

# WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 5.990

Volume 4, Issue 12, 1377-1383.

Research Article

ISSN 2277-7105

# EFFICACY OF TERMINALIA CATAPPA L.WOOD AGAINST SOME BACTERIAL PATHOGENS

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Article Received on 15 Oct 2015,

Revised on 05 Nov 2015, Accepted on 25 Nov 2015,

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#### **ABSTRACT**

The present study was carried out to evaluate the anti bacterial activities of the aqueous, ethyl acetate and hexane extracts of Terminalia catappa wood against some pathogenic bacteria. Anti bacterial activity was assessed by agar disc diffusion method. Among the three extracts, aqueous extract exhibited potent antibacterial activity against all the selected bacterial pathogens even at minimum concentration. The activity of the extracts was compared with a standard antibiotic Chloramphenicol.

**KEYWORDS:** Antibacterial activity, Agar disc diffusion, Terminalia catappa L. wood.

#### INTRODUCTION

Nature has bestowed upon us a very rich botanical wealth. The search for eternal health and longevity and for remedies to relieve pain and discomfort drove early man to explore his immediate natural surroundings and led to the use of many plants, animal products and minerals for the development of a variety of therapeutic agents.<sup>[1]</sup>

In the recent past, the frequency of life threatening infections has increased dramatically. Even though antibiotics serve as the most powerful and successful treatment strategy to control infectious diseases, the rate of resistance of pathogenic microorganisms to conventionally used antimicrobial agents is increasing with an alarming frequency.<sup>[2,3,4]</sup> In addition to this problem antibiotics are sometimes associated with adverse side effects on the host, which include hypersensitivity, depletion of beneficial gut and mucosal

microorganisms, immunosuppression and allergic reactions.<sup>[5]</sup> The emergence and spread of multidrug-resistant bacterial pathogens have substantially threatened the current antibacterial therapy.<sup>[6]</sup>

Therefore, it is necessary to search the other alternatives that can potentially be effective in the treatment of these problematic bacterial infections. Plants are one of the bedrocks for modern medicine to attain new principles.<sup>[7]</sup> The usefulness of plant extracts for antimicrobial therapy and/or other diseases have been observed to be promising remedies since ancient time in Ayurveda and other traditional medicinal practices. Plant based antimicrobials represent a vast untapped source of medicine with enormous therapeutic potential and can serve the purpose without any side effects.

Terminalia catappa, is an important medicinal plant with diverse pharmacological spectrum. There are a number of phytochemicals present in this plant such as gallic acid, ellagic acid, corilagin and unidentified tannins which are responsible for many of the pharmacological activities. Due to the presence of number of phytoconstituents, the different extracts have exhibited anti-inflammatory and antioxidant<sup>[8,9]</sup>, antibacterial<sup>[10,11]</sup>, antidiabetic<sup>[12]</sup>, antifungal<sup>[13]</sup>, antitumor<sup>[14]</sup> activities. Since evidences are lacking about the antibacterial activity of wood, in the present study attempts are made to evaluate the antibacterial activity of aqueous, ethyl acetate and hexane extracts of T.catappa wood.

#### MATERIALS AND METHODS

#### **Collection of plant material**

Fresh wood of Terminalia catappa was collected from Mannargudi, Thiruvarur Dt, Tamil Nadu, India, which was carefully identified and authenticated in the department of CARISM, SASTRA University, Thirumalaisamudhram, Thanjavur. The wood was cut into pieces and washed thoroughly 2-3 times with running water and once with sterile distilled water, then the plant material was air-dried on sterile blotter under shade.

## Microorganisms

Microorganisms such as Escherichia coli, Bacillus subtilis, Klebsiella pneumoniae, Salmonella typhi, Proteus vulgaris, Streptococcus pyogenes and Staphylococcus aureus were obtained from SASTRA University, Thirumalaisamudhram, Thanjavur. Each Bacterial strain was suspended in Muller Hinton broth and incubated for 24 hrs at 37°C.

#### **Extraction of plant Material**

The coarse powder of Terminalia catappa wood was used for the extraction purpose. Extraction process was carried by soaking the coarse powder in Distilled water, Ethyl acetate and N - Hexane kept in shaker for 48 hrs, the extracts were filtered through whattman filter paper and evaporated the extracts using water bath.

# **Determination of Antimicrobial Activity**<sup>[15]</sup>

Agar well diffusion method was followed to determine the antimicrobial activity. Mullar Hinton agar (Hi media) medium plates were prepared by sterilizing the medium with the use of autoclave at  $121^{\circ}$  C and 15 lbs pressure for 15 minutes; petriplates were also sterilized using autoclave. After sterilization the cooled medium was poured into petriplates of about 25 ml and allowed to solidify. Muller Hinton agar medium plates were swabbed (sterile cotton swabs) with 24 hrs broth culture of bacteria. Four wells (10mm diameter) were made in each of these plates using sterile cork borer. The test solution was prepared by dissolving 1mg of extract in 1 ml of DMF. About 50, 100, 150  $\mu l$  from  $1\mu g$  /  $1\mu l$  concentration of aqueous, ethyl acetate and N - Hexane extracts of T. catappa wood , DMF (control) were added using micropipette into the wells and allowed to diffuse at room temperature for 2 hours. Chloramphenicol was used as standard antibiotic. The plates were incubated at room temperature for 24 hours. Diameters of the inhibition zones were recorded in mm.

#### RESULTS AND DISCUSSION

The use of higher plants and preparations made from them to treat infections is a longstanding practice in a large part of the population, especially in the developing countries, where there is dependence on traditional medicine for a variety of ailments.<sup>[16]</sup> Interest in plants with antimicrobial properties increased because of current problems associated with the antibiotics.<sup>[17,18]</sup> Recently, the antimicrobial effects of various plant extracts against certain pathogens have been reported by a number of researchers.<sup>[19,20,21,22,23]</sup>

Disc diffusion method is the most widely used procedure for testing antimicrobial susceptibility. The disc diffusion procedure (Kirby-Bauer method) has been accepted by the Food and Drug Administration (FDA) and as a standard by the National Committee for Clinical Laboratory Standards. [25]

Considering the vast potentiality of T. catappa, the current investigation was undertaken to screen the antibacterial activities of three different extracts of T. catappa wood against certain

pathogenic bacteria. The antibacterial activities of aqueous, ethyl acetate and hexane extracts of wood of Terminalia catappa were evaluated by the disc diffusion method against seven pathogenic organisms namely Escherichia coli, Bacillus subtilis, Klebsiella pneumoniae, Salmonella typhi, Proteus vulgaris, Streptococcus pyogenes and Staphylococcus aureus.

Aqueous extract of Terminalia catappa wood at various concentrations (50, 100, 150 $\mu$ l) were found to be effective against both gram positive and gram negative bacteria. The maximum inhibitory activity was observed for Bacillus subtilis (30mm) and then for Salmonella typhi (27mm) (Table 1). The activity of the herbal extract was compared with a standard antibiotic Chloramphenicol (10 $\mu$ g). Maximum inhibitory activity of the ethyl acetate extract was against Salmonella typhi (27mm/ 150  $\mu$ g/ml). The extract has no effect against Streptococcus pyogenes (Table 2).

In the case of hexane extract, there is no inhibitory activity against Proteus vulgaris and Klebsiella pneumoniae (Table 3). Maximum activity was revealed against Bacillus subtilis (29 mm), Streptococcus pyogenes (28 mm).

The results of the present study showed that aqueous extract of T. catappa wood has more antibacterial activity than ethyl acetate and hexane extracts even at minimum concentration. . This might have been due to the capacity of water to extract the antibacterial principles present in T. catappa wood.

Table 1: Antibacterial activity of the aqueous extract of T.catappa wood

S.No.	Bacteria	Zone of Inhibition (mm)			
5.110.	Бастепа	50μg	100µg	150µg	DMF (Control)
1	Proteus vulgaris	12	15	22	-
2	Escherichia coli	16	20	24	-
3	Streptococcus pyogenes	13	18	23	-
4	Staphylococcus aureus	14	18	24	-
5	Klebsiella pneumoniae	17	21	25	-
6	Salmonella typhi	10	15	27	-
7	Bacillus subtilis	7	18	30	-

Table 2: Antibacterial activity of the Ethyl acetate extract of T.catappa wood

S.No.	Bacteria	Zone of Inhibition (mm)			
5.110.	Dacteria	50μg	100µg	150µg	DMF (Control)
1	Proteus vulgaris	-	20	22	1
2	Escherichia coli	10	14	18	-
3	Streptococcus pyogenes	-	-	-	-
4	Staphylococcus aureus	10	13	21	-
5	Klebsiella pneumoniae	10	15	21	-
6	Salmonella typhi	9	20	27	-
7	Bacillus subtilis	5	19	21	-

Table 3: Antibacterial activity of the Hexane extract of T.catappa wood

S.No.	Bacteria	Zone of Inhibition (mm)			
5.110.	Dacteria	50μg	100µg	150µg	DMF (Control)
1	Proteus vulgaris	-	ı	1	-
2	Escherichia coli	-	16	23	-
3	Streptococcus pyogenes	-	17	28	-
4	Staphylococcus aureus	12	14	19	-
5	Klebsiella pneumoniae	-	-	-	-
6	Salmonella typhi	13	20	26	-
7	Bacillus subtilis	6	11	29	-

Table 4: Antibacterial activity of Chloramphenicol

S.No.	Bacteria	Zone of Inhibition (mm) Chloramphenicol (10µg/ml)
1	Proteus vulgaris	26
2	Escherichia coli	25
3	Streptococcus pyogenes	20
4	Staphylococcus aureus	12
5	Klebsiella pneumoniae	26
6	Salmonella typhi	56
7	Bacillus subtilis	49

## **CONCLUSION**

The need of the hour is to find new antimicrobials because the microorganisms are getting resistant to the existing antibiotics. The persistent increase in multi drug resistant strains compels the search for new effective and affordable antimicrobial drugs. The results of the present study signify the potentiality of Terminalia catappa wood as a source of therapeutic agent which may provide leads in the ongoing search for antimicrobial botanicals.

#### **ACKNOWLEDGEMENT**

Authors extend a deep sense of gratitude to the Management, S.T.E.T. Women's College, Mannargudi and Vice Chancellor, SASTRA University, Thirumalaisamudhram for providing necessary infrastructural facilities.

#### REFERENCES

- 1. Nair R, Chanda S Antibacterial activities of some medicinal plants of the Western region of India. Turkish Journal of Biology, 2007; 31: 231-236.
- 2. Ge Y, Difuntorum S, Touami S, Critchley I, Burli R, Jiang V, Drazan K, Moser H In vitro antimicrobial activity of GSQ1530, a new heteroaromatic polycyclic compound. Antimicrobial Agents and Chemotherapy, 2002; 46: 31683174.
- 3. Nair R, Chanda S Anticandidal activity of Punica granatum exhibited in different solvents. Pharmaceutical Biology, 2005; 43: 21-25.
- 4. Neogi U, Saumya R, Mishra RK, Raju KC Lipid content and in vitro antimicrobial activity of oil of some Indian medicinal plants. Current Research in Bacteriology, 2008; 1: 1-6.
- 5. Al-Jabri AA Honey, milk and antibiotics. African Journal of Biotechnology, 2005; 4: 1580-1587.
- 6. Boucher HW, Talbot GH, and Bradley JS "Bad bugs, no drugs: no ESKAPE! An update from the Infectious Diseases Society of America," Clinical Infectious Diseases, 2009; 48(1): 1–12.
- 7. Evans CE, Banso A, Samuel OA Efficacy of some medicinal plants against Salmonella typhi: an in vitro study. Journal of Ethnopharmacology, 2002; 80: 21–24.
- 8. Sivaranjani C, Venkatalakshmi P, and Brindha P In Vitro Anti Inflammatory and Antioxidant Activities on Fruits of Terminalia catappa L. Research J. Pharm. and Tech, 2015; 8(10): 1409-1411.
- 9. Venkatalakshmi P, Brindha, P, Vellingiri Vadivel In vitro Antioxidant and Antiinflammatory Studies on Bark, Wood and Fruits of Terminalia catappa L. International Journal of Phytomedicine, 2015; 7(3): (In press).
- 10. Neelavathi P, Venkatalakshmi P, Brindha P Antibacterial activities of aqueous and ethanolic extracts of Terminalia catappa leaves and bark against some pathogenic bacteria. Int. J. Pharm. Pharm. Sci, 2013; 5(1): 114–120.
- 11. Sangavi R, Venkatalakshmi P and Brindha P Anti bacterial activity of Terminalia catappa L.Bark against some bacterial pathogens. World Journal of Pharmacy and Pharmaceutical Sciences, 2015; 4(9): 987-992.
- 12. Nagappa AN, Thakurdesai PA, Venkat Rao N, Singh J Antidiabetic activity of Terminalia catappa Linn fruits. J. Ethnopharmarcol, 2003; 88: 45–50.

- 13. Parimala Gandhi P, Venkatalakshmi P and Brindha P Efficacy of Terminalia catappa L. Wood and Bark against Some Fungal Species. International Journal of Current Microbiology and Applied Sciences, 2015; 4(9): 74-80.
- 14. Venkatalakshmi P, Brindha P, Induja K Invitro anti oxidant and anti tumour activities of Terminalia catappa Bark. Int. J. Pharm. Pharm. Sci, 2014; 6(1): 1–3.
- 15. Perez J and Chanda S. In vitro antifungal activity of methanol extracts of some Indian medicinal plants against pathogenic yeast and moulds. African journal of Biotechnology, 1990; 7: 4349-4353.
- 16. Ahmad ZM and Mohammad F Screening of some Indian medicinal plants for their antimicrobial properties. Journal of Ethnopharmacology, 1998; 62: 183-193.
- 17. Emori TG and Gaynes RP An overview of nosocomial infections, including the role of the microbiology laboratory. Clinical Microbiology Reviews, 1993; 6: 428-442.
- 18. Pannuti CS and Grinbaum RS An overview of nosocomial infection control in Brazil. Infection Control and Hospital Epidemiology, 1995; 16: 170-174.
- 19. Ahmed I and Beg AZ Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multidrug resistant human pathogens. Journal of Ethnopharmacology, 2001; 74: 113-123.
- 20. Erasto P, Bojase-Moleta RR, Majinda RR Antimicrobial and antioxidant flavonoids from the root wood of Bolusanthus speciosus. Phytochemistry, 2004; 65: 875-880.
- 21. Carneiro BAL, Teixeira MFS, de Oliveira VMA, Fernandes OCC, Cauper GSB, Pohlit AM Screening of Amazonian plants from the adolpho ducke forest reserve, Manaus, state of Amazonas, Brazil, for antimicrobial activity. Mem Inst Oswaldo Cruz, Rio de Janeiro, 2008; 103: 31-38.
- 22. Liasu MO, Ayandele AA Antimicrobial activity of aqueous and ethanolic extracts from Tithonia diversifolia and Bryum coronatum collected from Ogbomoso, Oyo state, Nigeria. Advances in Natural and Applied Sciences, 2008; 2: 31-34.
- 23. Parekh J, and Chanda S In vitro antimicrobial activity of Trapa natans L. fruit rind extracted in different solvents. African Journal of Biotechnology, 2007; 6: 760-770.
- 24. Sambath Kumar R, Sivakumar T, Sundram RS, Sivakumar P, Nethaji R, Gupta M, Mazumdar UK (2006) Antimicrobial and antioxidant activities of Careya arborea Roxb. Stem bark. Iranian Journal of Pharmacology and Therapeutics 5: 35-41.
- 25. Barry AL and Thornsberry C (1985) In: Manual of Clinical Microbiology, Lennette EH (Ed.), American Association for Microbiology, Washington pp. 978-987.