

WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 6.805

Volume 5, Issue 5, 722-727.

Research Article

ISSN 2277-7105

ANTIMICROBIAL ACTIVITY OF MORINGA OLEIFERA (MORINGACEAE) AND BARLERIA PRIONITIS (ACANTHACEAE) METHANOLIC CRUDE LEAVES EXTRACT

Padma Shrivastava^{1*} and Priyam Singh²

¹Dept. of Biotechnology, Govt. P.G. College, BHEL, Bhopal (M.P.), India.

Article Received on 02 March 2016, Revised on 23 March 2016, Accepted on 11 April 2016 DOI: 10.20959/wjpr20165-6043

*Corresponding Author Padma Shrivastava

Dept. of Biotechnology, Govt. P.G. College, BHEL, Bhopal (M.P.), India.

ABSTRACT

The nature provides the mankind vast therapeutic flora with a wide variety of medicinal potential. According to WHO world's 80% population depends on traditional medicines for healthcare. Phytochemical screening show the presence of saponin, flavonoids, carbohydrate, glycosides, terpenes and steroids in *Moringa oleifera* methanolic crude leaves extract. Phytochemical screening of *Barleria prionitis* showed the presence of terpenes, glycosides and flavonoids. Both the plants showed the anti microbial activity in the dose dependent manner. *M. oleifera* mehanolic crude leaf extract showed antimicrobial activity against *Pseudomonas aeruginosa* with zone of

inhibition of 6.0 ± 0.05 , 10.0 ± 0.02 , 11.0 ± 0.01 , 14.0 ± 0.033 mm in concentration of 25,50,75,100 % respectively of crude extract as compare to Streptomycin 9.9 ± 0.47 with concentration of $10\mu g/disc$. *B. prionitis* methanolic crude extract showed antimicrobial activity against *Streptococcus mutans* with zone of inhibition of 6.0 ± 0.05 , 10.0 ± 0.02 , 6.0 ± 0.01 , 7.0 ± 0.033 mm in concentration of 25,50,75,100 % respectively of crude extract as compare to Streptomycin 9.9 ± 0.47 with concentration of $10\mu g/disc$.

KEYWORDS: antimicrobial, medicinal potential, flavonoids, saponin, steroids, terpenes.

INTRODUCTION

Since time immemorial man has been using nature based therapeutics for treatment of various ailments. These nature based remedies not only possess potential for uprooting various deadly diseases but also act without any side effect. The potential for such properties lies in some chemical components present in plants known as secondary metabolites. The most

²Dept. of Botany, Govt. P.G. College, Narsinghgarh, Rajgarh (M.P.), India.

important of these bioactive compounds of plants are flavonoids, terpenes, alkaloids, tannins, saponins and phenolic compounds. There must be a continuous research for new drugs to cope up with multidrug resistance in pathogens which is one of the most serious threats to successful treatment of microbial diseases. ^[1] The increasing failure of the chemotherapeutics and antibiotic resistance exhibited by pathogenic microbial infectious agents has lead to the screening of several medicinal plants for their potential antimicrobial activity. ^[2] *Pseudomonas aeruginosa* is a common gram-negative rod-shaped bacterium that can cause disease in plants and animals, including humans. It is a species of considerable medical importance, due to multidrug resistant pathogen and its association with serious illnesses like nosocomial infections such as ventilator-associated pneumonia and various sepsis syndromes. In higher plants *P. aeruginosa* induces symptoms of soft rot, for example in *Arabidopsis thaliana* (Thale cress) ^[3] and *Lactuca sativa* (lettuce). ^[4,5] It is also pathogenic to invertebrate animals, including the nematode *Caenorhabditis elegans*, ^[6,7] the fruit fly *Drosophila melanogaster* and the moth *Galleria mellonella*. ^[9] The associations of virulence factors are the same for plant and animal infections. ^[10]

Streptococcus mutans is facultative anaerobe gram-positive bacterium commonly found in the human oral cavity and is a significant contributor to tooth decay and various oral caries. [11, 12]

MATERIALS AND METHODS

Collection of plant material

The plants of *Moringa oleifera* (Moringaceae) were collected from areas around Bhopal, and *Barleria prionitis* (Acanthaceae) from BHEL area, Bhopal, M.P. in India. The plant material was thoroughly washed with water and was kept for drying in shade at room temperature for 20days. The air dried plant material was grinded to powder to about 40-60 mesh size weighted and stored in large plastic bottles.

Crude Extraction

The plant powder was extracted according to Harborne and Baxter. ^[13] In this method the 40-60-mesh size powdered plant material was extracted with soxhlet apparatus using 50 % methanol after defatting with petroleum ether for 24 hr. The extraction was done for 48 hr. duration. Almost all the chlorophyll and lipid is deposited on the side of the flask and with skill it was removed. The crude was pipette off skilfully almost completely free of lipid

impurities and filtered through Whatmann filter paper no.1. Filtrate is then evaporated to 1/10th volume (<40°C).

Phytochemical screening

Phytochemical screening were performed to assess the qualitative chemical composition of different crude extracts using commonly employed precipitation and coloration reactions to identify the major secondary metabolites like alkaloids, flavonoids, saponins, phenolic compounds, tannins. The phytochemical analyses were carried out using standard procedures.^[14, 15]

Antibacterial assay

The plate disc diffusion assay was used to determine the growth inhibition of bacteria by plant crude extract. The bacterial cultures were maintained on nutrient agar media (Himedia). The bacteria were incubated at 37° for 24 hr. Nutrient agar media was prepared and poured in autoclaved Petri dishes. Inoculum containing 106 CFU/ml of test bacteria was spread on solidified plates with a sterile swab moistened with the bacterial suspension. Sterilized Whatmann filter paper discs were placed on the poured plates using sterile forceps and the 10µl of the crude extract was poured on the discs. The four different concentrations (25, 50, 75, 100 %) of leaves extract were prepared by dissolving in DMSO (Dimethyl Sulphoxide) and were tested for antibacterial activity. The experiment was performed in triplicates and the mean values were presented with standard deviation. Streptomycin (10µg/50µl) was used as standard. Control was prepared using DMSO only. Zone of inhibition were measured in mm.

OBSERVATIONS

Table 1. Phytochemistry of methanol crude extract of M. oleifera and B. prionitis leaves

C No	Compound	Methanolic Crude Extract			
S.No.		M. oleifera	B. prionitis		
1	Carbohydrate	+ve	-ve		
2	Glycosydes	-ve	+ve		
3	Terpenes	+ve	+ve		
4	Flavonoids	+ve	+ve		
5	Alkaloids	-ve	-ve		
6	Saponin	+ve	-ve		
7	Phenolic	VA	-ve		
/	Compounds	-ve			

Table 2. Antimicrobial activity of methanol crude extract of M. oleifera on P.aeruginosa

		Standard	Zone Of Inhibition(mm)				
S.No.	Bacterial	(Streptomycin)	Methanol Crude Extract				
	Strains		(%)				
		10μg/disc	25	50	75	100	
1	Pseudomonas aeruginosa	9.9 ± 0.47	6.0 ± 0.05	10.0 ± 0.02	11.0 ± 0.01	14.0 ± 0.033	

Table 3.Antimicrobial activity of methanol crude extract of B. prionitis on S. mutans

		Standard	Zone Of Inhibition(mm)			
S.No.	Bacterial	(Streptomycin)	Methanol Crude Extract			
	Strains		(%)			
		10μg/disc	25	50	75	100
1	Streptococcus mutans	9.9 ± 0.47	6.0 ± 0.05	10.0 ± 0.02	6.0 ± 0.01	7.0 ± 0.033

RESULTS AND DISCUSSION

Phytochemical screening show the presence of saponin, flavonoids, carbohydrate, glycosides, terpenes and steroids in Moringa oleifera methanolic crude leaves extract. Phytochemical screening of *Barlaria prionitis* showed the presence of glycosides, terpenes and flavonoids. Both the plants showed the antimicrobial activity in the dose depended manner. M.oleifera methanolic crude leaves extract showed antimicrobial activity against Pseudomonas aeruginosa with zone of inhibition of 6.0 ± 0.05 , 10.0 ± 0.02 , 11.0 ± 0.01 , 14.0 ± 0.033 mm in concentration of 25,50,75,100 % respectively of crude extract as compare to 9.9 ± 0.47 with concentration of 10µg/disc. B. prionitis methanolic crude extract showed antimicrobial activity against S. mutans with zone of inhibition of 6.0 ± 0.05 , 10.0 ± 0.02 , 6.0 ± 0.01 , $7.0 \pm$ 0.033 mm in concentration of 25,50,75,100 % respectively of crude extract as compare to 9.9 ± 0.47 with concentration of $10\mu g/disc$. As flavonoids are synthesized by plants in response to microbial infection it should not be surprising that they have been found invitro to be effective antimicrobial substances against a wide array of microorganisms. Generally alkaloids are extremely toxic though they do have a marked therapeutic effect in minute quantities. For this reasons alkaloids were not often used in folk medicine but for external application only. Pure isolated plant alkaloids and their synthetic derivatives are used as basic medicinal agents over the world for their analgesic, antispasmodic and bactericidal effects. [17] This antibacterial activity would support the folk therapy of infections whose symptoms might involve bacteria. [18] Antibiotics provide the base for treatment of bacterial infections. But, for the possession of high genetic variability bacteria easily develop antibiotic resistance. Thus, there has been a continuing search for new and more potent antibiotics. [19]

CONCLUSIONS

The two botanicals showed antibacterial activity in dose dependent manner. Methanolic crude extract from *Moringa oleifera* (Moringaceae) and *Barleria prionitis* (Acanthaceae) leaves showed potent antibacterial activity thus this plant can be used for isolation of bioactive compounds, as lead compound for development of novel antibiotics.

ACKNOWLEDGEMENT

The Author is highly thankful to the Dr. Shobna Bajpai Maru, Principal, Govt. P.G. College, BHEL, Bhopal (M.P.), India for providing research facilities. The author is also thankful to Dr. Abhishek Gupta, Director, Center for Microbiology and Bio-Technology (CMBT) research and training institute, Bhopal, (M.P.), India for providing microbial strains. The author is also thankful to Dr. Priyam Singh, Guest Faculty, Dept. of Botany, Govt. P.G. College, Narsinghgarh, Dist. Rajgarh, (M.P.), India.

REFERENCES

- 1. Chaudhry NMA, Tariq P. Invitro antibacterial activities of kalongi, cumin and poppy seeds, Pak. J. Bot., 2008; 40: 461-467.
- 2. Colombo ML, Bosisio E. Pharmacological activities of *Chelindonium majus* L. (Papaveraceae). Pharmacol. Res. 1996; 33: 124-34.
- 3. Walker TS, Bais HP, Déziel E, Schweizer HP, Rahme LG, Fall R, Vivanco JM. "*Pseudomonas aeruginosa*-plant root interactions. Pathogenicity, biofilm formation, and root exudation". Plant Physiol., 2004; 134 (1): 320–331. doi:10.1104/pp.103.027888. PMC 316311. PMID 14701912.
- 4. Rahme LG, Stevens EJ, Wolfort SF, Shao J, Tompkins RG, Ausubel FM. "Common virulence factors for bacterial pathogenicity in plants and animals". Science, 1995;268 (5219): 1899–1902. doi:10.1126/science.7604262. PMID 7604262.
- Rahme LG, Tan MW, Le L, Wong SM, Tompkins RG, Calderwood SB, Ausubel FM.
 "Use of model plant hosts to identify *Pseudomonas aeruginosa* virulence factors". Proc.
 Natl. Acad. Sci. U.S.A., 1997; 94 (24): 13245–50. doi:10.1073/pnas.94.24.13245. PMC
 24294. PMID 9371831.
- Mahajan-Miklos S, Tan MW, Rahme LG, Ausubel FM. "Molecular mechanisms of bacterial virulence elucidated using a *Pseudomonas aeruginosa-Caenorhabditis elegans* pathogenesis model". Cell, 1999; 96 (1): 47–56. doi:10.1016/S0092-8674(00)80958-7. PMID 9989496.

- 7. Martínez C, Pons E, Prats G, León J. "Salicylic acid regulates flowering time and links defence responses and reproductive development". Plant J., 2004; 37(2): 209–17. doi:10.1046/j.1365-313X.2003.01954.x. PMID 14690505.
- 8. D'Argenio DA, Gallagher LA, Berg CA, Manoil C. "Drosophila as a model host for *Pseudomonas aeruginosa* infection". J. Bacteriol. 2001; 183(4): 1466–71. doi:10.1128/JB.183.4.1466-1471.2001. PMC 95024. PMID 11157963.
- 9. Miyata S, Casey M, Frank DW, Ausubel FM, Drenkard E. "Use of the *Galleria mellonella* caterpillar as a model host to study the role of the type III secretion system in *Pseudomonas aeruginosa* pathogenesis". Infect. Immun., 2003; 71(5): 2404–13. doi:10.1128/IAI.71.5.2404-2413.2003. PMC 153283. PMID 12704110.
- 10. Rahme LG, Ausubel FM, Cao H, Drenkard E, Goumnerov BC, Lau GW, Mahajan-Miklos S, Plotnikova J, Tan MW, Tsongalis J, Walendziewicz CL, Tompkins RG. "Plants and animals share functionally common bacterial virulence factors". Proc. Natl. Acad. Sci. 2000; U.S.A. 97(16): 8815–21. doi:10.1073/pnas.97.16.8815. PMC 34017. PMID 1092204.
- 11. Ryan KJ, Ray CG (editors). Sherris Medical Microbiology(4ed.).Mc Graw Hill: ISBN 0-8385-8529-9: 2004.
- 12. Loesche WJ."Ch.99: Microbiology of Dental Decay and Periodontal Diseases". In Baron S *et al.*, Baron's Medical Microibiology(4th ed.). University of Texas Medical Branch. ISBN 0-9631172-1-1. 1996, PMID 21413316.
- 13. Harborne SB, Baxter H. Phytochemical Dictionary. A handbook of bioactive compounds from plants. London; Taylor and Francis: 1995.
- 14. Sofowora A, Medicinal plants and Traditional Medicine in Africa. Spectrum Books, Ibadan, 1993; 150.
- 15. Trease GE, Evans WC, Pharmacognosy. 13th edn. London; Bailliere Tindal, 1989, 176-180.
- 16. Anon, The Indian Pharmacopoeia 3rd edition. Government of India. New Delhi. Ministry of Health and Family Welfare, 1996.
- 17. Stary F. The natural guide to medicinal herbs and plants. Barnes and Noble Inc., 1996.
- 18. Veerpoorte R, Tjin A, Van Doorne H, Baerheim Svendsen A. Medicinal plants of Surinam. Antimicrobial activities of some medicinal plants. J. Ethnopharmacol., 1982; 5: 221-226.
- 19. Heisig P. Planta Medica, 2001; 67: 4-12.