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ASSOCIATION OF MATERNAL OBESITY AT BIRTH WITH INCREASED OFFSPRING BODY SIZE AND SERUM LEPTIN

*Neveen Helmy Abou El-Soud¹, Nahla Abdel Moniem Barakat², Mai Mahmoud Youssef³, Manal Abdel Moniem Mohsen³, Gamila Mohamed El-Saeed⁴

¹Complementary Medicine Department; ²Pediatric Department; ³ Child Health Department; ⁴Biochemistry Department. National Research Centre- El-Behouth Street, Dokki, 12311 Cairo, Egypt.

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*Correspondence
For Author
Prof. Dr. Neveen Helmy
Abou El-Soud
Complementary Medicine
Department, Medical
Researches Division,
National Research Center,
El-Behouth Street,Dokki,
12311 Cairo, Egypt.

ABSTRACT

Background: Maternal obesity and weight gain during pregnancy are risk factors for child obesity. There is a potential role of leptin in the regulation of body weight, energy expenditure and therefore fetal growth and development. The aim of this study was to examine the association of maternal obesity at birth with offspring body size and serum leptin. **Subjects and methods:** Two hundred mothers at delivery with their term healthy human newborns were enrolled in this study. One hundred mothers of them were obese (body mass index $(BMI) \geq 30 \text{ kg/m}^2$) and one hundred act as control (BMI between 20 - 24.9 kg/m²). Neonatal weight, length, head circumference and ponderal index were determined. Leptin concentrations in venous cord blood were measured using specific enzyme –linked immunosorbent assay (ELISA) method. **Results:** Neonates of obese mothers had

significantly higher levels of leptin $(21.17\pm16.7 \text{ vs } 11.33\pm10.55 \text{ ng/ml}, p < 0.001)$, birth weight $(3.64\pm0.32 \text{ vs } 3.2\pm0.67 \text{ kg}, p < 0.05)$, ponderal index $(3.24\pm0.38 \text{ vs } 2.77\pm0.65 \text{ gm/cm}^3)$, p < 0.05) when compared to non-obese group. Cord serum leptin was found to correlate with maternal BMI (r=0.46, p < 0.05) and neonatal body measurements; weight (r=0.50, p < 0.01), length (r=0.59, p < 0.001), head circumference (r=0.56, p < 0.01) and ponderal index (r=0.40, p < 0.05). **Conclusion:** Maternal obesity at delivery is associated with increased neonatal serum leptin and body size with high risk of children obesity later in life.

KEYWARDS: Maternal obesity; neonatal birth size; leptin.

INTRODUCTION

Obesity and overweight in children is associated with a wide spectrum of adverse conditions and can negatively affect every organ in the body. Consequences can be insulin resistance, hypertension, dyslipidemia and fatty liver disease.^[1] Recent evidence has linked childhood obesity with liver cancer in adulthood.^[2]

In addition, overweight and obese children might experience psychological and social problems.^[3] Obesity in childhood usually continues into adulthood.^[4] A number of epidemiological studies have shown a link between birth weight and BMI in childhood and adulthood,^[5,6] and thus potentially creating a life-long condition.

Maternal obesity and gestational weight gain, resulting in over-nutrition of the fetus, are major contributors to obesity and metabolic disturbances in the offspring.^[7,8]

Leptin is an adipocyte-derived hormone which is involved in the regulation of food intake and body weight. ^[9] Through stimulation of sympathetic nerve activity, it leads to a significant increase in energy expenditure. ^[10] The biologic actions of leptin are thought to be mediated through the activation of leptin receptor in the hypothalamus. ^[11] Numerous studies have found that plasma leptin concentrations are elevated in several models of animal and human obesity in proportion to the degree of adiposity. ^[12,13]

This study aimed to investigate the association of maternal obesity at birth with offspring body size and serum leptin.

SUBJECTS AND METHODS

This study was a case control study conducted from Mars 2013 to September 2014. It was observational cross sectional one consisting of three components: a maternal body mass index before delivery, umbilical cord blood collection at the time of delivery to measure serum leptin, and neonatal anthropometric measurements at birth. The study protocol was approved by the ethical committee of the National Research Centre and Obstetric Department of El-Azhar University. Participants gave informed, written consent for themselves and their newborns to enrol into this study.

SUBJECTS

Two hundred healthy pregnant women carrying singleton pregnancies, aged 18-40 years between week 37 and 40 of gestation were recruited in this study. They were attending the Obstetric Department, El-Azhar University for labour. None of them was taking any medication, except for vitamins and iron supplementation.

Women were excluded from this study if they had preeclampsia, chronic hypertension and pregravid type 1 or type 2 diabetes. At delivery, participants were excluded if delivery was caesarean, newborns were born preterm (< 37 weeks of gestation), small for gestational age (SGA) or with severe birth asphyxia (5-min Apgar score <6 or cord blood pH < 7), congenital metabolic or endocrine disease, with major malformations, CNS deficit, congenital infections, or disorders known to affect growth.

The studied women and their offspring were divided into two groups according to maternal BMI: obese group (BMI \geq 30 kg/m²); control group with normal body mass index (BMI: 20-24.9 kg/m²).

METHODS

All pregnant women were evaluated by history, clinical examination, and routine investigations. Newborns gestational age, sex and mode of delivery were recorded. Apgar scores at 1,5 minutes were recorded, each determined as the sum of scores (0,1 or 2) related to five neonatal vital signs (heart rate, respiratory rate, muscle tone, excitability and skin colour).

Measurements of anthropometric parameters

Maternal weight and height were measured before delivery. Maternal body mass index was assessed as weight in kg/height in meters 2 . Neonatal weight , height and head circumference were determined. Ponderal index (in kg/m3) was calculated as body weight in grams/ length in centimetres X 100. Anthropometric data were assessed as originally described by Cameron. $^{[14]}$

Biochemical analysis

Umbilical cord blood was collected at delivery and cord serum was separated by centrifugation and stored at -20 °C until analysis. Serum leptin was determined using specific enzyme -linked immunosorbent assay (ELISA) method using commercial kit

(Medizyme leptin from Medipan Diagnostica, Germany.) according to manufacture instructions. For each sample $100~\mu l$ of serum per tube was used for duplicate. The limit of detection of lipid assay was 0.45~ng/ml. The intra-assay and interassay coefficient of variation were 8% and 10.8% respectively.

Statistical analysis

Data was analysed using SPSS version 17 and expressed as mean \pm SD. Data was divided into two groups according to maternal BMI with the cutoff point 30 kg/m². Data of the two groups were compared using t-test for independent variables. When dividing the offspring data according to sex no significant differences were detected. Pearson correlation coefficient was used to determine the relation between continuous variables. Values of p <0.05 were considered significant.

RESULTS

Mothers mean age was 29.1 ± 5.53 years with range 19-40 years and of weight 65.63 ± 8.7 kg with range 60-86 kg. Their offspring mean birth weight was 3.39 ± 0.49 kg with range 2.6-4.0 kg and gestational age was 38.6 ± 1.29 weeks with range 37-40 weeks.

The mean cord serum leptin was 16.17 ± 12.4 ng/ml with rage 1-50 ng/ml. There was no leptin sex differences (16.9 ± 10.2 ng/ml for males vs 18.71 ± 11.4 ng/ml for females). When comparing the two groups based on maternal BMI at delivery; obese (BMI $\geq 30 \text{ kg/m}^2$) and control (BMI: $20\text{-}24.9 \text{ kg/m}^2$), it was found that neonates of obese mothers had significantly higher levels of leptin ($21.17 \pm 16.7 \text{ vs } 11.33 \pm 10.55 \text{ ng/ml}$, p < 0.001), neonatal birth weight ($3.64 \pm 0.32 \text{ vs } 3.2 \pm 0.67 \text{ kg}$, p < 0.05), ponderal index ($3.24 \pm 0.38 \text{ vs } 2.77 \pm 0.65 \text{ gm/cm}^3$, p < 0.05) (Table 1).

Table 1. Characteristics of study participants

Mother data	Obese group (n=100) BMI \geq 30 kg/m ²	Control group (n=100) BMI : (20- 24.9 kg/m ²)	P value
Age (years)	28.73±3.69	28.53±6.92	NS
Parity	2.07 ± 0.88	2.46±1.40	NS
Height (meter)	158.66± 4.08	159.26±5.31	NS
Weight (kg)	80.13± 4.36	68.66±4.8	< 0.001
BMI (Kg/m ²)	32.05±3.21	26.76±2.4	< 0.01
Neonatal data			

G.A (weeks)	38.66±1.04	38.93±1.28	NS	
Weight (Kg)	3.64±0.32	3.20±0.52	< 0.05	
Length (cm)	49.00±1.46	48.60±0.82	NS	
H.C (cm)	33.86±0.91	34.40±1.18	NS	
P.I (cm)	3.24±0.38	2.77±0.67	< 0.05	
Leptin (ng/ml)	21.17±16.70	11.33±10.55	< 0.001	
H.C:Head circumference ; P.I :Ponderal index; NS: Non- significant				

Cord serum leptin was found to correlate with birth weight (r = 0.50, p < 0.01), length (r = 0.59, p < 0.001), head circumference (r = 0.56, p < 0.01), ponderal index (r = 0.40, p < 0.05) and maternal BMI (r = 0.46, p < 0.05) (Table 2).

Table 2. Correlation of cord serum leptin with maternal body mass index and neonatal anthropometric measurements.

	Leptin		
Variable	r	p	
Maternal BMI	0.46	< 0.05	
Weight	0.50	< 0.01	
Length	0.59	< 0.001	
H.C	0.56	< 0.01	
P.I	0.40	< 0.05	

H.C:Head circumference; P.I:Ponderal index; r: Pearson correlation coefficient.

DISCUSSION

Increasing overweight influence the health of pregnant women, and also possess critical risk complications to maternal and Feto-Neonatal healthcare.^[15]

The impact of maternal obesity on the fetus has been investigated in several populations, and a range of adverse outcomes such as: Large for Gestational Age (LGA, birth weight > 90th percentile of gestational age- and sex-specific references) and macrosomia (birth weight \ge 4500g), shoulder dystocia , birth defects , preterm delivery , stillbirth and early neonatal death have consistently been reported. [16,17,18,19]

Results from the large Hyperglycemia and Adverse Pregnancy Outcome (HAPO) study has provided strong evidence of an association between maternal obesity and birth weight. A recent meta-analysis estimated that maternal obesity increases the risk of LGA, high birth weight (> 4000g) and macrosomia, with odds ratios of 2.08, 2.00 and 3.06, respectively.

Our data show that offspring of obese mothers have significantly higher body weight and body size. Interestingly, it was reported that the increased birth weight in offspring of obese mothers seems to be the result of increased fat mass, rather than lean mass.^[22,23] suggesting that the in utero metabolic environment affects primarily growth of fat mass, not lean mass.^[24]

Leptin, the product of the obesity (ob) gene, is a hormone of 16 kDa comprising 167 amino acids. ^[25] The central source of leptin is the adipose tissue (white and brown), both maternal and fetal adipose tissues although it can be also produced in other sites, including the placenta. ^[26] While its receptors are expressed in the uterine endometrium and trophoblast and the fetus. ^[27]

Although leptin, which has a molecular weight of 16,000 probably does not cross the placenta, there was a significant correlation in leptin concentrations between maternal serum and cord serum. This could be explained by the production of cord serum leptin by neonatal adipose tissue, placenta or both. In the present study cord serum leptin was correlated with maternal BMI before delivery, in agreement with previous studies.

Leptin have an essential role in regulation energy homeostasis and consequently body weight. ^[26] The presence of leptin in human cord blood and its relation to birth weight has been reported ^[31] We observed significant positive association of cord leptin with neonatal birth weight, in agreement with the results of other investigators. ^[32,33] Mastuda et al ^[33] reported correlation of serum leptin with body weight (r= 0.55; p <0.001). A similar correlation (r= 0.57; p <0.001) was reported by Schubring et al. ^[28], who observed in addition a positive association between cord serum leptin and placental weight (r= 0.50; p <0.01), suggesting the positive relation between leptin synthesis and release in circulation with birth weight and fetal fat mass during gestation. However, other investigators denied this association. ^[34] Leptin levels have been widely accepted as a marker for neonatal fat mass, and leptin receptors are expressed in several tissues including fetal cartilage, bone, lung, kidney, and hypothalami, suggesting that leptin may exert biological functions in the fetus and/or early in life. ^[35]

CONCLUSION

Maternal obesity at delivery is associated with increased neonatal serum leptin and body size with high risk of children obesity later in life.

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