of an Ebola virus vaccine under the Animal Rule.

Table 3: Most promising vaccine platforms with efficacy in non-human primates.

Platform	Immunogen	Characteristic	Preexposure (Efficacy)	Post-exposure (Efficacy)	Concerns
Adenovirus, type 5 (Ad5)	GP; NP; GP/NP	Replication- deficient	Single vaccination (100%)	Not reported	Preexisting immunity; high dose
Virus-like particles (VLPs)	VP40/NP/GP	Replication-deficient	Multiple vaccinations (100%)	Not reported	Production; boost immunization
Venezuelan Equine Encephalitis virus (VEEV) replicon	GP; NP; GP/NP	Single round replication	Multiple vaccinations (0%–100%)	Not reported	Boost immunization
DNA	GP; NP	Replication-deficient	Multiple vaccinations (0%–100%)	Not reported	Boost immunization; efficacy questionable
Human para- influenza virus, type 3 (HPIV3)	GP; GP/NP	Replication- competent (live- attenuated)	Single vaccination (50%100%)	Not reported	Preexisting immunity; safety
Vesicular stomatitis virus (VSV), Indiana serotype	GP	Replication- competent (live- attenuated)	Single vaccination (100%)	Single treatment (50%–100%)	Safety

Note: GP- glycoprotein, NP- nucleoprotein, VP 40- virion protein 40 k Da.

Study population demographics

A total of 27 healthy adult volunteers were enrolled, with 5 in the 2-mg dose group, 8 each in the 4-mg and 8-mg dose groups, and 6 in the placebo group. The subject population was 66.7% male and 33.3% female with a mean age of 33.1 years (range of 18 to 44 years). Subjects were predominantly white (96.3%) and non-Hispanic/Latino (96.3%). The mean BMI was 25.6 (range, 19.5 to 42.2). All subjects had an educational level of high school or higher, with 48.1% having college level degrees and 33.3% holding advanced degrees."Fig.3"

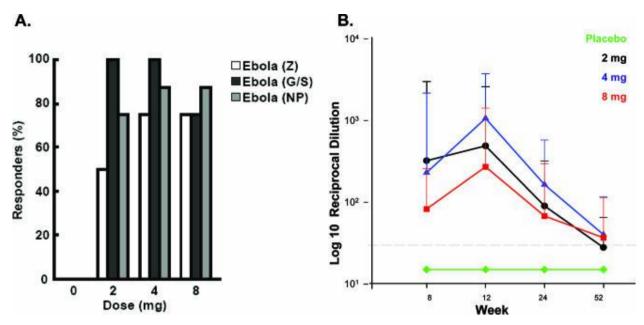


Figure 3: Kinetics and frequency of antibody responses.

(A) Percentages of responders following the third vaccination by the IP-Western assay for all subjects are shown. The y axis represents the percentage of responders with a positive assay, and the x axis represents the vaccine dose group. White bars, GP (Z); black bars, GP (S/G); gray bars, NP (Z). (B) Kinetics of the antibody response for all subjects is shown over the 52 weeks of the study. The geometric mean titer of the \log_{10} reciprocal dilution and standard deviation of the antibody response to GP (S/G) are plotted against the number of weeks after initial vaccination for each of the three dose levels. Vaccinations were given at 0, 4, and 8 weeks. The threshold for positivity in this assay was a reciprocal dilution of 30 and is shown as a dashed line. Of note, only six of eight subjects in the 8-mg dose group received all three vaccinations in the series, yet all vaccines are included in the immunogenicity analysis.

Vaccine safety

Due to a theoretical concern over GP-mediated cytopathicity, coagulation parameters of study subjects were closely monitored. At enrollment and throughout the study, D-dimer, prothrombin time, partial thromboplastin time, fibrinogen, complete blood count, and red blood cell smears were evaluated. There were no reportable coagulation laboratory abnormalities.^[14]

Two subjects were withdrawn from the vaccination schedule due to serious adverse events that were assessed as "possibly" related to vaccination: a grade 4 creatine phosphokinase elevation 2 weeks after first vaccination and a grade 2 herpes zoster thoracic dermatome

eruption 3 weeks after the second vaccination, both in 8-mg recipients. Of note, the grade 4 creatine phosphokinase elevation was associated with vigorous exercise. These events resolved without sequelae, and these subjects continued to participate in the study and attended all study visits. Although only six of eight subjects in the 8-mg dose group received all three injections, the immunogenicity and safety laboratory values for all subjects are included in the analyses. One subject in the 2-mg dose group chose to withdraw after the second vaccination; another subject (in the placebo group) withdrew after the third injection. Neither of these subjects returned for further visits and, therefore, they were not included in the immunogenicity analysis due to a lack of samples at time points following their withdrawal. As a result, 20 of 21 vaccines had immune responses assessed. All subjects are represented in the safety data through the time points available.

The diary cards showed that 90.5% (19/21) of subjects who received vaccine (at any dose level) experienced at least one local injection site symptom (mild to moderate pain/tenderness, mild induration, or mild skin discoloration) following a vaccination. The systemic symptoms recorded on diary cards included malaise, myalgia, headache, nausea, and fever, as well as local injection site symptoms. The study vaccinations were well-tolerated and safe in healthy subjects, ages 18 to 44 years.

DISCUSSION – PUBLIC HEALTH IMPACT

The subsequent signs and symptoms includes systemic (prostration), gastrointestinal (anorexia, nausea, vomiting, abdominal pain, diarrhea), respiratory (chest pain, shortness of breath, cough, nasal discharge), vascular (congectival injection, postural hypotension, oedema), and neurological (headache, confusion and coma), manifestations.

Patients develop clinical signs during infections and die typically between day 6 and 16 with hypovolemic shock and multiorgan failure. Hemorrhages can be serious but are only present in fewer than half of patient.

Pregnant woman may have an increased risk of miscarriage and clinical findings suggest high death rate for children of infected mothers. This high death rate could be due to transmission from the infected mother to the child during breast feeding, either through milk or close contact.

People who are living in or travelling to affected areas of Africa may be at risk of infection; however, this risk is extremely low unless there has been direct exposure to the bodily fluids of an infected person (including unprotected sexual contact with confirmed cases up to three months after they have recovered), or infected animal (alive or dead).

Caring for relatives with EVD is a known risk factor for infection, and healthcare workers, particularly those in resource poor settings with inadequate infection control are also at risk. During outbreaks of Ebola HF, the major prevalence of the virus includes healthcare workers, their family and friends. Medical professionals are also at high risk of infection.

CONCLUSION

The broad immunogenicity of this Ebola virus DNA vaccine suggests that immunization by plasmid DNA delivery is a viable platform and merits further development. Nonhuman primate studies have shown that a rAd5 vaccine effectively prevents disease, and DNA vaccination prior to boosting with rAd5 also confers protection and markedly increases the magnitude of the immune response. Therefore, future formulations of this DNA product will include multiple GP constructs encoding GP in either its wild-type form or a modified form to optimize vaccine potency. Because Ebola virus from Ivory Coast has been observed in only one limited outbreak and is closely related to Ebola virus Zaire, it is not included in vaccine formulations.

Since the prophylactic efficacy of an Ebola virus vaccine cannot feasibly or ethically be demonstrated in a human trial, the combination of safety and immunogenicity data from phase I, II, and III human trials and efficacy data from nonhuman primate studies will ultimately need to be utilized to obtain licensure of an Ebola virus vaccine under the Animal Rule.

The successful evaluation of a DNA vaccine to multiple Ebola virus subtypes reported here provides the opportunity for further clinical evaluation of candidate Ebola virus DNA vaccines alone or in combination with Ebola virus rAd vaccines as a heterologous prime-boost strategy.

There is a need of development of safe, effective and affordable Ebola vaccine that constitutes a top priority for Ebola hemorrhagic fever research and formidable scientific and

public health challenges at the dawn of this year, so that Ebola could not become world's next AIDS.

AKNOWLEDGEMENT

An enormous body of the work has been contributed to the knowledge of status of Ebola vaccine and Research Development. With that said, we acknowledge the research that has been carried out is mentioned in this review. This work has been carried out at Siddhant College of Pharmacy, Sudumbare, Pune.

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