

WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 5.045

Volume 4, Issue 4, 701-708.

Research Article

ISSN 2277-7105

701

SCREENING PROCEDURE FOR SELECTING FUNGI WITH POTENTIAL FOR USE IN THE BIOREMEDIATION OF CONTAMINATED SOIL

Merlyn Stephen* and A. Panneerselvam

PG & Research Department of Botany and Microbiology, A.V.V.M.Sri pushpam college (autonomous) Poondi, Thanjavur Dt.Tamil Nadu, India.

Article Received on 27 Jan 2015,

Revised on 19 Feb 2015, Accepted on 09 Mar 2015

*Correspondence for Author

Merlyn Stephen

PG & Research
Department of Botany and
Microbiology, A. V. V.
M. Sri pushpam college
(autonomous) Poondi,

Thanjavur Dt. Tamil Nadu,

India.

ABSTRACT

In the present investigation fungi on the hydrocarbon polluted soils were screened. These are conducted by the enumeration of the fungal population and the identification. The following fungal organisms such as *Aspergillus, Penicillium, Fusarium, Cladosporium, Mucor, Curvularia, Helminthosporium*, and *Trichoderma Species* were sereened and identified. The growth of fungal diversity was higher due to more carbon concentration in the hydrocarbon polluted sites and the dominant species present in all stations are *Aspergillus* and *penicillium Species*. The study focuses its attention on the survey of fungi from the zone of hydrocarbon polluted area and their potential ability to bring about degradation of hydrocarbons.

KEYWORDS: Soil, Pollution, Hydrocarbon, Biodegradation, Fungi.

INTRODUCTION

Hydrocarbons are causing wide spread pollution in both the aquatic and terrestrial environment. The petroleum industry is responsible for the generation of large amounts of organic residues, as well as for the pollution of soils, rivers and seas. One of the best approaches to restoring contaminated environments is to make use of the physiological potential of microorganisms able to degrade the pollutants in a bioremediation process. It is an attractive approach to cleaning up hydrocarbons because it is simple to maintain, applicable over large areas, cost-effective and leads to the complete destruction of the contaminant (Bento et al., 2005). Oil spillage is the accidental discharge or pouring of crude oil into the environment which involves the contamination of the environment with liquid

hydrocarbon. These spills endanger public health, drinking water and natural resources and disrupt the economy (Gesinde, *et al.*, 2008).

Strategies for controlling environmental contamination by petroleum and its derivatives have been the subject of various studies over the past three decades. When a spillage occurs the first action is to remove the oily phase by mechanical or by physical-chemical means through the application of surfactants to disperse the layer of oil. Bioremediation is an alternative that has been used to eliminate or minimise the effects of pollutants by using microorganisms which have biodegradation potential (Atlas, 1995).

In recent times, an increasing amount of microbiological research has been devoted to bioremediation of oil-contaminated sites using various microbial species Numerous microorganisms are known for their ability to degrade hydrocarbons. The biodegradation capabilities of bacteria have been recognized, but fungi have been the subject of recent research (Colombo *et al.*, 1996; Krivobok *et al.*, 1998; Salicis *et al.*, 1999; García *et al.*, 2000; Garon *et al.*, 2000; Chaillan *et al.*, 2004; Santos and Linardi, 2004; Potin *et al.*, 2004), due to their ability to synthesize relatively unspecific enzymes involved in cellulose and lignin degradation, which are capable of degrading high molecular weight, complex or more recalcitrant compounds, including aromatic structures. The full potential of biodegradation by filamentous fungi for bioremediation purposes has not been fully investigated. The use of filamentous fungi isolated from contaminated soil may offer advantages for several reasons. Owing to their ability to extend through the soil by hyphal elongation, fungi can access xenobiotics. In addition fungi are capable of growing under stressful environmental conditions.

The biological treatment are cheaper than chemical and physical ones. The degradation of crude oil in soil matrix through microorganisms are able to transform petroleum hydrocarbons in less toxic substances Therefore, the main goal of this work was to evaluate the colony growth rates of filamentous fungi isolated from contaminated soil area of Thanjavur District, using different petroleum hydrocarbons or derivatives as the only carbon source, with a view to selecting strains for future employment in bioaugmentation schemes.

MATERIALS AND METHODS

Sources of soil sample: The five oil contaminated soil samples used for the isolation were from five different stations located at the environment. The five different stations were 1.

Automobile shop at Orathanadu, Thanjavur. 2. Petrol bunk at Mariyamman Kovil, Thanjavur 3. Heavy Vehicle Work shop centre at Karanthai, Thanjavur 4. Heavy vechicle workshop at New bus stand, Thanjavur. 5. Heavy vehicles workshop centre at Thiruchirapalli Samples from each station were collected randomly from different locations just 1cm below the soil surface and transported to the laboratory in white plastic bags and kept in a refrigerator (in order to keep the organisms viable and free from any contaminant) before analysis.

Soil dilution method: 1gm of soil sample was suspended in 10ml of double distilled water to make soil suspensions and 1ml of soil suspension of each concentration were added to sterile Petri dishes (triplicate of each dilution) containing 15 ml of sterile Potato Dextrose Agar and Czapek Dox Agar. One percent streptomycin solution was added to the medium before pouring into petriplates for preventing bacterial growth. The Petri dishes were then incubated at 28±2 °C in dark. The plates were observed everyday up to three days.

Physico-chemical analysis of soil: The collected soil was characterized for its physicochemical properties. The physico -chemical parameters were measured by standard methods. Physical and chemical parameters of soil such as pH, salinity, organic carbon, nitrogen, phosphorous and potassium were analyzed. Moisture content was estimated by finding the weight difference of known quantity of soil before and after drying in a hot air oven at 60°C for 6 hours. Soil samples after removing the debris were suspended in distilled water (1:2 w/v) and allowed to settle down the sand particles. The pH of the suspension was read using pH meter (Systronics, India), to find out the soil pH. Electrical conductivity (Jackson, 1973), Cation exchange capacity (CEC) of the soil was determined by using 1 N Ammonium acetate solution as described by (Jackson, 1973). Organic carbon content was determined by adopting chromic acid wet digestion method as described by Walkley and Black, 1934, Available nitrogen (Subbiah and Asija, 1956) Available phosphorus by Brayl method as described by Bray and Kutz, 1945. Available potassium (Standford and English, 1949) Calcium (Jackson, 1973) Available micronutrients such as Zn, Cu and Mn (Lindsay and Norwell, 1978). Other nutrients such as magnesium, sodium and available iron were analysed following the method of (Barnes 1959 and Muthuvel and Udayasoorian, 1999). The physico chemical parameters of the soil samples were analyzed at Soil Testing Laboratory Tiruchirappalli, Tamilnadu, India.

Methods of microscopic examination: For light microscope the optical equipment used were dissecting microscope, research microscope (10x and 15x eye pieces and 10 x to 100x

objectives), equipment for microphotography, camera Lucida ocular, stage micrometers.

Identification of the soil fungi: Fungal morphology were studied macroscopically by observing colony features (Colour and Texture) and microscopically by staining with lacto phenol cotton blue and observed under compound microscope for the conidia, conidiophores and arrangement of spores .The fungi were identified with the help of standars manuals. Manual of soil fungi (Gillman,1957). Hyphomycetes (Subramaniyan,1971) A manual of penecillia (Raper and Thom 1949) the genus *Aspergillus* (Raper and Fennell,1965)

RESULTS

The soil samples examined for isolating fungi from petroleum hydrocarbon contaminated site. Totally 15 strains were isolated from the hydrocarbon pollued soil. They were identified as *Aspergillus, Penicillium, Fusarium, Cladosporium, Mucor, Curvularia, Helminthosporium*, and *Trichoderma Species*, Among them *Aspergillus and Penicillium sp* are the most commonly encountered genera of hydrocarbon degraders in oil contaminated stations, in agreement with the present results.

Table :1 List of fungi isolated from five different stations

ISOLATE	STATION	STATION	STATION	STATION	STATION
IDENTIFICATION	I	II	III	IV	\mathbf{V}
Aspergillus fumigatus	+	+	+	+	+
A. flavus	+	+	+	+	+
A.niger	+	+	+	+	+
A. versicolar	+	-	+	+	+
A. japonicus	+	-	+	+	+
A.nives	+	-	+	-	-
A.terreus	+	+	+	+	+
Penicillium lanosum	+	-	+	-	-
P.chrysogenum	+	+	+	+	+
P.lanosum	+	+	+	+	+
P.corylophilum	+	-	+	+	+
Trichoderma viride	+	-	-	-	-
Cladosporium herbarum	_	+	-	+	+
Mucor racemosus	-	+	-	+	-
Helminthosporium solani	+	+	-	-	-

- (+) Indicates the presence of the microbes
- (-) Indicates the absence of the microbes

Table 2 Physico-chemical properties of hydrocarbon contaminated soils

CI NO	SOIL PARAMETERS	STATIONS					
SI NO		I	II	III	IV	V	
1	Texture Sandy Loam	7.86	7.56	7.43	7.48	7.91	
2	Temperature(°C)	50	30	40	40	30	
3	рН	8.5	8.0	7.0	7.5	7.0	
4	Total nitrogen (kg/hectare)	112.3	106.6	103.8	105.2	97.9	
5	Totalphosphorous (kg/hectare)	3.75	4.23	4.72	4.50	4.01	
6	Total potassium (kg/hectare)	118	121	124	117	125	
7	Zinc (ppm)	0.90	0.79	0.82	0.96	1.02	
8	Copper (ppm)	0.48	0.50	0.42	0.48	0.52	
9	Iron (ppm)	4.87	4.58	4.63	4.56	4.62	
10	Manganese(ppm)	2.16	2.58	24.6	27.1	28.6	
11	Electricalconductivity(Ec)	0.48	0.52	0.26	0.42	0.27	

DISCUSSION

Studies on the isolation of filamentous fungi in environments containing oil or its sub products found a very similar diversity of genera to that found in our study, such as Aspergillus and Penicillium, Recently, it was recorded that the genera of fungi such as Penicillium, Aspergillus, Fusarium, Rhizopus associated with petroleum hydrocarbon contaminated soil. In their studies they isolated Penicillium and Aspergillus from hydrocarbon contaminated soil and identified as hydrocarbon degrading fungi along with Trichoderma, and Rhizopus sp. The similar results of our study were also obtained by Obire et al.,2009; in their studies on effect of different concentrations of crude oil on fungal populations of soil. The fungal isolates obtained in their study were mainly Aspergillus species, while others were Penicillum, Rhizopus and Rhodotorula species which were all able to utilize hydrocarbon as carbon source from contaminated soil, which were able to degrade crude oil.

In the present investigation Aspergillus and Penicillium species were present in dominant numbers. Our finding coincides with the work of Elisane et al.,2008 who also isolated four strains from the contaminated soil. They were identified as Aspergillus are the most commonly encounter hydrocarbon degraders in oil contaminated tropical soils, which are in agreement with the present work fungi from total hydrocarbon contaminated soil and identified by microscopy as Penicillium, Aspergillus The different result from our findings

were obtained who also isolated many fungal species that were able to degrade polycyclic aromatic hydrocarbons. The species isolated were *Coniothyrium fuckelii*, *Gliocladium virens Phialophora hoffmannii*, *Scopulariopsis brumptii Trichoderma harzianum* along with genera were similar to our finding. The growth rate of each fungus shows that *Rhizopus sp.* had the highest growth diameter in low petro contaminated PDA media culture and *Aspergillus and penicillium species* had the highest growth diameter in high petro contaminated PDA medium.

CONCLUSIONS

Biological recovery processes of soils contaminated by hydrocarbons and their derivatives have been based on the stimulation of native microorganisms and in some cases, on the increase of the microbial population, through incorporation of native/exogenous organisms. These microorganisms do not seem to be competitive in comparison with the native microbial population, which is already adapted to the environment. One alternative would be the isolation of species from contaminated soil and their posterior growth and reintroduction into the same system. The problem is that conventional isolation methods are only able to extract a small part of viable microorganisms from the environment, thus limiting the achievement of species of interest. This fact will lead to a future development of better studies with these fungi, as well as with those that grow in both conditions, for specific purposes of use in biodegradation.

In the present study the *Penicillium and Aspergillus sp* was the most frequent genus found in uncontaminated soil. However, it was not isolated from contaminated soil. The existence of species that grow with or without the presence of oil allow inferences on their use as an contamination indicator or on how these hydrocarbons are degraded, having potential for the treatment of environments.

ACKNOWLEDGEMENTS

The authors gratefully thank the Secretary and correspondent, Principal and Dean Faculty of science for their support for providing laboratory and working facilities.

REFERENCE

- 1. Atlas, R. M., Bioremediation of Petroleum Pollutants. Internet. Biodeterior. and Biodegrad, 1995; 317- 327.
- 2. Barnes, H., 1959. Apparatus and methods of Oceanography, Part I Chemical, Allen and

- Unwin Ltd., London.
- 3. Bento FM, Camargo FAO, Okeke BC, Frankenberger WT. Comparative bioremediation of soils contaminated with diesel oil by natural attenuation, biostimulation and bioaugmentation. Bioresour. Technol. 2005; 96: 1049-1055.
- 4. Bray, R.H., and Kutz, L.T., Determination of total organic and available phosphorus in soils. Soil Sci., 1945; 59: 39-45.
- 5. Colombo JC, Cabello M, Arambarri AM. Biodegradation of aliphatic and aromatic hydrocarbons by natural soil microflora and pure cultures of imperfect fungi. Environ. Pollut. 1996; 94: 355- 362.
- 6. Chaillan F, Fleche AL, Bury E, Phantavong Y, Crimount P, Saliot A and Oudot J. identification and biodegradation potential of tropical aerobic hydrocarbon-degrading microorganism research in Microbiology, 2004; 155: 587-595.
- Elisane ODS, Celia FCDR, Catia TDP, Ana VLS, Janaina FDMB, Susana JK and Carlos AVB., Prescreening of filamentous fungi isolated from a contaminated site in Southern Brazil for bioaugmentation purposes, African Journal of Biotechnology, 2008; 7: 1314-1317.
- 8. García IG, Peña PRJ, Venceslada JLB, Martín AM, Santos MAM, Gómez ER. Removal of phenol compounds from olive mill wastewater using Phanerochaete chrysosporium, Aspergillus niger, Aspergillus terreus and Geotrichum candidum. Process Biochem. 2000; 35: 751-758.
- 9. Garon D, Krivobok S, Seigle-Murandi F. Fungal degradation, 2000; 40: 91-97.
- 10. Gesinde, A.F., E.B.Agbo, M.O. Agho and E.F.C.Dike, Bioremediation of some Nigeria and Arabian crude oils by fungal isolates. Int. J. Pure Applied Sci., 2008; 2:37-44.
- 11. Gillman, J.C., A manual of Soil Fungi Revised 2nd edition Oxford and IBH publishing company (Indian reprint) Calcutta, Bombay, New Delhi: 1957, pp. 436.
- 12. Jackson, M.L., Soil Chemical Analysis. New Delhi: Prentice Hall of India Private Limited:1973, pp. 180-189.
- 13. Krivobok S, Miriouchnike E, Seigle-Murandi F, Benoit-Guyod JL Biodegradation of anthracene by soil fungi. Environ. Pollut. 1998; 37: 523
- 14. Lindsay, W.C., and Norwell, A., Development of a DTPA soil test for zinc, iron, manganese and copper. Proc. Soil Sci. Sol. Am., 1978; 42:421-428.
- 15. Muthuvel, P. and Udayasoorian, C., 1999. Soil, plant, water and agrochemical analysis,
- 16. O. Obire, E. C. Anyanwu and R. N. Okigbo, "Saprophytic and Crude Oil Degrading Fungi from Cow Dung and Poultry Droppings as Bioremediating Agents," Journal of

- Agricultural Science and Technology, 2009; 4: 81-89.
- 17. Potin O, Rafin C, Veignie E Bioremediation of an aged polycyclic aromatic hydrocarbons (PAHs) contaminated soil by filamentous fungi isolated from the soil. Int. Biodeterior., Biodegradation. 2004; 54: 45-52.
- 18. Raper K B & Fennell D I, The genus Aspergillus, (The Williams and Wilkins Co., Baltimore), 1965; 686.
- 19. Raper, K.B., and Thom, C.(1949). A manual of Penicillia. Williams and Wilkins Co., Baltimore, Md., U.S.A.
- 20. Santos EO, Rosa CFC, Passos CT, Sanzo AVL, Burkert JFM, Kalil SJ, Burkert CAV. 2008. Pre-screening of filamentous fungi isolated from a contaminated site in Southern Brazil for bioaugmentation purposes. African Journal of Biotechnology, 2008; 7(9):1314-1317.
- 21. Salicis F, Krivobok S, Jack M, Benoit-Guyod JL Biodegradation of soil fungi. Environ. Pollut. 1999; 38: 3031-3039.
- 22. Standfold, S., and English, L., Use of flame photometer in rapid soil test K and Ca. Agron. J., 1949; 41: 446-447.
- 23. Subbiah, B.V., and Asija, G.L., A rapid method for estimation of available nitrogen in soil. Curr. Sci., 1956; 25: 258-260.
- 24. Subramanian, C.V., 1971. Hypomycetes: An account of Indian species, Indian Counc. Agri. Res., New Delhi.
- 25. Walkley, A. A critical examination of a rapid method for determining organic carbon m soils: Effect of variations in digestion conditions and of inorganic soil constituents. Soil Sci. 1947; 63:251-263.