

WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 6.805

Volume 5, Issue 6, 2021-2030. Research

Research Article

ISSN 2277-7105

IN-VITRO FREE RADICAL SCAVENGING ACTIVITY STUDIES OF EXTRACTS AND ISOLATED COMPOUNDS OF EUGENIA JAMBOLANA LAM. SEEDS

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Article Received on 19 April 2016,

Revised on 09 May 2016, Accepted on 30 May 2016

DOI: 10.20959/wjpr20166-6426

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ABSTRACT

Background: Eugenia jambolana seeds of Myrtaceae family are substantial ayurvedic medicine in treating diabetes. It is also narrated to have anti-inflammatory, hypoglycemic, antiulcerative, hypolipidemic, antiviral, ameliorative and antioxidant activities due to flavonoids, tannins, triterpenoids, saponins and glycosides. **Objective:** Various successive Soxhlet extractives [petroleum ether (PEE), CHCl₃ (CE), ethyl acetate (EAE) and methanol (ME)] and isolated compounds [IC1 and IC2] of E. jambolana seeds were inflicted to free radical scavenging activities employing DPPH and H₂O₂ radicals procedure which could be the main cause of degenerative diseases. Materials and Methods: All the extracts and isolated compounds were established to predicate good antioxidant activity against DPPH

and H_2O_2 radicals as compared to standard Ascorbic acid. PEE and CE showed very less antioxidant activity when compared to standard, EAE, ME, IC1 and IC2. Standard, PEE, CE, EAE, ME, IC1 and IC2 offered utmost DPPH radical scavenging activity of 88.49, 17.40, 22.59, 61.08, 72.12, 54.55 and 66.62 μ g/ml at concentration of 500 μ g/ml and for H_2O_2 scavenging, ultimate activity of 82.09, 24.25, 27.23, 67.61, 78.85, 60.53 and 71.36 μ g/ml at concentration of 50 μ g/ml. The antioxidant activity was extreme in ME and IC2. **Results:** The IC₅₀ of Standard, EAE, ME, IC1 and IC2 for DPPH radical and H_2O_2 radical was

pioneered 265, 360, 295, 380 and 220 μ g/ml and 26, 33, 27,37 and 27 μ g/ml respectively. **Conclusion:** The separation and identification of flavonoids, tannins, glycosides and triterpenoid saponins present in seeds of *E. jambolana* can help the researchers to detect new molecules as natural antioxidants.

KEYWORDS: Antioxidant, DPPH, *Eugenia jambolana*, H₂O₂, IC₅₀, Isolated compound.

INTRODUCTION

Antioxidants are substances, which help to defend the body against cell damage by various free radicals. Antioxidants are molecules that can neutralize free radicals by accepting or donating an electron to eliminate the unpaired condition. Free radicals are molecules containing unpaired electrons. Most of the free radicals are produced by mitochondria and most free radicals in biological system are derivatives of oxygen ("Reactive Oxygen Species" ROS), but there are also derivatives of nitrogen ("Reactive Nitrogen Species" RNS). Synthetic anti-oxidants, such as butylated hydroxyl toluene (BHT) and butylated hydroxyl anisole (BHA) have restricted use in food industry as they are suspected to be carcinogenic. Moreover, the synthetic anti-oxidants also show low solubility and moderate anti-oxidant activity. Therefore, many researchers are in search for anti-oxidants of natural origin. [1]

Eugenia jambolana Lam. (Synonym: Syzygium cumini Linn.) of family Myrtaceae, is widely spread in the plains from sub-Himalayas to South India^[2] and is a popular traditional medicinal plant in India. E. jambolana is prescribed for management of diabetes and several-marketed formulations contain the seeds of Eugenia jambolana as an ingredient. Seed powder is widely used in diabetes and urine related troubles. It is also used in indigestion, loss of appetite, vomiting and fever. It has been reported to have hypoglycemic, antidiabetic, ameliorative, antioxidant, anti-hyperlipidemic, antiviral, anti-inflammatory which may be due to the presence of flavonoids, saponins, glycosides and triterpenoids in the extract. ^[3-24] Therefore, our aim was to evaluate the free radical scavenging activity of various successive extracts and isolated compounds of E. jambolana seeds to determine its anti-oxidant properties and its uses in different diseases.

MATERIALS AND METHODS

Raw material characterization and preparation

The seeds of the plant *E. jambolana* were acquired from the forest region of Coimbatore District of Tamil Nadu, India. The seed material of the plant was ascertained by Agronomy

Dept., The Tamil Nadu Agricultural University, Coimbatore, India. The accrued material was dried under shade, powdered well with a mechanical grinder and stored in an air-tight container.

Chemicals

DPPH (1, 1-diphenyl-2-picryl-hydrazyl) was received from Sigma Chemicals, USA. The other chemicals and solvents of analytical reagent grade accomplished from S. D. Fine Chemicals Ltd., India, Qualigens Fine Chemicals Ltd., Mumbai, India and Ranbaxy Chemicals Ltd., New Delhi, India.

Extraction

The seed powder was condemned to successive extraction with petroleum ether, chloroform, ethyl acetate and methanol utilizing Soxhlet Extractor at 30-45 °C (5 days for each extraction).

Preliminary phytochemical evaluation

The preliminary phytochemical screening of the four consecutive extracts was executed to know the aberrant constituents existing in the seeds of *E. jambolana* as per the standard procedures. The extracts were tested for tannins, flavonoids, chlorophyll, alkaloids, glycosides and triterpenoids. The presence of flavonoids and tannins was confirmed by Thin layer chromatographic analysis using mobile phase as CHCl₃:CH₃COCH₃: HCOOH (75:16.5:8.5 %v/v/v) and detected under UV light. [28]

Isolation

Two unknown isolated compounds were recovered from ethyl acetate extract by Preparative TLC method using mobile phase as CHCl₃: CH₃COCH₃: HCOOH (75:16.5:8.5 %v/v/v) at the R_F values of 0.4 (IC1) and 0.9 (IC2). The isolated compounds were checked for purity (single spot/compound) by TLC, HPTLC and HPLC analysis. The spots were distinguished under UV light because the extract contains fluorescence compounds which avoid using spray reagents. The extracts and isolated compounds obtained were weighed and stored at -4 °C until further analysis.

DPPH free radical scavenging activity^[29]

Free radical scavenging potentials of PEE, CE, EAE and ME of *Ej* seeds and IC1 & IC2 were proven against a methanolic solution of DPPH.^[29] 0.1 mM solution of DPPH in methanol was

prepared and 1.0 ml of this solution was added to 3.0 ml of different concentrations (100-500 μ g/ml) of PEE, CE, EAE and ME of Ej seeds prepared in water. It was incubated at room temperature for 30 minutes and the absorbance was measured at 517 nm against the corresponding blank solution. Ascorbic acid was taken as reference. Percentage inhibition of DPPH free radical was calculated based on the control reading, which contained DPPH and distilled water without any extract using the following equation.

% Scavenging Activity = $[(Ac-As)/Ac] \times 100$

Where, Ac is the absorbance of the control reaction and As is the absorbance in the presence of the sample of the extracts. The antioxidant activity of the extract was expressed as IC_{50} . The IC_{50} value was defined as the concentration (in $\mu g/ml$) of extracts that inhibits the formation of DPPH radicals by 50%.

Scavenging of Hydrogen Peroxide^[30]

A solution of hydrogen peroxide (40 mM) was prepared in phosphate buffer (pH 7.4). The concentration of hydrogen peroxide was determined by recording the absorbance at 230 nm. Different concentrations of PEE, CE, EAE and ME of *Ej* seeds and IC1 & IC2 (10-50 μg/ml) in distilled water were added to a hydrogen peroxide solution (0.6 ml, 40 mM). The absorbance of hydrogen peroxide at 230 nm was determined after ten minutes against a blank solution containing phosphate buffer without hydrogen peroxide. The percentage of hydrogen peroxide scavenged by PEE, CE, EAE and ME of *Ej* seeds and IC1 & IC2 and ascorbic acid (standard) was calculated using following equation.

% Scavenged $[H_2O_2] = [(Ac - As)/Ac] \times 100$

The IC_{50} value was defined as the concentration (in $\mu g/ml$) of extracts that inhibits the formation of hydrogen peroxide radicals by 50%.

RESULTS AND DISCUSSION

DPPH free radical scavenging activity

DPPH scavenging activity has been used by various researchers as a quick and reliable parameter to assess the in-vitro antioxidant activity of crude plant extracts.^[31-32] DPPH is a stable free radical at room temperature and accepts an electron or hydrogen radical to become a stable diamagnetic molecule.^[33] The absorption maximum of a stable DPPH radical in methanol was at 517 nm. The decrease in absorbance of DPPH radical caused by

antioxidants, because of the reaction between antioxidant molecules and radical, progresses, which results in the scavenging of the radical by hydrogen donation.^[34]

Fig. 1 and fig. 2 epitomize DPPH scavenging activity of PEE, CE, EAE and ME and of IC1 and IC2 are compared with standard Ascorbic Acid. Table No.1 emphasizes % maximum antioxidant activity and IC₅₀ values of standard, PEE, CE, EAE, ME, IC1 and IC2.

Scavenging of Hydrogen Peroxide

Although hydrogen peroxide itself is not very reactive, it can sometimes cause cytotoxicity by giving rise to hydroxyl radicals in the cell. Thus, removing H_2O_2 is very important throughout food systems. Scavenging of H_2O_2 by antioxidants may be due to donation of electrons to H_2O_2 , thus neutralizing it to water.^[35]

Fig. 3 and fig. 4 delineate H_2O_2 scavenging activity of PEE, CE, EAE and ME and of IC1 and IC2 are compared with standard Ascorbic Acid. Table No. 2 annotates % maximum antioxidant activity and IC₅₀ values of standard, PEE, CE, EAE, ME, IC1 and IC2.

Figures

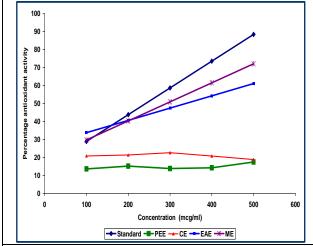


Fig. 1 - DPPH Scavenging Activity of PEE,

CE, EAE and ME are compared with a

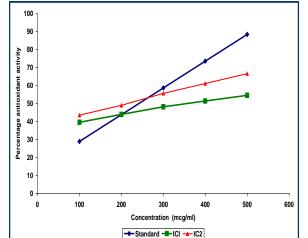


Fig. 2 - DPPH Scavenging Activity of IC1 and IC2 are compared with Standard Ascorbic acid

Standard Ascorbic Acid

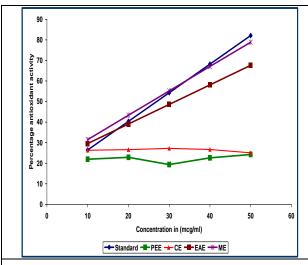


Fig. 3 - Hydrogen Peroxide Scavenging Activity of PEE, CE, EAE and ME are compared with Standard Ascorbic Acid

Fig. 4 - Hydrogen Peroxide Scavenging Activity of IC1 and IC2 are compared with Standard Ascorbic Acid

Table No. 1 - % Maximum DPPH free radical scavenging activity and IC_{50} of Standard, PEE, CE, EAE, ME, IC1 and IC2

| S. No. | Sample | % maximum Antioxidant activity [%] | IC ₅₀ |
|--------|----------|------------------------------------|------------------|
| 1 | Standard | 88.4878 | 265 |
| 2 | PEE | 17.3969 | _ |
| 3 | CE | 22.5945 | _ |
| 4 | EAE | 61.0825 | 360 |
| 5 | ME | 72.1219 | 295 |
| 6 | IC1 | 54.5533 | 380 |
| 7 | IC2 | 66.6237 | 220 |

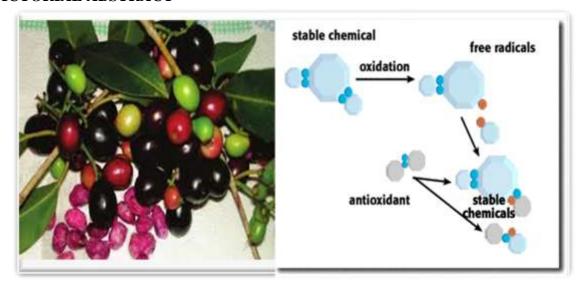
Note: PEE – Petroleum ether extract; CE – Chloroform extract; EAE – Ethyl acetate extract; ME – Methanol extract; IC1 – Isolated compound 1; IC2 – Isolated compound 2; IC $_{50}$ – 50% Inhibitory concentration.

Table No. 2 - % Maximum H_2O_2 free radical scavenging activity and IC_{50} of Standard, PEE, CE, EAE, ME, IC1 and IC2

| S. No. | Sample | % maximum Antioxidant activity [%] | IC ₅₀ |
|--------|----------|------------------------------------|------------------|
| 1 | Standard | 82.0938 | 26 |
| 2 | PEE | 24.2550 | |
| 3 | CE | 27.2350 | _ |
| 4 | EAE | 67.6082 | 33 |
| 5 | ME | 78.8546 | 27 |
| 6 | IC1 | 60.5338 | 37 |
| 7 | IC2 | 71.3656 | 27 |

Note: PEE – Petroleum ether extract; CE – Chloroform extract; EAE – Ethyl acetate extract; ME – Methanol extract; IC1 – Isolated compound 1; IC2 – Isolated compound 2; IC $_{50}$ – 50% Inhibitory concentration

PICTORIAL ABSTRACT



CONCLUSION

The preliminary phytochemical investigation and TLC studies indicate the presence of flavonoids, tannins, triterpenoid saponins and glycosides in seeds of *E. jambolana* Lam. Polyphenols like flavonoids and tannins are the well known natural antioxidants. So, the antioxidant potential of extracts and isolated compounds may be due to the presence of flavonoids and tannins. The results of the present study show that the extracts and isolated compounds of seeds of *E. jambolana* Lam. possess good antioxidant activity when compared with standard Ascorbic acid. The antioxidant activity was highest in methanolic extract and IC2. PEE and CE showed very less antioxidant activity when compared with other extracts (EAE and ME) standard and isolated compounds. Hence, *E. jambolana* Lam. seeds can be used as the potential source for natural antioxidants. Characterization and formulation development of IC1 and IC2 can be used in modern medicine for treating the diseases caused by free radicals.

ACKNOWLEDGEMENT

We capitulate our gratitude and admiration to the Management and the Principal of College of Pharmacy, Sri Ramakrishna Institute of Paramedical Sciences, Coimbatore, for the proficiency provision for presiding over this research work. We surrender indebted recognition and reverence to the Principal and the Management of Karpagam College of Pharmacy, Coimbatore, for the great espouse to publish this investigation.

REFERENCES

- 1. Dhrubo Jyoti Sen. Herbal antioxidants: A great hope for future. Pharma Times., 2008; 40(12): 20 -21.
- 2. Kapoor LD. Handbook of Ayurvedic Medicinal Plants, CRC press, Boca Raton, London, New York: Washington, D. C., 2005; 179 -180.
- 3. Grover JK, Vats V and Rathi SS. Anti-hyperglycemic effect of *Eugenia jambolana* and *Tinospora cordifolia* in experimental diabetes and their effects on key metabolic enzymes involved in carbohydrate metabolism. Journal of Ethnopharmacology., 2000; 73(3): 461-470.
- 4. Pepato MT, Folgado VBB, Kettelhur IC and Brunetti IL. Lack of antidiabetic effect of a *Eugenia jambolana* leaf decoction on rat streptozotocin diabetes. Brazilian Journal of Medical and Biological Research., 2001; 34(3): 389 -395.
- 5. Timbola AK, Szpoganicz B, Branco A, Monache FD and Pizzolatti MG. A new flavonol from leaves of *Eugenia jambolana*. Fitoterapia., 2002; 73: 174 -176.
- 6. Madhuri Pandey and Aqueel Khan. Hypoglycemic effect of defatted seeds and water soluble fibre from the seeds of *Syzygium cumini* (Linn.). *Indian Journal of* Experimental Biology., 2002; 40: 1178 -1182.
- 7. Grover JK, Rathi SS and Vats V. Amelioration effect of experimental diabetic neuropathy and gastropathy in rats following oral administration of plant (*Eugenia jambolana*, *Mucuna pruriens and Tinospora cordifolia*) extracts. Indian Journal of Experimental Biology., 2002; 40: 273 -276.
- 8. Ravi K, Sivagnanam K and Subramanian S. Anti-diabetic activity of *Eugenia jambolana* seed kernels on Streptozotocin-induced diabetic rats. Journal of Medicinal Food., 2004; 7(2): 181 -191.
- 9. Kasi Ravi, Balasubramanian Ramachandran and Sorimuthu Subramanian. Protective effect of *Eugenia jambolana* seed kernel on tissue antioxidants in Streptozotocin-induced diabetic rats. Biological & Pharmaceutical Bulletin., 2004; 27(8): 1212-1217.
- 10. K. Ravi, D. S. Sekar and S. Subramanian. Hypoglycemic activity of inorganic constituents in *Eugenia jambolana* seed on Streptozotocin-induced diabetic rats. Biol Trace Elem Res., 2004; 99(1-3): 145-155.

- 11. Sridhar SB, Sheetal UD, Pai MR and Shastri MS. Preclinical evaluation of the antidiabetic effect of *Eugenia jambolana* seed powder in Streptozotocin-induced diabetic rats. Braz J Med Biol Res., 2005; 38(3): 463-468.
- 12. Kasiappan Ravi, Subbaih Rajasekaran and Sorimuthu Subramanian. Antihyperlipidemic effect of *Eugenia jambolana* seed kernel on Streptozotocin-induced diabetic rats. Food and Chemical Toxicology., 2005; 43(9): 1433 -1439.
- 13. Mutalik S, Chetana M, Sulochana B, Uma Devi P and Udupa N. Effect of dianex, a herbal formulation on experimentally induced diabetes mellitus. Phytotherapy Research., 2005; 19(5): 409 -415.
- 14. Teixeira MD CC, Fuchs FD, Weinert MD LS and Esteves MD J. The efficacy of folk medicines in the management of type 2 diabetes mellitus: results of a randomized controlled trial of *Syzygium cumini* (L.) Skeels. Journal of Clinical Pharmacy and Therapeutics., 2006; 31(1): 1-5.
- 15. Kotowaroo MI, Mahomoodally MF, Gurib/Fakim A and Subratty AH. Screening of traditional antidiabetic medicinal plants of *Mauritius* for possible alpha/ amylase inhibitory effects in vitro. Phytother Res., 2006; 20(3): 228 -31.
- 16. Bala SS, Afreena N, Madhava PK and Suryanarayana MP. Antihyperglycemic effect of the fruit-pulp of *Eugenia jambolana* in experimental diabetes mellitus. Journal of Ethnopharmacology., 2006; 104(3): 367 -373.
- 17. Singh N and Gupta M. Effects of ethanolic extract of *Syzygium cumini* (Linn) seed powder on pancreatic islets of alloxan diabetic rats. Indian Journal of Experimental Biology., 2007; 45(10): 861 -867.
- 18. Aditi Chaturvedi, Mohan Kumar M, Bhawani G, Harish Chaturvedi, Mohan Kumar and Goel RK. Effect of ethanolic extract of *Eugenia* jambolana seeds on gastric ulceration and secretion in rats. Indian Journal of Physiol Pharmacol., 2007; 51(2): 131 -140.
- 19. Bhavna Sharma, Viswanath G, Rajani Salunke and Partha Roy. Effects of flavonoid-rich extract from seeds of *Eugenia jambolana* (L.) on carbohydrate and lipid metabolism in diabetic mice. Food Chemistry., 2008; 110(3): 697 -705.
- 20. Bhavna S, Chandrajeet B and Partha R. Hypoglycemic and hypolipidemic effecs of flavonoid rich extract from *Eugenia jambolana* seeds on Streptozotocin induced diabetes rats. Food and Chemical Toxicology., 2008; 46(7): 2376 -2383.
- 21. Helmstadter. *Syzygium cumini* (L.) Skeels (Myrtaceae) against diabetes 125 years of research. Die Pharmazie., 2008; 63(2): 91 -101.

- 22. Bhanuprakash V, Hosamani M, Balamurugan V, Gandhale P, Naresh R, Swarup D and Singh RK. *In-vitro* antiviral activity of plant extracts on goatpox virus replication. Indian Journal of Experimental Biology., 2008; 46(2): 120 -127.
- 23. Kumar A, Ilavarasan R, Jayachandran T, Deecaraman M, Mohan Kumar R, Arvindan P, Padmanabhan N and Krishan MRV. Anti-inflammatory activity of *Syzygium cumini* seed. African Journal of Biotechnology., 2008; 7(8): 941 -943.
- 24. Sharma SB, Reenu, Nasir A, Prabhu KM and Murthy PS. Ameliorative effect of herbal compound isolated from seeds of *Eugenia jambolana* on diabetic dyslipidemia. Clinical Chemistry & Laboratory Medicine., 2008; 46: Supplement 1: S7.
- 25. Khandelwal KR, Kokate CK, Pawar and Gokhale. Practical Pharmacognosy. Nirali Prakashan., October 1996; 27, 102, 106, 119, 145, 154.
- 26. Rasheeduz Zafar Neerja Gandhi. Practical Pharmacognosy. CBS Publications., 1994; 38-39.
- 27. Kokate CK, Purohit AP and Gokhale SB. Pharmacognosy. 24th ed. Nirali Prakashan., September 2003; 197.
- 28. Wagner H, Bladt S and Zgainski EM. Plant Drug Analysis. Springer Verlag. Berlin Heidelberg. New York: Tokyo., 1984; 118: 163-173.
- 29. Sharma SK and Gupta VK. *Invitro* antioxidant studies of *Ficus racemosa* Linn. root. Phcog Mag., 2008; 4(13): 70 -74.
- 30. Ruch RJ, Cheng SJ and Klaunig JE. Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chines green tea. Carcinogenesis., 1989; 10: 1003 -1008.
- 31. Navarro MC, Montilla MP, Martin A, Jimenez J and Utrilla MP. Free radicals and antihepatotoxic activity of *Rosmarinus tomentosus*. Planta Med., 1992; 59: 312 -14.
- 32. Thabrew MI, Hughes RD and Mc Farlane IG. Antioxidant activity of *Osbeckia aspera*. Phytother. Res., 1998; 12: 288 -90.
- 33. Soares JR, Dins TCP, Cuaba AP and Ameida LM. Antioxidant activity of some extracts of *Thymus cygi*. Free Rad. Res., 1997; 26: 469 -478.
- 34. Chang LW, Yen WJ, Haung SC and Duh PD. Antioxidant activity of sesame coat. Food Chem., 2002; 78: 347 -354.
- 35. Elmastas M, Gulcin I, Kufrevioglu OI, Ibaoglu K and Aboul-Enein HY. Radical scavenging activity and antioxidant capacity of Bay leaf extracts. J. Iran. Chem. Soc., 2006; 3(3): 258 -266.