

**EVALUATION OF ANTIULCER AND ANTIOXIDANT ACTIVITIES
OF *MURRYA KOENIGI* AND *JATROPHA CURCAS* IN ALBINO RATS****Shweta Sao*¹, Saurabh Dubey² and Nameer – Al – Hasan³**

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ABSTRACT

India has a wide variety of medicinal plants. Medicinal plants synthesize variety of phytochemicals that help to defend against various types of neoplasm. Due to environment pollution and feeding habits of people, risk of generation of Free radicals and reactive oxygen species is also increasing. These free radicals are responsible for generation of various ulcers, tumors and Neoplasms. The phytochemicals present in *M. koenigi* and *J.curcas* has antioxidant effects which may prevent against various Neoplasms. Use of plants as a source of medicine has been an ancient practice and is an important component of health system in India. General awareness, academic and

government interest in traditional medicine is growing rapidly due to increased side effects of adverse drug reactions and cost factors of modern system of medicine. The present study focuses on evaluation of anticancer and antioxidant activities of *Murrya koenigi* and *Jetropha curcas* plant extracts.

KEYWORDS: *Murrya koenigi* and *Jetropha curcas*.**INTRODUCTION**

Cancer is also called Neoplasm and the growth of cancer is called Neoplasia. The term neoplasia means “new growth”. Thus, the term Neoplasm may be defined as – ‘A mass of tissue formed as a result of abnormal, excessive, uncontrolled, uncoordinated, autonomous proliferation of the cells.’ Neoplasm may be Benign when they are slow growing and localized without causing much effect on the host or Malignant when they proliferate rapidly, spread throughout the body and eventually cause death of the host.

The common term used for the entire malignant tumor is Cancer. Hippocrates (460-377 BC), coined the term Karkinos for the cancer of breast. The word Cancer means Crab, thus reflecting the true characteristic of cancer, since it sticks to the affected part stubbornly like a crab.

TYPES OF PEPTIC ULCER

Duodenal Ulcers (DU), Gastric Ulcers (GU), Stress Ulcers (SU), NSAID Induced Ulcers

1.1. PEPTIC ULCERS AND HELICOBACTER PYLORI

H.pylori a gram negative bacteria that colonise the gastric mucosa and has been found in the cases of chronic gastritis and peptic ulcers. Its prolonged infection may lead to Lymphoma and Gastric carcinoma.

RESULT AND DISCUSSION

Table 1.2 Evaluation of antiulcer activity of *Murrya koenigi* and *Jetropha curcas* leaf extracts by Ethanol-induced ulcer model.

Groups	SOD	CATALASE	GSH
Group-1			
Rat-1	1	1	1
Rat-2	1	1	1
Rat-3	1	1	1
Rat-4	1	1	1
Rat-5	1	1	1
Rat-6	1	1	1
Group-2			
Rat-1	3	2	3
Rat-2	3	3	3
Rat-3	2	3	3
Rat-4	3	3	3
Rat-5	3	2	3
Rat-6	3	3	3
Group-3			
Rat-1	2	2	2
Rat-2	2	2	0
Rat-3	2	1	2
Rat-4	1	0	1
Rat-5	2	1	2
Rat-6	1	2	0
Group-4			
Rat-1	1	1	1
Rat-2	1	1	0
Rat-3	2	1	1
Rat-4	1	1	0
Rat-5	1	1	0

Rat-6	0	0	0
Group-5			
Rat-1	0	1	1
Rat-2	0	0	1
Rat-3	1	0	2
Rat-4	0	1	1
Rat-5	0	1	1
Rat-6	0	1	1

TABLE 1.3 Evaluation of Antioxidant levels in Ethanol induced peptic ulcer -

Groups	Body Wt. (gm)	Treatment	Ulcer Index						Total Score
			Normal Stomach	Red Coloration	Spot Ulceration	Hemorrhagic Streaks	Ulcer	Perforation	
1.	180	Normal Control	0	0.5	-	-	-	-	0.5
	175		0	0.5	-	-	-	-	0.5
	190		0	0.5	-	-	-	-	0.5
	205		0	-	-	-	-	-	0
	200		0	-	-	-	-	-	0
	195		0	0.5	-	-	-	-	0.5
2.	185	Control	-	0.5	1.0	1.5	2.0	-	5.0
	180		-	0.5	1.0	1.5	2.0	-	5.0
	190		-	0.5	1.0	1.5	2.0	-	5.0
	195		-	0.5	1.0	1.5	2.0	3.0	8.0
	198		-	0.5	1.0	1.5	2.0	-	5.0
	200		-	0.5	1.0	1.5	2.0	-	5.0
3.	180	Standard (20mg/kg)	-	0.5	1.0	-	-	-	1.5
	220		-	-	-	-	-	-	0
	191		-	0.5	1.0	-	-	-	1.5
	198		-	0.5	-	-	-	-	0.5
	205		-	0.5	-	-	-	-	0.5
	200		-	-	-	-	-	-	0
4.	220	Methanolic Extract of M.koenigi (100mg/kg)	-	0.5	1.0	1.5	-	-	3.0
	180		-	0.5	1.0	-	-	-	1.5
	190		-	0.5	1.0	-	-	-	1.5
	195		-	0.5	-	-	-	-	0.5
	180		-	0.5	-	-	-	-	0.5
	210		-	0.5	-	-	-	-	0.5

5.	220	Methanolic extract of J.curcas	-	0.5	-	-	-	-	0.5
	180		-	0.5	1.0	-	-	-	1.5
	190		-	0.5	1.0	1.5	-	-	3.0
	195		-	0.5	-	-	-	-	0.5
	180		-	0.5	1.0	-	-	-	1.5
	210		-	0.5	1.0	-	-	-	1.5

(key: 0=negligible level, 1=Mild level, 2=Moderate level, 3=High level)

4.1 DISCUSSION

Peptic Ulcer Disease (PUD) encompassing gastric and duodenal ulcer is the most prevalent chronic gastrointestinal disorder and is inflammatory in nature, Valle (2008). The pathophysiology of PUD involves an imbalance between offensive or injurious (acid, pepsin and *Helicobacter pylori*) and defensive mucosal factors (mucin, prostaglandin, bicarbonate, nitric oxide and growth factors), Wallace *et al.*, (2011).

Most widely used method for producing experimental gastric ulcers is Ethanol induced, as it is suitable for first line antiulcer screening, because the agents are retained in the stomach and may act by a variety of mechanisms, Vimlesh *et al.*, (2013). Reproducibility and high incidence of ulceration has been reported by this method. A major advantage of this method is that one can measure gastric secretory rate, percentage ulceration and ulcer severity in the same animal.

Intragastric application of absolute ethanol is a reproducible method to produce gastric lesion in experimental animals, Robert *et al.*, (1979) and Szabo *et al.*, (1980). The protective effect against various irritants has been called cytoprotective activity, Robert (1979). The concept of gastric cytoprotection signifies protection against mucosal injury by a mechanism other than inhibition of acid secretion was introduced long ago, Rober *et al.*, (1979) and Vogel (2008). In the ethanol-induced gastric ulceration model, ethanol produces necrotic lesions by direct necrotizing action which in turn reduces defensive factors like the secretion of bicarbonate and production of mucus, Marhuenda *et al.*, (1993). Ethanol-induced gastric lesions impaired gastric defensive factors such as mucus and mucosa circulation, Ferreira *et al.*, (2008). Ethanol causes necrotic lesions of the gastric mucosa in a multifactorial way. It can reach the mucosa by disruption of the mucus-bicarbonate barrier and cause cell rupture in the wall of blood vessels. These effects are probably due to biological actions, such as of lipid peroxidation, formation of free radicals, intracellular oxidative stress, changes in permeability and depolarization of the mitochondrial membrane prior to cell death, Sannomiya *et al.*, (2005).

Oral administration of absolute ethanol is noxious to the stomach since it affects the gastric mucosa topically by disrupting its barrier and provoking pronounced microvascular changes within a few minutes after its application, Moleiro *et al.*, (2009). In addition, it produces linear hemorrhagic lesions, extensive submucosal edema, mucosal friability, inflammatory cells infiltration, and epithelial cell loss in the stomach, which are typical characteristics of alcohol injury, Jelski *et al.*, (2009). The pathogenesis of ethanol-induced gastric mucosal damage occurs directly and indirectly through various mediators such lipoxxygenase, cytokines and oxygen-derived free radicals, Abdel-Salam *et al.*, (2001). Mucus secretion is regarded as a crucial defensive factor in the protection of the gastric mucosa from gastric lesions, Oluwole *et al.*, (2008). Thus in the present study, it could be assumed that the existence of the cytoprotective effect of compound is present in the methanol extract of the *Murrya koenigi* and *Jatropha curcas* leaves because it showed the significantly less ulcer index in the observation and high mucin concentration when the results were compared with the control group.

An antioxidant is any substance that retards or prevents deterioration, damage or destruction by oxidation, Dekkers *et al.*, (1996). A free radical is a compound with one or more unpaired electrons in their outer orbital, Jesberger *et al.*, (1991). The most dangerous free radicals are

the atomic and molecular varieties of oxygen which is known as Reactive Oxygen Species (ROS). While ROS are not technically free radicals, they are highly reactive with the molecules around them, Sharma *et al.*, (1996). To neutralize these free radicals antioxidants play an important role, Vani *et al.*, (1997). Antioxidant enzymes such as super oxide dismutase (SOD), Catalase and Glutathione peroxidases are known to attenuate the generation of ROS by removing potential oxidants or by transforming ROS and Reactive nitrogen species into stable compounds, Ashok, (2001).

The level of CAT, SOD and GSH was significantly decreased in diseased control group as compared to normal group. Administration of methanolic extract of both the plants had shown significant increased in the levels of CAT, SOD and GSH as compared to the diseased control animals, which suggests its efficacy in preventing free radical-induced damage.

REFERENCES

1. Ahamad Nisar, Fazal Hina, Haider Bilal, Abbasi. (2010). Efficient Free Radical Scavenging Activity of Ginkgo biloba, Stevia rebaudiana and Parthenium hysterophorus Leaves through DPPH. 2(3).
2. Ahirrao R A, Patel M R, Pokal D M, Patil J K, Suryawanshi H P. (2011). Phytochemical screening of leaves of Jatropha curcas plant. 2(4): 1324.
3. Ahsan, Rajib, Islam, Monirul K M, Haque E, Mossaddik A. (2009). In vitro Antibacterial Screening and Toxicity Study of some Plants. 5(5): 617-621.
4. Aiyelaagbe O O, Adeniyi B A, Fatunsin O F, Arimah B D. (2007). In vitro Antimicrobial activity and Phytochemical Analysis of Jatropha curcas Roots. International Journal of Pharmacology. 3(1): 106-110.
5. Asthana Amrita, Mall H V, Dixit Kalpana, Gupta S. (1989). Fungitoxic Properties of Latex of Plants with special reference to that of Croton bonplandianum boill. Pharma Biology. 27(1): 25-28.
6. Beevi S S, Rasheed M H, Geeha A. (2007). Evidence of Oxidative and Nitrosative Stress in Patients with Cervical Squamous Cell Carcinoma. Clin Chim Acta. 375(1-2): 119-23.
7. Dwivedi Deepak, Kumar Ashok, Rana R, Vihan V S. Antibacterial activity of methanolic extracts of some local plants against pathogenic E.coli isolates from kid diarrhea. Journal of veterinary practitioner, 8(2): 119-120.

8. Ehsan Oskueian, Norhani Abdullah, Wan Zuhainis Saad, Omar Rahman Abdul, Ahamad Syahida, Wen Bin Kuan. (2011). Antioxidant, Anti-inflammatory and Anti-cancer Activities of Methanolic Extracts from *Jatropha curcas*. 5(1): 49-57.
9. Ghais uddin, Abdur Rauf, Naveed Muhammad, Shabana, Nadia Malik, Mohsina. Phytochemical and pharmacological studies of the whole plant of *Calotropis procera*. Middle East Journal of Medicinal Plants Research., 2012; 1(4): 71-74.
10. Hachem C Y, Clarridge J E, Evans D G, Graham D Y. (1995). Comparison of Agar Based Media for Primary Isolation of *Helicobacter pylori*. Clin Pathol. 48: 714-716.
11. Igbiosa O O, Igbiosa E O, Aiyengoro O A. (2009). Antimicrobial Activity and Phytochemical Screening of Stem Bark Extracts from *Jatropha curcas*. African Journal of Pharmacology. 3(2): 058-062.
12. M Smita K. Naidu, Suryakar A N, Sanjay C, Kathkam R V, Kumbar K M, (2007). Oxidative stress and Antioxidant Status in cervical Cancer Patients. Indian Journal of Clinical Biochemistry. 22(2): 140-144.
13. Manmohan S, Klanjiappan K, Kayalvizi M. (2004). Enhanced Lipid Peroxidation and Impaired Enzymatic Antioxidant Activities in Erythrocytes of Patients with Cervical Carcinoma. Cell Mol Bio Lett. 9(4A): 699-07.
14. Moustafa A M, Ahamad S H, Hussein A A, Omran M A, Extraction and Phytochemical investigation of *Calotropis procera* effect of plant extract on the activity of diverse muscles. Pharm Biol., 2010; 1080-190.
15. Nihal Singh Verma, Sumeet Dwivedi, Debadash, S.K.Gupta. (2011), Antibacterial activity of root bark of *Nyctanthes arbor tristis*. International Journal of Drug Discovery and Herbal Research. 1(2): 61-62.
16. Notani P N. (2001). Global Variation in Cancer Incidence and Mortality. Current Science., 81: 465-74.
17. Pittaya, Kanokon, Walter C Taylor. (2013). Chemical Constituents from the Flowers of *Nyctanthes arbor tristis*. Science Asia, 29: 21-30.
18. Priya K, Ganjewala, Deepak. (2007). Antibacterial activity and phytochemical analysis of different plant parts of *Nyctanthes arbor tristis*. 1(2): 61-67.
19. Rahman, Shahedur, Salehin, Md. Fauzus, Jamal, Md. Abuhena Mostufa, Pravin, Anzana, Alam, Md. Khasrul. (2011). Antibacterial activity of *Argemone maxicana* against water born microbes. Research Journal of Medicinal Plants. 5(5): 621-626.
20. Saini Satish Chand, Reddy G B S, Birari Pankaj. (2013). *Murraya Koenigii*, I.O.R.S Journal of Pharmacy and Biological Sciences. 7(6): 15-18.

21. Sarvanan V Sakthi; Shanmugapandiyn P; Mahesh K. Antimicrobial activity of chloroform extract of leaves of *C. roseus*. Asian Journal of Chemistry, 2012; 24(7): 3126-3128.
22. Savita G Aggarawal, Sanjay Goyal. (2013). *Nyctanthes arbor tristis* against pathogenic bacteria, Journal of Pharmacognosy and Phytochemistry. 2(3): 124-127.
23. Singh Sarita, Singh Amitabh, Jaiswal Jyostana Singh T D, Singh V P, Pandey V B, Tiwari Aparna, Singh U P. (2010). Taylor and Francis Archives of Phytopathology and Plant Protection. 43(8): 769-774.
24. Vinoth B, Manivasagaperums R, Balamurugan S. (2012). Phytochemical Analysis and Antibacterial Activity of *Moringa oleifera*. International Journal of Research in Biological Sciences. 2(3): 98-102.
25. Yamanaka N, Fukushima M, Koizami K, Nishida K, Kato T, Ota K,. (1998), Enhancement of DNA Chain Breakage by Bleomycin and Biological Free Radicals Producing Systems,. Oxygen Biomembranes (New York) North Holland; 56-69.