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Research Article

ASSESSMENT OF THE PROTECTIVE ROLE OF HESPERIDIN AGAINST GENOTOXIC AND BIOCHEMICAL EFFECTS OF CYTOXAN IN MICE

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ABSTRACT

Hesperidin (HDN) is one of naturally occurring flavonoid widely found in citrus fruits. Flavonoids were found to inhibit DNA and chromosomal damage induced by cyclophosphamide. Chemoprevention is one of the most promising and realistic approaches in the prevention of side effects produced by cytoxan treatment as an anticancer drug. Therefore, the present study was designed to evaluate the protective effect of HDN against genotoxic and biochemical effects of cytoxan (anti-tumor drug) in Swiss albino male mice. Mice were divided into 19 groups (5 animals each). The 1st gp served as control (Their row is under the tables). The

2nd , 3rd , 4th , gps had divided into 9 subgroups and received orally 50, 100 and 200mg/kg bw of cytoxan for one, two and three weeks, respectively. The 5th , 6th , 7th gps had divided into 9 subgroups and received 50, 100 and 200 mg/kg bw of cytoxan in combination with 50 mg of HDN for 1, 2 and 3 weeks, respectively. At the end of the experimental periods, animals were subjected to genetic and biochemical analysis. The results revealed that MN frequency significantly increased in cytoxan treated mice. However, HDN treatment showed a significant decreament in MN frequency in mice bone marrow cells compared to control. As well, a significant inhibition in DNA content using comet assay in cytoxan treated groups with about 3 fold lower than control, whereas the treatment with HDN in improved that content. Also, there was a significant reduction in DNA, RNA and protein content in brain and kidney tissues and functions (creatinine and urea level elevation). However, administration of HDN in combination with cytoxan significantly improved all parameters studied in brain and kidney tissues. A significant decrease in the level

of tissue antioxidants like superoxide dismutase (SOD), cholinesterase (CHE) and increase in malondialdhyde (MDA), cholesterol, glucose and triglycerol levels was found in cytoxan treated groups. In conclusion, HDN supplementation significantly ameliorate these parameters due to antioxidant activity, thereby showing potent antigenotoxic and chemopreventive effects against toxicity induced by cytoxan in mice.

KEYWORDS: Hesperidin, Antioxidant, Cytoxan, Genotoxicity, biochemical, mice.

INTRODUCTION

Hesperidin (HDN) is one of naturally occurring flavonoid widely found in citrus fruits. HDN as a flavonoid was found to inhibit DNA & chromosomal damage induced by cyclophosphamide (CPA). Flavonoid compounds have many biological properties, including hepatoprotective, antibacterial, antimutagenic and anticancer activities. HDN is a flavonone glycoside, belonging to the flavonoid family. This natural product is found in citrus species. Citrus extract had a significant protective effect on genotoxicity induced by cyclophosphamide. HDN was reported to have many biological effects including antiinflammatory, antimicrobial, anticarcinogenic, antioxidant effects, and decreasing capillary fragility. [3]

Cytoxan (Cyclophosphamide, CPA) is an alkylating anti-tumor drugused for the treatment of various cancer and noncancer disorders. It is a member of oxazophorine group and its chemical formula is C7H15Cl2N2O2P. [4] CPA shows antitumor activities against a broad range of cancers including malignant lymphomas, myeloma, leukemia, neuroblastoma, adeno-carcinoma, retino-blastoma, and breast-carcinoma. [5-7] CPA is also used for the mobilization of hematopoietic progenitor cells from the bone marrow into peripheral blood. [8,9] It is also a well known immunosuppressive agent used for graft rejection in case of renal, hepatic, and cardiac transplantation. [10]

For micronucleus (Mn) assay, the studies of ^[15,16] showed that oral administration of CPA in mice and rats resulted in a significant increase in the frequency of Mn and that depends on the dose and sampling times. In addition, when CPA was administered to rat, a significant increase in chromosomal aberration and Mn formation was found in a dosedependent manner in BM cells of rat.^[17] Administration of citrus extract before CPA treatment significantly reduced the frequency of Mn in mice compared with the group treated with CPA alone. Citrus extract, particularly flavonoids constituents with antioxidant

activity, reduced Mn 2.8 fold against genotoxicity induced by CPA.^[2] The same findings were found by.^[18,19] A significant increase in comet tail length and Mn percentage in both acute (single dose) and subacute (multiple doses repeated every 24 h for 7 times) studies of CPA treated mice.^[20,21] The same finding was reported by.^[22] CPA inhibits embryonic DNA synthesis and does so prior to its effect on RNA or protein synthesis.^[23]

Although CPA is known to produce DNA cross-links, other DNA lesions are produced as well. CPA mediates G0/G1 and S phase arrest. Accumulation of cells in G0/G1 in comparison to the control, whereas higher concentrations causes dose-dependent G0/G1, Sphase and G2/M phase inhibition.^[24] However, orange juice was found to reduced the extent of DNA damage caused by CPA in mice due to its antigenotoxic effect.^[25, 26] The protective effect against that damage was associated with modulation of lipid peroxidation as well as an increase in GSH and the GSH-dependent enzyme glutathione S-transferase.^[26] These findings indicate that intake of HDN can lead to protection against invivo genotoxicity and oxidative stress. Effect of CPA on biochemical parameters studied by^[27,28] in rats showed the decreased level of serum total protein and increased levels of urea and creatinine than the control animals.

An increase in malondialdhyde (MDA) level and decrease in superoxide dismutase (SOD) level was found in CPA treated Wistar male rats, that also happened in mice due to reproductive toxicity.^[29,30] However, HDN has shown to increase the level of SOD in the mice and rats.^[31,32]

In addition, an increase in cholesterol, glucose, triglycerol, MDA and Mn frequency levels in CPA treated mice, whereas a decrease in total protein level where found by.^[33] To our knowledge this is the first report to study the effect of HDN as a chemopreventive agent in combination with cytoxan on cholesterol, glucose and triglycerol levels in mice.

Therefore, the present study was designed to evaluate the protective effect of hesperidin (HDN) against genotoxic and biochemical effects of cytoxan (anti-tumor drug) treated Swiss albino male mice.

MATERIALS AND METHODS

Animals

Adult Swiss albino male mice weighting 25-30 gm were used in the present study. Animals were housed in cages of the Animal house laboratory of the National Research Center, Dokki, Cairo, Egypt and had free access to water and pellet diet for one week for adaptation.

Drugs

- I Cytoxan were purchased from Sigma Aldrich Chemical Private Ltd., India is used in the treatment of mice under study.
- II- Hesperidin was supplied by Sigma Aldrich Chemical private Ltd., India and suspended in distilled water and administered orally and used as a protective.

Treatment

Mice were divided into 19 groups (5 animals each). The 1st gp served as control (Their row is under the tables). The 2rd, 3th, 4th, gps had divided into 9 subgroups and received orally 50, 100 and 200mg/kg bw of cytoxan for one, two and three weeks, respectively. The 5th, 6th, 7th gps had divided into 9 subgroups and received 50, 100 and 200 mg/kg bw of cytoxan in combination with 50 mg of

HDN for 1, 2 and 3 weeks, respectively. At the end of the experimental periods animals were anaesthetized and sacrificed by cervical dislocation and subjected to genetical and biochemical analysis.

Genetic analysis

Micronucleus assay: Mice were sacrificed and both femurs of mice were removed and aspirated with fetal calf serum.^[34] The bone marrow smears were made, fixed and stained with Giemsa.^[35] 2000 polychromatic erythrocytes were scored per animal.

Comet assay: The comet assay were performed according to Comet assay reagent kit for single cell gel electrophoresis assay (Catalog \neq 4250-050-k). DNA migration, image length, nuclear size and DNA damage were calculated in mice brain cells.

Determination of Nucleic Acids

Nucleic acids (DNA and RNA) were determined using a simplified method for determination of specific DNA or RNA. [36]

Biochemical analysis

Blood samples were collected in heparinized tubes and centrifuged at 5000 rpm for 10 min for quantitative measurement of lipid peroxidase malondialdhyde (MDA) according to.^[37] Superoxide dismutase (SOD) activity was assayed by the method of.^[38] Cholinesterase (CHE) was assayed by the method of.^[39] Biochemical estimation of cholesterol was developed according to.^[40] Triglyceride was measured according to^[41] and urea, creatinine and glucose were determined by a differential pH technique according to.^[42] Total protein content was measured colorimetrically according to.^[43]

Statistical analysis

Data are presented as Means \pm SE. One way analysis of variance (ANOVA) and Tukey's HSD test were used for multiple comparisons of data.

RESULTS AND DISCUSSION

In the present study, HDN was investigated for its potential antigenotoxic and chemopreventive effects in CP treated Swiss albino mice by genetic (Mn and comet assays) and biochemical analysis.

The results of the present study revealed that the treatment with cytoxan drug resulted in significant increase in the frequency of MN with 2 fold compared to control (Table 1). However, HDN treatment showed a significant decrease in MN frequency in mice bone marrow cells compared to cytoxan treated groups.

The comet assay (mean comet tail length) results showed a significant increase in comet tail length in cytoxan treated groups up to 3 fold higher than control, whereas the treatment with HDN improved that results (Table 2).

The antigenotoxic activity of HDN was evaluated by measuring their inhibitory effect on CP (cytoxan) induced genotoxicity in mice. It is indicated that CP induced chromosomal damage in mouse bone marrow cells. These fragmented chromosomes were condensed to form Mn [44]. HDN decreased the CPA induced formation of Mn, which may due to the inhibition of CPA induced chromosomal and DNA damage.

Cytoxan + HDN Cvtoxan Number Number of % of cells % of cells **Treatment** Dose examined of period with with mg/kg $M \pm S.E.$ $M \pm S.E.$ animals cells micronuclei micronuclei 2000 11.2±0.88 50.0 5 2.80 2.15 8.6 ± 0.60 5 2000 2.20 1 Week 100.0 3.10 12.4±1.04* 8.8 ± 0.55 200.0 5 3.40 13.8±0.65 2.25 2000 9.0 ± 0.61 5 2000 3.00 12.0±0.79^{*} 9.2 ± 0.42 50.0 2.30 5 13.0±0.80° 2Weeks 100.0 2000 3.20 2.35 9.4 ± 0.57 5 200.0 2000 3.70 15.0±1.00° 2.40 9.6 ± 0.57 50.0 5 2000 3.60 14.4±1.20° 2.50 10.0 ± 0.80 3Weeks 100.0 5 2000 3.90 15.6±1.03° 2.60 10.4 ± 0.84 200.0 5 2000 4.10 16.4±0.91 2.70 10.8±1.08 Control 0.0 2000 2.05 8.4 ± 0.57

Table 1: Effect of HDN on Cytoxan induced micronuclei in male mice.

Results are shown as mean \pm SE

The last row indicate the control group data.

*=P<0.05, **=P<0.01 and ***=P<0.001.

The antitumor agent CPA induced genotoxicity in mice and rats based on evaluation of Mn frequencies. A dose response for CPA —induced DNA damage was detectable in mice and rats by the comet assay. Evaluation of the nature of the CPA— induced Mn in mice and rats revealed that the Mn were primarily due to breakage events and contained chromosomal fragments. While Showed that oral administration of CPA in mice and rats resulted in a significant increase in the frequency of Mn and that depends on the dose and sampling times. As well with, who reported a significant increase in comet tail length and Mn frequency in cytoxan treated mice.

Table2: Mean comet tail length (μm) of mice brain exposed to cytoxan and /or HDN.

Treatment	Dose	Mean comet tail length (Mean ± S. E)	Mean comet tail length (Mean ± S. E)
period	mg/kg	Cytoxan	Cytoxan + HDN
1 Week	200.0	18.15±1.839*	13.94±1.034
2 Weeks	200.0	22.29±1.72***	17.842±2.078**
3 Weeks	200.0	29.34±2.872***	20.828±1.301***

	Control	0.00	13.17±1.072	
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Results are shown as mean \pm SE

The last row indicate the control group data.

*= P<0.05**= P<0.01 and***= P<0.001

Consumption of orange juice containing HDN can protect DNA from damage induced by CPA. Such protective effects of orange juice may be mediated by, (1) modulation of phase I and II enzymes; (2) substrate competition for the nucleophilic action of CPA or quenching of CPA metabolities and side-products (reactive species); and (3) enhancement of DNA repair. [25]

Oral administration of cytoxan (50, 100 and 200mg/kg bw) to male mice for 1, 2 and 3 weeks caused a significant reduction in DNA, RNA and protein content in brain (Table 3) and kidney (Table 4) tissues. These findings are in accordance—with, who found that CPA inhibits DNA synthesis and does so prior to its effect on RNA or protein synthesis. As well, a significant damage in kidney function (creatinine and urea) in 3 fold with 3 doses used in the study. However, administration of HDN in combination with cytoxan significantly improved all parameters studied in brain and kidney tissues and that coincide with who said that increased of urea could be due to increased protein level or due to impairment of renal function in CPA intoxication. As well a significant decrease in total protein level was found in CPA treated rats. However, no differences were found in creatinine and urea levels between CPA and morin (a naturally occurring flavonoid) treated groups. CPA mediates G0/G1 and S phase arrest. Accumulation of cells in G0/G1 in comparison to the control, whereas higher concentrations causes dose-dependent G0/G1, Sphase and G2/M phase inhibition.

Table 3: Effect of Cytoxan and/or HDN on DNA, RNA and protein of mice brain.

Treatment	Dose		Cytoxan			Cytoxan + HDN	
period	mg/kg	DNA	RNA	Protein	DNA	RNA	Protein
	50.0	$0.39^{***} \pm 0.02$	0.272*±0.013	$9.046^* \pm 0.105$	0.491±0.012	0.312±0.012	9.606±0.131
1 Week	100.0	0.39***±0.12	0.250**±0.14	8.687**±0.237	0.481±0.240	0.304±0.014	9.453±0.251
	200.0	0.37±0.01	0.235***±0.006	8.473***±0.166	0.469±0.012	0.296±0.012	9.392±0.137
	50.0	$0.38^{***} \pm 0.01$	0.264*±0.014	8.685**±0.276	0.488±0.025	0.310±0.013	9.501±0.195
2 Weeks	100.0	0.36±0.01	0.244**±0.008	8.356**±0.253	$0.450^* \pm 0.016$	0.294±0.012	9.410±0.187
	200.0	$0.35^{***} \pm 0.02$	0.218±0.010	8.344***±0.110	$0.448^* \pm 0.012$	0.283±0.0017	9.201±0.223
	50.0	$0.36^{***} \pm 0.02$	0.257**±0.017	8.276***±0.207	$0.458^* \pm 0.011$	0.291±0.008	9.283±0.204
3Weeks	100.0	$0.34^{***} \pm 0.02$	0.235***±0.006	8.150***±0.193	$0.440^* \pm 0.240$	0.280±0.014	9.101±0.149
3 WEEKS	200.0	0.31±0.02	0.198±0.006	8.080±0.170	0.436*±0.019	0.267±0.001	9.060±0.220

Control 0.0 0.491±0.01 0.323 ±0.014 9.835±0.283

Results are shown as mean \pm SE

The last row indicate the control group data.

*= P<0.05, **= P<0.01 and ***=P<0.001.

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Table 4: Effect of HDN on kidney (DNA, RNA, Protein) and kidney function (Creatinine, Urea) of mice treated with Cytoxan

Treatment	Dose			Cytoxan				C	'ytoxan + H	DN	
period	mg/kg	DNA	RNA	Protein	Creatinine	Urea	DNA	RNA	Protein	Creatinine	Urea
	50.0	0.331***	0.264^{*}	6.563*	0.660*	34.900**	0.417	0.316	7.859	0.588	30.880
	30.0	±0.015	± 0.018	± 0.449	±0.026	± 0.488	± 0.011	± 0.014	±0.490	±0.012	±0.285
1 Week	100.0	0.314***	0.250^{*}	6.149^{*}	0.699**	34.780***	0.394	0.290	7.062	0.616	31.160
1 Week	100.0	±0.019	±0.012	± 0.236	±0.030	± 0.339	± 0.009	± 0.018	±0.541	±0.012	±0.348
	200.0	0.308	0.218**	5.761**	0.743***	35.110****	0.381	0.278	6.647	0.625	32.050
	200.0	±0.013	± 0.006	± 0.382	±0.018	± 0.408	±0.019	± 0.009	± 0.550	±0.018	±0.618
	50.0	0.328	0.259**	6.443*	0.670^{*}	34.970***	0.407	0.308	7.486	0.600	31.850
	30.0	±0.013	± 0.023	± 0.549	±0.028	± 0.580	± 0.110	± 0.015	± 0.478	±0.010	± 0.567
2 Weeks	100.0	0.311***	0.203***	5.920**	0.701**	35.870***	0.391	0.284	6.961	0.626	32.350
2 WEEKS	100.0	± 0.008	± 0.006	± 0.200	±0.021	± 0.392	± 0.008	±0.012	± 0.464	±0.010	± 0.690
	200.0	0.304	0.192***	5.713**	0.814***	37.300****	0.377	0.274	6.628^{*}	0.639	33.160
	200.0	±0.014	± 0.010	± 0.253	±0.013	± 0.606	± 0.008	± 0.008	± 0.440	±0.020	± 0.844
	50.0	0.310	0.237^{**}	6.401^{*}	0.715***	36.920***	0.406	0.282	7.123	0.607	32.060
	30.0	± 0.008	± 0.008	± 0.516	±0.210	± 0.456	± 0.005	± 0.009	± 0.555	±0.012	±0.340
3 Weeks	100.0	0.306***	0.200***	5.701**	0.883***	37.670 ^{**}	0.365^{*}	0.270	6.815	0.630	33.820
3 WEEKS	100.0	±0.014	± 0.007	± 0.288	±0.025	± 0.444	± 0.017	± 0.017	± 0.586	±0.022	± 0.847
	200.0	0.295***	0.188***	5.438**	1.001***	40.160***	0.343^{*}	0.267	6.824*	0.649*	34.720 [*]
	200.0	±0.015	±0.014	± 0.376	±0.009	± 0.494	± 0.014	± 0.010	±0.326	±0.016	± 0.950

Control 0.0 0.429±0.02 0.319±0.019 8.166±0.431 0.571±0.023 30.670±0.727

Results are shown as mean \pm SE

The last row indicate the control group data.

*=P<0.05, **=P<0.01 and ***=P<0.001.

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Serum cholinesterase (CHE) with antioxidant SOD values were significantly decreased ($p \le 0.01$), whereas MDA level was increased with cytoxan treatment compared to control. That happened in mice due to reproductive toxicity. ^[30] However, the treatment with HDN improved these values in all treated groups (Table 5).

Table 5: Effect of HDN on blood MDA, SOD and CHE of Cytoxan intoxicated male mice.

Treatment	Cytoxan	Number of		Cytoxan		Cyt	oxan + HDN(50 mg	/kg bw)
period	Dose (mg/kg)	Animals	MDA	SOD	СНЕ	MDA	SOD	СНЕ
	50.0	5	9.65±0.69 [*]	$6.27\pm0.33^*$	6.009±0.40**	7.87±0.55	7.51±0.44	7.604±0.29
1 Week	100.0	5	10.19±0.87*	6.11±0.39*	5.773±0.32**	8.36±0.54	7.13±0.35	7.385±0.24
	200.0	5	10.51±0.83	5.78±0.36**	5.193±0.26***	8.96±1.06	6.74 ± 0.42	7.164±0.24
	50.0	5	$10.20\pm0.88^*$	$6.21\pm0.48^*$	5.863±0.35**	8.46±0.74	7.38 ± 0.50	7.495±0.23
2 Weeks	100.0	5	11.04±0.93	$6.06\pm0.45^*$	5.693±0.29**	8.66±0.57	6.83 ± 0.43	7.268±0.28
	200.0	5	11.89±0.58	5.49±0.35***	5.042±0.34***	9.04±0.82	6.64 ± 0.38	7.156±0.22
	50.0	5	11.18±1.13	5.53±0.42**	4.249±0.26***	9.23±0.80	7.06 ± 0.46	6.841±0.25*
3 Weeks	100.0	5	12.39 ± 0.92	5.33±0.34***	4.222±0.24***	9.61±0.88	6.74 ± 0.42	6.542±0.35*
	200.0	5	13.20±0.59	4.75±0.40	3.758±0.24***	9.66±0.65*	6.37±0.63	6.505±0.26*

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	Control	0.0	5	7.67 ± 0.28	7.90 ± 0.42	8.292±0.52

Results are shown as mean \pm SE

The last row indicate the control group data.

*=P<0.05, **=P<0.01 and ***=P<0.001.

Determination of serum lipids and glucose showed a significant increase ($p \le 0.01$) in cholesterol, triglyceride and glucose values in the cytoxan treated groups compared to control group (Table 6). However, administration of HDN in combination with cytoxan improved these values.

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Table 6: Effect of HDN on blood cholesterol, glucose and triglycerol of cytoxan intoxicated male mice.

Treatment	Dose		Cytoxan		Cytoxan	+ HDN (50 mg	g/kg bw)
period	mg/kg	Cholesterol	Glucose	Triglycerol	Cholesterol	Glucose	Triglycerol
periou	mg/kg	mg/dL	mg/dL	mg/dL	mg/dL	mg/dL	mg/dL
	50.0	159.55±0.79 [*]	72.86±1.02	97.29±1.84*	155.57±1.50	70.57±0.56	92.37±0.69
1 Week	100.0	161.54±0.79**	73.54±0.81*	99.73±1.66***	156.60±0.93	70.59±0.33	93.40±1.04
	200.0	163.54±0.53***	74.99±0.92**	100.82±1.47***	157.46±0.79	71.16±0.46	93.89±3.09
	50.0	$160.48\pm0.80^*$	73.84±1.1*	97.91±1.66**	156.40±1.12	70.81±0.34	92.50±0.82
2 Weeks	100.0	162.25±0.76**	74.99±0.92**	100.92±1.99***	157.51±0.96	71.35±0.58	93.74±0.94
	200.0	165.95±0.55***	75.53±0.92***	101.01±2.27***	158.46±0.71	71.85±0.90	94.11±1.08
	50.0	161.59±0.85***	74.15±0.98***	100.39±1.16****	157.57±0.96	71.81±0.35	93.66±1.01
	100.0	164.20±0.99***	76.2±0.48***	101.69±2.32***	158.98±0.88	72.26±1.06	95.90±4.01
3Weeks	200.0	167.07±0.96***	78.43±0.80***	102.73±1.93***	159.48±1.07	72.86±1.02	97.15±2.98

	91.02±1.1	70.54±0.50	15.42±1.5	0.0	Control
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Results are shown as mean \pm SE

The last row indicate the control group data.

*=P<0.05, **=P<0.01 and ***=P<0.001.

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The treatment with HDN in combination with cytoxan significantly (p < 0.05) lowered the degradation of membrane lipids than the cytoxan alone. Cytoxan induction provokes lipid bilayer repercussion by breaking down membrane phospholipids, therefore MDA, product of lipid peroxidation, acts as a marker for lipid bilayer damage. As the membrane damage progress, it results in the buildup of free radicals in normal animals which leads to greater membrane damage and inactivation or alteration of membrane bound enzymes. On treatment with rats, HDN decreased MDA level signifying attenuation in lipid peroxidation thereby proving its stabilizing power on membranes. Enough evidence has been garnered for HDN proving to be effective antioxidant in CPA mediated oxidative stress.

In conclusion, these findings substantiate the chemoprotective and antigenotoxic potential of HDN against toxicity induced by cytoxan in mice. It is clear that HDN may has antioxidative activity which reduced the oxidative stress, genotoxicity, and DNA damage induced by cytoxan in mice.

REFERENCES

- 1. K. Tiwari, Imbalance in antioxidant defense and human diseases: multiple approach of natural antioxidants therapy. Current Sciences, 2001; 81: 1179-1187.
- S. J. Hosseinimehr, M. Karami, Citrus extract modulates genotoxicity induced by cyclophosphamide in mice bone marrow cells. Journal of Pharmacy and Pharmacology, 2005; 57: 505-510.
- 3. A. Garg, S. Garg, J. D. Zaneveled, A. K. Singla; Chemistry and pharmacology of the citrus bioflavonoid hesperidin. Phytotherapy Research, 2001; 15: 655-669.
- 4. S. Khan, G.Jena; Sodium valproate, a histone deacetylase inhibitor ameliorates cyclophosphamide-induced genotoxicity and cytotoxicity in the colon of mice. J Basic Clin Physiol Pharmacol., Jan 2014; 27: 1-11.
- 5. M. J. Moore; Clinical pharmacokinetics of cyclophosphamide. Clin Pharmacokinet., 1991; 201: 194–208.
- 6. J. Gilbert, W. P. Petros.et.al., Pharmacokinetic interaction between ondanstetron and cyclophosphamide during high-dose chemotherapy for breast cancer. Cancer Chemother Pharmacol., 1998; 42: 497–503.
- 7. R. R.Chhipa, S. Singhet.al.;Doxycycline potentiates antitumor effect of cyclophosphamide in mice. Toxicol. Appl. Pharmacol., 2006; 202: 268–77.

- 8. E. Jantunen, M. Putkonenet.al.; Low-dose or intermediate-dose cyclophosphamide plus granulocyte colony-stimulating factor for progenitor cell mobilization in patients with multiple myeloma. Bone Marrow Transplant, 2003; 31: 347-51.
- 9. M. Karanth, S. Chakrabarti, R. A. Lovellet al.; A randomised study comparing peripheral blood progenitor mobilisation using intermediate-dose cyclophosphamide Plus lenograstim with lenograstim alone. Bone Marrow Transplant, 2004; 34: 399-403.
- 10. D. Anderson, J. B. Bishop et.al.; Cyclophosphamide: review of its mutagenicity fora assessment of potential germ cell risks, Mutat. Res., 1995; 330: 115–81.
- 11. W. F. Benedict, A. Banerjee, N. Venkatesan; Cyclophosphamide induced oncogenic transformation, chromosomal breakage and sister chromatid exchange following gmicrosomal activation. Cancer Res., 1978; 38: 2922-24.
- 12. U. Claussen, W. Hellnann, G. Pache; The embryotoxicity of the cyclophosphamide metabolite acrolein in rabbits, tested in vivo by i.v. injection and by theyolk-sac method. Arzneim.Forsch., 1980; 30: 2080-83.
- 13. Baibinder, C. Reich et al.; Relative mutagenicity of some urinary metabolites of the antitumor drug cyclophosphamide. Cancer Res., 1981; 41: 2967-72.
- 14. S. H. Robison, M. R. Odioet.al.; Assessment of the in vivo genotoxicity of 2-hydroxy-methoxyben zophenone, Environ. Mol. Mutagen., 1994; 23: 312–17.
- 15. A. Mishra, V. Gupta, H. Nagar, V. Shrivastav; Protective Effect of Murrayakoenigii (Curry Leaf) Leaves Extract Against Genotoxicity Induced by Cyclophosphamide in Mouse Bone Marrow Cells. Global Veterinaria, 2013; 10(2): 128-133.
- 16. T. Doherty, J. E. Hayes, J. Molloy, C.Wood, M. R. O'Donovan; Bone marrow micronucleus frequencies in the rat after oral administration of cyclophosphamide, hexamethylphosphoramide or gemifloxacin for 2 and 28 days. Toxicol Res., 2013; 2: 321-327.
- 17. S.Sharma, G. Sharma, A. Mehta; Antimutagenic protection of Ficus benghalensis extract against cyclophosphamide induced genotoxicity in rat bone marrow. Asian J Pharm Clin Res, 2012; 5(1): 84-86.
- 18. R.Edenharder, J. Frangart, M. Hager, P. Hofmann, R.Rauscher; Protective effects of fruits and vegetables against in vivo clastogenicity of cyclosphosphamide or benzo[a] pyrene in mice. Food and Chemical Toxicology, 1998; 36(8): 637-45.
- 19. A. Ahmadi, S. J. Hosseinimehr, A. Naghshvar, H. Ebrahim, M. Ghahremani; Chemoprotective Effects of Hesperidin Against Genotoxicity Induced by Cyclophosphamide in Mice Bone Marrow Cells. Arch Pharm Res, 2008; 31(6): 794-797.

- 20. P. Mittal, P. A. Patil, S. S. Torgal; Screening of codeine, dextromethorphan and dextropropoxyphene for their genotoxicity in swiss albino mice. Indian J. Med. Res., 2009; 129: 676-680.
- 21. L. Recio, C. Hobbs, W. Caspary, K. L. Witt; Dose-Response Assessment of Four Genotoxic Chemicals in a Combined Mouse and Rat Micronucleus and Comet Assay Protocol. J Toxicol Sci., 2010; 35(2): 149–162.
- 22. H. Zhao, Y. Xuan, L. Defang, C. Hongmei, J. Jiangtao, W. Zhiping, Z. Qiusheng; Isoliquiritigen Enhances the Antitumour Activity and Decreases the Genotoxic Effect of Cyclophosphamide. Molecules, 2013, 18(8): 8786-8798.
- 23. R. D Short, K. S. Rao, J. E. Gibson; The in vivo biosynthesis of DNA, RNA, and proteins by mouse embryos after a teratogenic dose of Cyclophosphamide. Teratology, 1972; 6: 129-138.
- 24. N. Singh, M. Nigamet.al.; Caspase Mediated Enhanced Apoptotic Action of Cyclophosphamide and Resveratrol-Treated MCF-7 Cells, Journal of pharmacological sciences, 2009; 109: 473-85.
- 25. S. I. R. Franke, D. Pra, B. Erdtmann, J. A. P. Henriques, J. da Silva; Influence of orange juice over the genotoxicity induced by alkylating agents: an in vivo analysis. Mutagenesis; 2005; 20(4): 279-283.
- 26. S. J. Hosseinimehre, A. Ahmadia, D. Beikib, E. Habibia, A. Mahmoudzadeh; Protective effects of hesperidin against genotoxicity induced by99mTc-MIBI in human cultured lymphocyte cells. Nuclear Medicine and Biology, 2009; 36: 863–867.
- 27. N. Nithya, K. Chandrakumar, V. Ganesan, S. Senthilkumar; Efficacy of Momordica charantia in attenuating hepatic abnormalities in cyclophosphamide intoxicated rats. J. Pharmacol. Toxicol., 2012; 7(1): 38-45.
- 28. A. Merwid-Ląd, M. Trocha, E. Chlebda, E. Sozański, J. Magdalan, D. Ksiądzyna, M. Pieśniewska, A. Szeląg; The Effects of Morin, a Naturally Occurring Flavonoid, on Cyclophosphamide-Induced Toxicity in Rats. Adv Clin Exp Med, 2011; 20(6): 683–690.
- 29. S. O. Abarikwu, C. A. Otuechere, M. Ekor, K. Monwuba, D. Osobu; Rutin Ameliorates cyclophosphamide-induced reproductive toxicity in male rats. Toxicol. Inter., 2012; 19(2): 207-214.
- 30. M. Shokrzadeh, A. Chabra, F. Naghshvar, A. Ahmadi; The Mitigating Effect of Citrullus colocynthis (L.) Fruit Extract against Genotoxicity Induced by

- Cyclophosphamide in Mice Bone Marrow Cells. The Scientific World Journal Volume 2013, Article ID 980480, 8 pages (2013).
- 31. T. SaiSampath, M. Kanaka Durga, Bh. Saranya, K. Kalyani; Review on plant derived natural products and their analogues with chemoprotective activity against genotoxicity of cyclophosphamide. International Journal of Pharma and Bio Sciences, 2011; 2(3): 375-386.
- 32. S. Kumar, N. Dhankhar, V. Kar, M. Shrivastava, S. Shrivastava; Myocardial Injury Provoked by Cyclophosphamide, Protective Aspect of Hesperid in in Rats. International Journal of Research in Pharmaceutical and Biomedical Sciences, 2013; 2(3): 1288-1296.
- 33. N. R. V. Dragano, V. P. de Venancio, F. B. A. Paula, F. Della Lucia, Fonseca, M. J. O. and Azevedo, L.: Influence of Marolo (Annonacrassiflora Mart.) pulp intake on the modulation of mutagenic/antimutagenic processes and its action on oxidative stress in vivo. Plant Foods Hum Nutr, 2010; 65: 319-325.
- 34. H. Valette, F. Dolle, M. Bottlaender, F. Hinnen, D. Marzin; Fluro-A-85380 demonstrated no mutagenic properties in vivo rat micronucleus and Amestests. Nuclear Medicine and Biology, 29: 849-853.
- 35. U. J. A. D'Souza, A. Zain, S. Raju; Genotoxic and cytotoxic effects bonemarrow of rats exposed to low dose ofpaquat via the dermal route. Mutation Research, 2002; 581: 187-190.
- 36. Porcher, M. C. Malinge, C. Picat, B. Grandchamp; A simplified method for determination of specific DNA or RNA copy number using quantitative PCR and an automatic DNA sequencer. Biotechniques, 1992; 13(1): 106-14.
- 37. H. Ohkawa, N. Ohishi, K. Yagi; Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Analytical Biochemistry, 1979; 95(2): 351–358.
- 38. Nishikimi, N. Appaji, K. Yagi; The occurrenceof superoxide anion in the reaction of reduced phenazinemethosulfate and molecular oxygen. Biochem Biophys Res Commun; 1972; 46(2): 849-54.
- 39. J. B. Henry; Clinical diagnosis and management by laboratory methods,19th ed. Philadelphia: WB Saunders; USA (1996).
- 40. W. Richmond, Preparation and properties of the cholesterol oxidase from nacordia sp. and its application to the enzymatic assay of total cholesterol in serum. Clin. Chem., 1973; 19: 1350-1359.

- 41. P.Fossati, L.Prencipe; Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. Clin. Chem., 1982; 28(10): 2077-80.
- 42. M. Ripamonti, A. Mosca, E. Rovida, M. Luzzana, L. Luzi, F. Ceriotti, F. Cottini, L. Rossi-Bernardi;): Urea, Creatinine and Glucose determined in plasma and whole blood by a differential pH technique. Clin.Chem., 1984; 30(4): 556-559.
- 43. T. Peters; Protein colorimetric method. Clin. Chem., 1968; 14: 1149-1159.
- 44. M. Hayashi, R. R. Tice, J. T. Macgregor, D. Anderson, D. H. Blakey; In vivo rodent erythrocyte micronucleus assay. Mutat. Res., 1994; 312: 293-304.
- 45. K. L. Witt, E. Livanos, E. G. Kissling, D. K. Tarous, W. Caspary, R. R. Tice, Recio; Comparison of flow cytometry and microscopy based methods for measuring micronucleated reticulocyte frequencies in rodents treated with nongenotoxic and genotoxic chemicals. Mutat. Res., 2008; 649: 101-113.
- 46. P. Abraham, E. Sugumar; Increased glutathione levels and activity of PON1 (phenyl acetate esterase) in the liver of rats after a single dose of cyclophosphamide a defense mechanism. Exp Toxicol Pathol., 2008; 59(5): 301-6.
- 47. D. Wills; Effects of lipid peroxidation on membrane –bound enzymes of the Endoplasmic reticulum. Biochem J., 1971; 123: 983- 991.
- 48. L. G. Menon, S. Picinich, R. Koneru, H.Gao, S. Y. Lin, M. Koneru, P. Mayer Kuckuk, J. Glod, D. Banerjee; Differential gene expression associated with migration of mesenchymal stem cells to conditioned medium from tumor cells or bone marrow cells. Stem Cells, 2007; 25(2): 520-528.