

**IN VITRO BIOLOGICAL EVALUATION OF CURCUMIN AS
POTENTIAL 5-LIPOXYGENASE (5-LO) INHIBITOR****A. Kanakaraju^{1,*} and Y. Rajendra Prasad²**

¹Pharmaceutical Chemistry Division, Vignan Institute of Pharmaceutical Technology,
Beside SEZ, Duvvada, Visakhapatnam, Andhra Pradesh, Pincode-5530049, India

²Pharmaceutical Chemistry Division, A U College of Pharmaceutical Sciences,
Andhra University, Visakhapatnam, Andhra Pradesh, Pincode-530003, India

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Correspondence*For Author****Prof. A. Kanakaraju**

Pharmaceutical
Chemistry Division,
Vignan Institute of
Pharmaceutical
Technology, Beside SEZ,
Duvvada, Visakhapatnam,
Andhra Pradesh, Pincode-
5530049, India.

ABSTRACT

Curcumin (**1**), known from ancient time as an Ayurvedic medicine, popular as a spice in Asian cuisine, and has recently acknowledged as a drug candidate with distinctive therapeutic applications. Therefore, in the present study, Curcumin (**1**) has been studied for its inhibitory potential against 5-lipoxygenase (5-LO) enzyme.

KEYWORDS: Curcumin, 5-lipoxygenase (5-LO)**INTRODUCTION**

Curcumin (**1**) is the β -diketone present in the turmeric (*Curcuma longa* L.). It is an important phytoconstituent in the Ayurvedic system of medicine.^[1-4] Although, extensive research has been done, still there is a huge demand for Curcumin due to its inherent diversity of multitargeted pharmacological mechanisms against several diseases.^[5,6]

Lipoxygenases are a class of non-heme, iron-containing enzymes that catalyze the incorporation of molecular oxygen into 1,4,-*cis,cis*-pentadiene-containing fatty acids (e.g. linoleic and arachidonic acids) to form hydroperoxide products.^[7] The human isozymes, 5-, 12- and 15-Lipoxygenases are associated with different disease states, which suggest that selective inhibition may be important in targeting them for therapeutic purposes. 5-Lipoxygenase (5-LO), which was first discovered in 1976, plays an essential role in the biosynthesis of leukotrienes (LTs) that exert a large number of different biological activities

mediated by specific G-protein coupled receptors. LTB_4 is a typical proinflammatory mediator that recruits and activates leukocytes, whereas cysteinyl-leukotrienes C_4 , D_4 and E_4 cause vascular permeability and smooth muscle contraction. In view of these properties, development of drugs with 5-LO inhibitory activity have been hypothesized to possess therapeutic potential for treatment of asthma, allergic disorders and other inflammatory diseases.^[8] Based on the mechanism of action, the lipoxygenase inhibitors have been classified into four distinct classes: (i) iron chelating inhibitors, (ii) competitive reversible inhibitors, (iii) inhibitors of the 5-LO activating protein (FLAP) and (iv) anti-oxidative.^[9] Intensive discovery efforts in the development of clinically useful drugs from the inhibitors of 5-LO enzyme have led to one marketed drug; Zileuton (A-64066) and others, namely MK-3000, MK-886, MK-0591, ZM 211965, AKBA, BW A4C, LDP-977, Bay-X-1005, and Abt-761, which are evaluated at different stages of drug development [8,9]. As a part of our ongoing research in systematic investigation for identifying some novel bioactive compounds in relation to their 5-LO inhibitory activity, we have selected Curcumin (**1**).

EXPERIMENTAL

Chemicals

The compound Curcumin (**1**) was received from Pharmaceutical Chemistry Research Labs, Asia Metropolitan University, Malaysia, as gift sample (Courtesy of Dr.VasudevaRao).

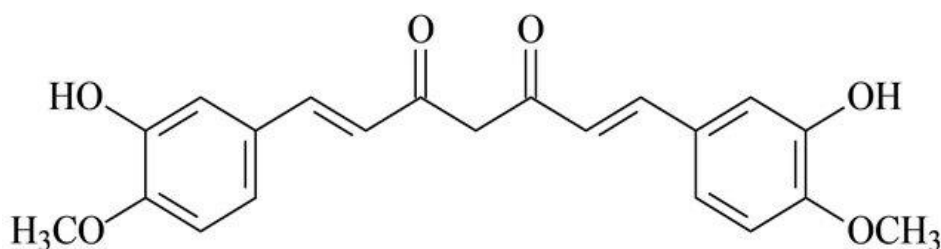


Figure 1: Chemical structure of Curcumin (**1**)

Enzyme 5-LO Inhibition Assay

The 5-LO inhibitory potential of the Curcumin (**1**) was determined by 5-LO inhibition assay (UV-Kinetic method) as described by Sircar *et al.*^[10] For the evaluation of 5-LO inhibitory activity, the enzymatic activity of 5-LO was measured spectrophotometrically using potato 5-LO,^[11] and an incubation mixture containing 80 mM linoleic acid and 50 mM sodium phosphate buffer (pH = 6.3). The reaction was initiated by the addition of an enzyme buffer mix to substrate (Linoleic acid) and the enzyme activity was monitored as an increase in rate

of absorbance at 234 nm on a UV/visible spectrophotometer (Varian Cary-50 UV-Visible spectrophotometer) for 120 sec. Each experiment was conducted by incubating along with control at various concentrations of the test substances with enzyme buffer mix for 2 min before addition of the substrate. The percentage inhibition was calculated by comparing slope or increase in absorbance of test substance with that of control enzyme activity. The assay was performed in triplicate and mean values were used for the calculation. The IC₅₀ values were obtained using fenny probed analysis software. The result for the test compound was compared with the positive control abietic acid (LI01020),^[12]

RESULTS AND DISCUSSION

Chemical synthesis

The chemical synthesis, physical and spectroscopic characterization of the selected compound Curcumin (**1**) was consistent with the earlier reports.^[13-15]

Enzyme 5-LO inhibition assay

From the analysis of *in vitro* 5-LO inhibitory activity screening data discovered that the Curcumin (**1**) showed comparatively more potential inhibitory activity, with IC₅₀ value of 3.37 ± 0.11 µg/mL in comparison with that of the standard drug (Abietic acid, IC₅₀ : 4.11 ± 0.23 µg/mL). The 5-LO inhibitory activity is significantly due to β-diketone moiety present in the compound. This observation has also been well supported by earlier report.^[16]

CONCLUSION

In summary, we could evaluate the 5-LO inhibitory potential of Curcumin (**1**) and it was appeared to be a significant inhibitor of potato 5-LO enzyme. This observation is noteworthy to develop some novel synthetic curcumin conjugated molecules as potential 5-LO inhibitors.

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REFERENCES

1. Srimal, R. C.; Dhawan, B. N. J. Pharm. Pharmacol., 1973; 25: 447.
2. Clarke, L.; Jackson, C. L. J. Am. Chem. Soc., 1908; 39: 696.
3. Spicer, C. P.; Strickland, J. D. H. J. Chem. Soc., 1952; 4644.

4. Tayyem, R. F.; Heath, D. D.; Al-Delaimy, W. K.; Rock, C. L. *Nutrition & Cancer.*, 2006; 55: 126.
5. Gerrard, W.; Lappert, M. F.; Shafferman, R. *Chem. Ind.*, 1958; 722.
6. Roughley, P. J.; Whiting, D. A. J. *Chem. Soc. Perkin Trans.*, 1973; 1: 2379.
7. Kenyon, V.; Chorny, I.; Carvajal, W. J.; Holman, T. R.; Jacobson, M.P. *J. Med. Chem.*, 2006; 49: 1356-1363.
8. Werz, O. *Curr. Drug Targets Inflamm. Allergy.*, 2002; 1: 23-44.
9. Babu, M. A.; Shakya, N.; Prathipati, P.; Kaskhedikara, S. G.; Saxena, A. K. *Bioorgan. Med. Chem.*, 2002; 10: 4035-4041.
10. Sircar, J. C.; Shwender, C. J.; Johnson, E. A. *Prostaglandins.*, 1983; 25: 393-396.
11. Reddenna, P.; Whelan, J.; Maddipati, R. K. R. *Methods Enzymol.*, 1990; 187: 268-277.
12. Ulusu, N. N.; Ercil, D.; Sakar, M. K.; Tezcan, E. E. *Phytother. Res.*, 2002; 16: 88-90.
13. Donaldson, E. M. *Talanta.*, 1981; 28: 825.
14. Spicer, C. P.; Strickland, J. D. H. *J. Chem. Soc.*, 1952; 4650.
15. Arrieta, A.; Dietze, F.; Mann, G.; Beyer, L.; Hartung, J. J. *Prakt. Chem.*, 1988; 330: 111.
16. Hong, J.; Bose, M.; Ju, J.; Ryu, J H.; Chen, X.; Sang, S. *Carcinogenesis.*, 2004; 25: 1671.