

UNLOCKING THE POWER OF PHYTOCHEMICALS IN PARKINSON'S DISEASE: BRIDGING CURRENT INSIGHT TO FUTURE THERAPEUTICS INNOVATIONS

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ABSTRACT

Phytochemicals have emerged as promising candidates in neurodegenerative disorder therapeutics, particularly parkinson's disease (PD). PD is characterized by the progressive degeneration of dopaminergic neurons and the aggregation of α -synuclein, leading to motor and non-motor symptoms. Current treatments like Levodopa, primarily address symptoms without halting disease progression or neuronal degeneration. This review highlights the neuroprotective potential of various naturally occurring phytochemicals, such as chrysin, daphnetin and theaflavin, etc. These compounds exhibit antioxidant, anti-inflammatory, and signaling modulation properties, targeting pathways like Nrf2, NF- κ B, and PI3K/Akt. The mechanisms of action, such as oxidative stress mitigation, mitochondrial function enhancement and autophagy regulation, offer a multi-faceted approach to combating PD. Furthermore, by investigating numerous experimental models that assess the potential neuroprotective efficiency of antioxidant phytochemical derivatives in terms of their

inhibitory effects on oxidative stress and neuroinflammation in the brain. Naturally derived antioxidant phytochemicals may be considered prospective pharmaceutical therapeutic options to alleviate symptoms or reduce the progression of parkinson's disease. However, additional well-designed clinical trials are needed to assess the preventive and therapeutic advantages of phytochemicals as prospective medications in the treatment of Parkinson's

disease. By bridging existing knowledge with therapeutic innovation, this study underscores the role of phytochemicals in developing future interventions for PD.

KEYWORDS: Parkinson's Disease, Phytochemicals, Neurodegeneration, Neuroprotection, α -synuclein.

1. INTRODUCTION

Parkinson's disease (PD) is a progressive, chronic neurodegenerative movement disorder, with prevalence increasing with age. In the Western world, PD affects approximately 315 per 100,000 people of all ages, and this prevalence is projected to double by 2030, significantly contributing to global mortality, morbidity, and socioeconomic burden.^[1] The clinical symptoms commonly associated with PD include bradykinesia, resting tremor, postural instability, rigidity, depression, and anxiety. A key hallmark of PD is the progressive degeneration of dopaminergic neurons in the substantia nigra pars compacta (SNpc), leading to dopamine (DA) depletion in the striatum (ST)—a major factor underlying the motor impairments characteristic of the disease.^[2] Beyond the dopaminergic system, other neuropathological processes involve nondopaminergic systems with some overlapping with Alzheimer's disease (AD) pathology. These include cholinergic dysfunction and disruptions in serotonergic, glutamatergic, and noradrenergic pathways, all contributing to dopaminergic neuronal death and/or DA system dysfunction. PD is also associated with a range of nonmotor symptoms, such as sleep disturbances, cognitive changes, autonomic dysfunction, mood alterations, depression, fatigue, and pain.^[3]

Since the aggregation of misfolded α -synuclein is a key component of Lewy bodies, a significant clinical characteristic of Parkinson's disease, PD is referred to as a synucleinopathy. Furthermore, α -synuclein has a special role in the etiology of Parkinson's disease and seems to be connected to both familial and sporadic forms of the illness.^[4] Remarkably, a number of neurotoxin pathways, such as posttranslational modifications, neuroinflammation, oxidative stress, mitochondrial dysfunction, altered mitochondrial morphology, synaptic dysfunction, phospholipids, induced endoplasmic reticulum (ER) stress and metal ions, have been broadly associated with α -synuclein accumulation. Misfolded α -synuclein may be responsible for the aging process in Parkinson's disease. Age-related failure of the antioxidant defense system and excessive production of ROS worsen oxidative stress in the brain.^[5]

Although, levodopa (L-dopa) is the most effective treatment for PD's early-stage motor symptoms, it is not thought to be a cure for the disease. Tremor might only be marginally lessened, although bradykinesia and stiffness react the best. Balance issues and other symptoms could not get any better. L-dopa, however, is ineffective in treating Lewy pathology, nonmotor symptoms, or neuronal loss. Patients eventually need larger dosages of L-dopa, which are linked to more adverse effects such as dyskinesia.^[6,7] By effectively scavenging oxygen free radicals and enhancing the cellular antioxidant defense system and associated molecules, plant products and their bioactive phytochemicals can shield cells from oxidative damage. These antioxidant phytochemicals have been shown in multiple studies to have neuritogenic potential, reestablishing synaptic connectivity by reversing the loss of neuronal processes. By focusing on many mechanisms beyond those previously described, a variety of antioxidant phytochemicals have demonstrated potentially neuroprotective qualities. Biologically active substances known as phytochemicals typically match secondary metabolites found in plants, such as terpenoids, flavonoids, and alkaloids.^[8]

Moreover, phytochemicals inhibit nuclear factor kappa B (NF- κ B) pathways and aid in the activation of the phosphatidylinositol 3-kinase/protein kinase B (PI3K/Akt) and extracellular signal-regulated kinase (ERK) pathways.^[9] This study provides information about the neuroprotective properties and mechanisms of action which recently identified naturally occurring phytochemicals that alter the course of Parkinson's disease at the cellular and molecular level to combat oxidative stress and neurodegeneration. Therefore, we explored the pharmacological innovative targets of various phytochemicals involved in neurodegeneration of parkinsons disease.^[10,11]

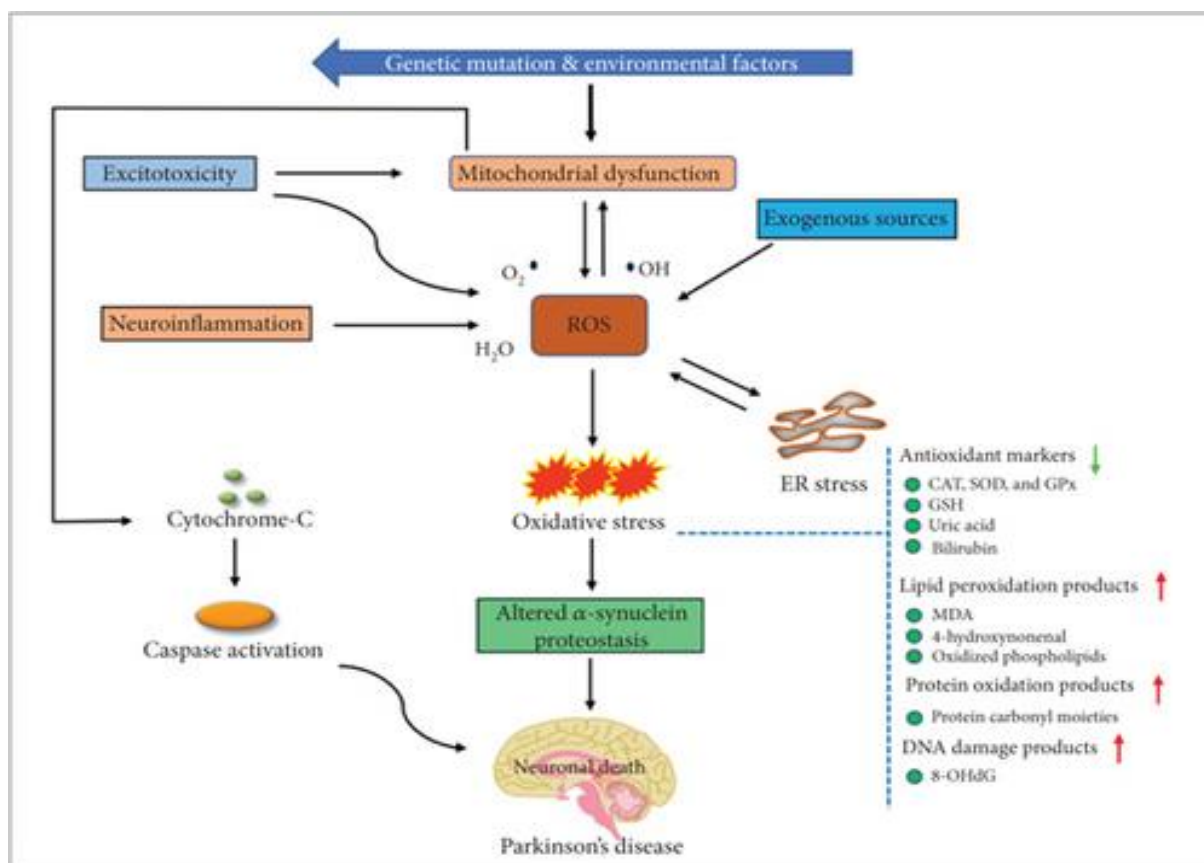


Figure 1: The pathogenesis of parkinson's disease.

Brain cells are susceptible to oxidative stress when ROS creation out spaces intracellular antioxidant defenses. In addition, earlier studies have demonstrated that increased ROS production causes significant and simultaneous dysregulation of several signaling pathways, such as RAS-MEK-ERK1/2,^[12] PI3K/AKT/GSK3 β ,^[13] Keap1-Nrf2-ARE,^[14] NF- κ B.^[15]

In recent decades, a number of scientific findings have shown that Nrf2 is extensively involved in the regulation of various physiological functions through its antioxidant, anti-inflammatory, autophagic, detoxifying, and proteasomal actions.^[16] Normally, these targets are thought of as antioxidant genes.^[17]

2. TARGETS OF PHYTOCHEMICALS IN PARKINSONS DISEASE

1) Chrysin

Chrysin is a naturally occurring polyphenolic substance called a flavonoid; flavonoids are found in a wide variety of foods, including fruits, vegetables, blue passion flowers, mushrooms, plants, and honey.^[11] Chrysin's neuroprotective effects, which stem from its anti-inflammatory, antioxidant, and other pharmacological characteristics, have been investigated.^[12] Chrysin's neuroprotective properties demonstrate that dopaminergic neuronal

death is inversely correlated with increases in dopamine (DA) levels in both in vitro and in vivo investigations.^[13,14] Additionally, as assessed by the Barnes maze, beam walk, horizontal and vertical grid, rotating behavior, and passive avoidance tests, chrysin treatment greatly reduced the animals' motor impairment and cognitive dysfunction. Chrysin thus shows promise as a disease-modifying substance that could also help in alleviate Parkinson's disease symptoms.^[15] Krishnamoorthy *et.al.* studied the effect of chrysin on mice which indicated the reduced dopaminergic neuronal loss in the SN area in an MPTP-based experimental paradigm. Chrysin protected against dopaminergic neurodegeneration and enhanced locomotor activity in MPTP-injected PD animal models, according to a fairly recent study.^[16] Krishnamoorthy *et.al.* and Del Fabbro *et.al.* investigated that there is an increase DA levels in MPP+ and MPTP-treated CGN cells as well as mice models of parkinson's disease by functioning as an inhibitor of Monoamine Oxidase B (MAO-B). Chrysin also raised the levels of DA, 3,4-dihydroxyphenylacetic acid (DOPAC), and homovanillic acid (HVA), according to a number of neurotoxin-induced in vivo tests.^[17] By suppressing proapoptotic proteins like caspase-3, caspase-9, and Bax and increasing the expression of the antiapoptotic protein Bcl-2 against MPP+ neurotoxicity, Guo B *et.al.* showed the neuroprotective effect at the molecular level. Furthermore, in 6-OHDA- and MPTP-induced PD mice models, chrysin therapy produced neuroprotectivity by controlling or reestablishing BDNF and glial cell-derived neurotrophic factor (GDNF) levels in the ST area.^[18] Therefore, chrysin's many protective functions in parkinson's disease, including neuroprotective and symptom-relieving, are very valuable and could lead to new avenues for innovative therapeutic management of PD; more clinical research to be required. Chrysin has antiviral and antibacterial properties in addition to parkinson's disease.^[19]

2) Daphnetin

Among the coumarins included in daphne are daphnetin, DAP-8-glucoside, daphnin, esculin, umbelliferone, and acetyl-umbelliferone. Although Daphne species also had trimeric coumarin metabolites such as triumbellin and daphneretusin B, dimeric coumarins include rutarensin, daphnoretin, daphneretusin-A, and dimethyl-daphnoretin-7-O-glucoside.^[20] Fan X *et.al.* showed the nephroprotective effect against cisplatin-induced nephrotoxicity by activating the Nrf2 pathway and suppressing the NF- κ B signaling pathway.^[21] Reactive oxygen species (ROS) and inflammatory signaling molecules such as interleukin (IL)-6 and tumor necrosis factor alpha (TNF- α) are the primary causes of neurodegenerative disorders. It also inhibits JAK/STAT activation, which decreases levels of COX-2 and inducible nitric oxide synthase

(iNOS) and increases pro-inflammatory cytokines and enzymes. It significantly enhances Nrf-2 expression was noted by Vinayagam R et.al.^[22] According to the in vitro investigation on neuronal-like rat pheochromocytoma PC12, DAP increases heat shock protein (HSP)-70 by molecularly downregulating the expression of NF- κ B and mitogen-activated protein kinase (MAPK). By modifying the phosphorylation of pro-apoptotic proteins and an anti-apoptotic protein (Bax/Bcl-2), these enzymes subsequently regulate neuronal death.^[23] Fever, lumbago, cancer, rheumatoid arthritis (RA), coagulation problems, and various skin disorders have all been treated with the DAP was reported by Qi Z et.al.^[24] Among its many pharmacological qualities were analgesic,^[25] antipyretic,^[26] anti-arthritis,^[27] anti-inflammatory, antioxidant, anti-proliferative, antibacterial, and anti-inflammatory.^[28]

3) Theaflavin

A class of polyphenols known as theaflavin (TF), which comprises theaflavin-3-gallate, theaflavin-3'-gallate, and theaflavin-3,3'-digallate, affects the quality and color of black tea.^[29] Numerous recent scientific publications have suggested that TF may have neuroprotective properties against NDDs. TF has been shown to be equally efficient as EGCG in reducing neurotoxicity caused by β -amyloid and α -synuclein due to its potential antioxidant properties.^[30] Recently, a randomized, double-blind, placebo-controlled study was carried out.^[31] Luo Z et.al. investigated that the cell lines were used to investigate the mechanism of action of TF, 6-OHDA-induced SH-SY5Y. The findings revealed reduced nuclear morphology, reduced apoptosis, elevated MMPs, decreased intracellular NO levels, and lessened cell viability loss. These results demonstrated that TF protected against 6-OHDA-induced apoptosis by preventing the production of NO and ROS.^[32] Grelle G et.al. studied that oxidative stress was eliminated by giving TF to the PC12 cell line treated with H₂O₂, which led to an increase in Bcl-2 expression and a decrease in the production of the proteins Bax and caspase-3. This finding implies that TF possesses antiapoptotic properties that provide cytoprotection as well as neuroprotection.^[33] According to a different study, TF acted as a potent inhibitor of β -amyloid and α -synuclein fibrillogenesis and promoted the innocuous assembly of these proteins. These results suggest that TF may remove dangerous amyloid buildup.^[34] TF increased the expression of DAT and VMAT-2 in the PD animal model and decreased the effects of oxidative stress in MPTP-induced neurotoxicity in mice. Additionally, TF has been shown to ameliorate dopaminergic neuronal degeneration and behavioral deficits.^[35] Anandhan A et.al. investigated those mice treated with MPTP/p, TF increased nigral TH and DAT expression and decreased caspase-3, caspase-8, and caspase-9;

these effects were linked to increases in regulated behavioral function.^[36] Additionally, it was discovered that proinflammatory cytokine production and microglia activation were also brought on by cholinergic system dysfunction. The anti-inflammatory markers IL-4 and IL-10 were more abundant in MPTP-injected rats as a protective measure against neural inflammation.^[37] According to a recent *in vivo* study, TF treatment significantly decreased neuroinflammation and apoptosis, hence reducing chronic MPTP-induced neurotoxicity in mice's SN and ST studied by the Szuster-Ciesielska A *et.al.*^[38,39,40] Furthermore, TF therapy significantly reduced the overproduction of IL-4 and IL-10 and lessened the behavioral abnormalities brought on by MPTP injection, such as catalepsy and akinesia. Anti-inflammatory, anti-influenza, cardioprotective, and anti-cancer capabilities are among the other biological actions that have been discovered.^[41,42]

4) Epigallocatechin-3-Gallate

Epigallocatechin-3-gallate (EGCG), a polyphenol found in green tea, has been demonstrated to have a variety of physiologically beneficial effects on cancer, non-communicable illnesses, and inflammatory disorders in humans.^[43–45] By altering several transcription factors, proteins, and other important growth factors, EGCG has been connected to a variety of pharmacological properties, including anti-inflammatory, antioxidant, metal-chelating, radical-scavenging, antiapoptotic, and anticarcinogenic properties.^[46, 47] EGCG has many potential advantages for supporting healthy aging since it promotes the morphologic and functional changes that occur naturally in an aging brain. These alterations include improving learning and memory, lessening brain oxidative damage, and preventing cognitive dysfunction.^[48–50] Green tea consumption and cognitive impairments in parkinson's disease disorders have been linked in a number of human studies to an inverse dose-response association.^[51,52] EGCG therapy enhanced mitochondrial function, increased succinate dehydrogenase (SDH) activity, decreased lipid peroxidation production and NO levels, and elevated ATPase and ST catecholamine levels in rotenone-induced rat PD models. Furthermore, EGCG administration enhanced motor function and reduced the levels of apoptotic markers and neuroinflammatory cytokines.^[53] When EGCG was given to MPTP-induced animals, the neurotoxin-induced decrease in the ST antioxidant enzymes SOD and CAT was stopped, and the overall brain activity of both enzymes was elevated.^[54] Additionally, in SH-SY5Y cells, EGCG administration completely inhibited STAT3 activity and promoted the growth of neuronal cells triggered by 6-OHDA.^[55] According to the most current research, EGCG's neurorescue action inhibited functional and neurochemical deficits

in MPTP-induced PD rats, decreased oxidative stress, and controlled the iron-export protein ferroprotein in the SN.^[56] Levites Y et.al. found that EGCG preserved TH-positive cells in the SN area and restored movement behavior in the MPTP-injected animal model. EGCG administration increased the ratio of CD3⁺CD4⁺ to CD3⁺CD8⁺ T-cell lymphocytes in the peripheral blood and decreased the expression of inflammatory markers such TNF- α and IL-6 in the serum, according to flow cytometric analysis.^[57] Wang L et.al, indicated that the additionally, effect in PQ-induced PC12 cell models, EGCG demonstrated antiapoptotic actions by maintaining MMP and preventing the elevation of caspase-3 activity and the downregulation of the proapoptotic SMAC protein in cytosol expression.^[58]

It was also observed that EGCG treatment reduced TNF- α and NO inflammatory mediators and attenuated loss of midbrain DA levels triggered by LPS-induced neurotoxicity.^[59] Hou RR et.al. conducted both in vitro and in vivo and showed that cotreatment with EGCG reduced glutamate-induced oxidative cytotoxicity in HT22 cells by inhibiting the activation of NF- κ B. Furthermore, rats administered with the carbidopa and L-dopa showed less buildup of 3-O-methyldopa in their plasma and ST after receiving EGCG treatment; EGCG also had a potent therapeutic impact on PD animals' hippocampal oxidative neuronal death caused by kainic acid.^[60] Many neurodegenerative disease, including parkinson's disease, have been closely associated with the process of α -synuclein and other protein deposition.^[61] EGCG may interact with α -synuclein amino acid positions on peptide membranes, according to other studies. These results support the idea that EGCG may be a powerful remodeling agent of α -synuclein accumulation and a possible disease-modifying medication for the treatment of parkinson's disease. It was suggested that EGCG binds to α -synuclein via unstable hydrophobic interactions.^[62,63,64]

5) Alpha and beta asarone

Alpha- (α -) asarone and beta- (β -) asarone compose an important antioxidant aromatic chemical constituent that is extracted from the rhizomes of *Acorus calamus*. Consequently, both α - and β -asarone have been reported to have one or more similar pharmacological properties that may offer beneficial effects in the therapeutic management of several diseases.^[65] Importantly, the delivery of α - and β -asarone in the brain is extensive, demonstrating its ability to cross the BBB, a desirable characteristic of compounds used for the treatment of numerous neurodegenerative diseases.^[66] Lu J et.al., studied the effect in PD model, α -asarone treatment reduced neural inflammation and suppressed IL- β , IL-6, and

TNF- α production in LPS-stimulated BV-2 cells. In addition, α -asarone treatment effectively inhibited the LPS-stimulated activation via regulation of NF- κ B by blocking degradation of inhibitor NF- κ B signaling in BV-2 microglial cells. In vivo studies also demonstrated that prophylactic administration with α -asarone inhibited microglial activation and attenuated PD-like behavioral deficits in MPTP-injected PD mice.^[67] Hei X et.al. investigated the effect in the 6-OHDA-induced PD model, β -asarone improved the behavioral function of rats in the initiation time, open field, stepping time, and rotarod tests. Research has also found that β -asarone increases the levels of HVA, DOPAC, and 5-HIAA in the ST region. In addition, administration with β -asarone elevated the level of TH-positive neurons and inhibited the expression of LC3-II in SN4741 cells. Moreover, *in vivo* experimental results showed that β -asarone affected the expression of Bcl-2, Beclin-1, JNK, and p-JNK in 6-OHDA-injected PD rats. The neuroprotective effect of β -asarone occurs primarily by downregulating JNK and p-JNK expressions and then indirectly increasing Bcl-2 expression. Additionally, β -asarone may inhibit the function of Beclin-1, thereby inhibiting autophagy activation.^[68] Activated autophagy is an important process that may play a defensive role through clearance of toxic aggregated α -synuclein in neurons.^[69] On the other hand, dysfunction of the autophagy-lysosomal pathway has been associated with the development of PD.^[70] Fang YQ et.al. showed that endoplasmic reticulum (ER) stress may induce autophagy.^[71] Zhang S et.al. investigated the effect in 6-OHDA-induced PD rat models, β -asarone administration may decrease the levels of Beclin-1, CHOP, GRP78, and p-PERK while significantly increasing the level of Bcl-2. β -Asarone may increase Bcl-2 by inhibiting the p-ERK pathway, and Bcl-2 may inhibit the expression of Beclin-1. The results of the study suggested that β -asarone may regulate autophagy and ER stress via the PERK/CHOP/Bcl-2/Beclin-1 pathway.^[72] Kuma A et.al. reported the β -asarone can effectively inhibit neuronal apoptosis through the CaMKII/CREB/Bcl-2 signaling pathway and regulate Bcl-2 family proteins.^[73] Pan T et.al. documented that β -asarone significantly lowered the expression levels of MALAT1 and α -synuclein in the midbrain of MPTP-injected PD mice. In addition, immunoprecipitation and RNA pull-down assays confirmed that MALAT1 was associated with α -synuclein, leading to the increased stability of α -synuclein and its expression in SH-SY5Y cells. β -Asarone treatment could increase the viability of cells exposed to MPP⁺.^[74] Jheng JR et.al. demonstrated that β -asarone exerted antioxidative effects on H₂O₂-stimulated PC12 cells by reducing oxidative stress via activation of the protective Nrf2/HO-1 pathway.^[75] The documented other biological activities are antifungal,^[76] anthelmintics,^[77] for the treatment of myocardial ischemia,^[76] anticancer.^[77]

6) Baicalein

Baicalein (5,6,7-trihydroxyflavone; $C_{15}H_{10}O_5$) is an important flavonoid compound mainly isolated from the roots of *Scutellaria baicalensis*.^[78] Ning B et.al. reported that baicalein effectively protected glutamate, amyloid- β (A β), 6- hydroxydopamine (6-OHDA), hydrogen peroxide (H_2O_2), 1-methyl-4-phenylpyridinium(MPP⁺), 1-mehtyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) and methamphetamine-induced neurotoxicity in cell line and animal models.^[79] baicalein protects mitochondrial functions by modulating mitochondrial membrane potentials, Bcl2/Bax ratio and reducing oxidative stress.^[80] In addition, baicalein can exerts its anti-inflammatory action by inhibiting glial activation as well as reducing pro-inflammatory enzymes and cytokines, including interleukin (IL)-1 β .^[81] IL-1 β is known to be activated by caspase 1, a pro-inflammatory caspase activated during the inflammasome activation.^[82] Hung KC et.al. summarized the therapeutical actions of baicalein against alzheimer's and parkinson's diseases. Baicalein effectively prevents alzheimer's and parkinson's diseases by reducing oxidative stress, inhibiting aggregation of disease-specific amyloid proteins, inhibiting excitotoxicity, stimulating neurogenesis, and anti-apoptosis as well as anti-inflammatory properties.^[83] Mu X et.al. reported the neuronal cell damage or death is the most important factor for various neurodegenerative disorders. Oxidative stress, neuroinflammation, misfolding of protein and mitochondrial dysfunction are the major pathways responsible for neurodegeneration. Ju XN et.al. investigated that the several transcription factors play an important role in the pathophysiology of neuronal cell damage including nuclear factor erythroid-derived 2 (NF-E2) related factor (Nrf2), nuclear factor-kappa B (NF- κ B), mitogen-activated protein kinases (MAPKs), cAMP-response element binding protein (CREB), Wnt, janus kinase/signal transducer and activator of transcription and TLR-4, etc.^[84] In this connection, the main focus of neuroprotective agents is to protect neuronal cell damage/death for preventing the neurodegenerative diseases.^[85,86] The reported other biological activities like antibacterial,^[87] antiviral,^[88] anti-inflammatory,^[89] antitumor,^[90] cardiovascular^[91] and neuroprotective activities.^[92]

7) Betulin

The naturally occurring triterpene, betulin (lup-20(29)-ene-3 β ,28-diol) is mostly found in shrubs and trees and other forms of the main extractive (up to 30% of dry weight) of birch tree bark.^[93] The triterpene found in nature is betulin. It is frequently separated from birch tree bark. Up to 30% of the dry weight of silver birch bark is made up of it.^[94] Ju XN et.al. studied the neuroprotective roles of betulin in 6-OHDA-induced DA neuron degeneration,

food-sensing behavior defects, and life span which may be related to its antioxidant and antiapoptotic activity. It can affect pathways in parkinson's disease by upregulating the Akt/Nrf2 signaling pathway, downregulating the apoptotic pathway gene, egl-1.^[95] Zhou QB et.al. reported that the effect of betulin reduced α -syn accumulation, thereby blocking its toxic effect in cells. Previous research has demonstrated that a ubiquitin-proteasome mechanism repairs cells from damage under stressful conditions.^[96] Ci et al. reported that betulin has the effect on regulating AMPK/AKT/Nrf2 signaling to mitigate inflammation^[97] Zhou et. al. found that betulin can modulate immune response to alleviate liver damage from hepatitis.^[98] Based on these accumulating evidence and results, presumed that DJ-1 had same merits in EBI caused by subarachnoid hemorrhage(SAH) via regulating Akt/Nrf2 signaling pathway, for kindred mechanisms play similar roles in multiple neurological diseases.^[99,100] It is found that the outer bark of birch trees has various biological activities on animal and human health including the following properties like antibacterial,^[111] antiparasitic,^[112] antiviral,^[113] antiinflammatory,^[114] anticancer,^[115] antivenom^[116] liver protective,^[117] kidney protective,^[118] lung protective,^[119] anticonvulsant and a cognitive enhancer in patients with alzheimer's disease, it also improves diet-induced obesity, ameliorates the stability of atherosclerotic plaques, and can be used to treat type II diabetes.^[120]

CONCLUSION

Parkinson's disease (PD) is a chronic neurological condition that largely impairs mobility. It is caused by the degeneration of dopamine-producing neurons in the substantia nigra, resulting in a lack of dopamine, a crucial neurotransmitter involved in motor function. L-dopa does not effectively treat neuronal loss, nonmotor symptoms or Lewy pathology. Patients gradually demand increasing dosages of L-dopa, which are accompanied with significant side effects such as dyskinesia.

Naturally derived phytochemicals and their derivatives have a potential neuroprotective effect due to their multifaceted ability to regulate and modulate chronic inflammation, oxidative stress, and downstream signaling of different pathways, all of which are hallmarks of parkinson's disease.

In this review, we have unlocked the power of different phytochemicals in parkinson's disease. Chrysin which acts on the dopamine transporter (DAT) pathways and increased the levels of dopamine. Daphnetin acts by downregulations of pathways like janus kinase (JAK/STAT), nuclear related factor (Nrf-2), Nf-kB, beclin-2(Bax/Bcl-2). Theaflavin,

increased the expression of dopamine transporter protein (DAT) and vesicular monoamine transporter-2 (VMAT-2). Epigallocatechin-3-gallate inhibited the overexpression of STAT3, NF- κ B. (α -) asarone and (β -) asarone downregulated JNK and p-JNK, PERK/CHOP/Bcl-2/Beclin-1 pathways. Baicalein suppressed the nuclear factor erythroid-derived 2 (NF-E2) related factor (Nrf2), and regulated AMPK/AKT/Nrf2 signaling to mitigate inflammation (NF- κ B), mitogen-activated protein kinases (MAPKs), cAMP-response element binding protein (CREB), janus kinase/signal transducer and activator of transcription and TLR-4. Betulin causes significant effect on AMPK/AKT/Nrf2 signaling. Thus, these phytochemicals will help to act by bridging current insight to future therapeutics innovations.

Future research should focus on increasing clinical acceptance of claims derived from in vitro and in vivo preclinical investigations, as well as conducting additional clinical trial studies of numerous more possible drugs and combinations thereof, to detect and prevent any undesired side effects. The efficacy of phytochemicals in clinical research will determine their pharmacological relevance in humans and nutritional intervention programs will diminish oxidative neuroinflammatory damage and halt the progression of parkinson's disease.

REFERENCE

1. Balakrishnan R, Azam S, Cho DY, Su-Kim I, Choi DK. Natural phytochemicals as novel therapeutic strategies to prevent and treat Parkinson's disease: current knowledge and future perspectives. *Oxidative Medicine and Cellular Longevity*, 2021; 2021(1): 6680935.
2. Duty S. Targeting glutamate receptors to tackle the pathogenesis, clinical symptoms and levodopa-induced dyskinesia associated with Parkinson's disease. *CNS drugs*, 2012 Dec; 26: 1017-32.
3. Zeng XS, Geng WS, Jia JJ. Neurotoxin-induced animal models of Parkinson disease: pathogenic mechanism and assessment. *ASN neuro*, 2018 May; 10: 1759091418777438.
4. Fink AL. The aggregation and fibrillation of α -synuclein. *Accounts of chemical research*, 2006 Sep 19; 39(9): 628-34.
5. Chagraoui A, Boulain M, Juvin L, Anouar Y, Barrière G, De Deurwaerdère P. L-DOPA in parkinson's disease: Looking at the "false" neurotransmitters and their meaning. *International journal of molecular sciences*, 2019 Dec 31; 21(1): 294.
6. Picconi B, Centonze D, Håkansson K, Bernardi G, Greengard P, Fisone G, Cenci MA, Calabresi P. Loss of bidirectional striatal synaptic plasticity in L-DOPA-induced dyskinesia. *Nature neuroscience*, 2003 May; 6(5): 501-6.

7. Prakash D, Sharma G, editors. *Phytochemicals of nutraceutical importance*. CABI, 2014.
8. Moosavi F, Hosseini R, Saso L, Firuzi O. Modulation of neurotrophic signaling pathways by polyphenols. *Drug design, development and therapy*, 2015 Dec 21; 23-42.
9. Mandel S, Youdim MB. Catechin polyphenols: neurodegeneration and neuroprotection in neurodegenerative diseases. *Free Radical Biology and Medicine*, 2004 Aug 1; 37(3): 304-17.
10. Limanaqi F, Biagioni F, Busceti CL, Ryskalin L, Polzella M, Frati A, Fornai F. Phytochemicals bridging autophagy induction and alpha-synuclein degradation in parkinsonism. *International journal of molecular sciences*, 2019 Jul 3; 20(13): 3274.
11. Mani R, Natesan V. Chrysin: Sources, beneficial pharmacological activities, and molecular mechanism of action. *Phytochemistry*, 2018 Jan 1; 145: 187-96.
12. Walle T, Otake Y, Brubaker JA, Walle UK, Halushka PV. Disposition and metabolism of the flavonoid chrysin in normal volunteers. *British journal of clinical pharmacology*, 2001 Feb; 51(2): 143-6.
13. Goes AT, Jesse CR, Antunes MS, Ladd FV, Ladd AA, Luchese C, Paroul N, Boeira SP. Protective role of chrysin on 6-hydroxydopamine-induced neurodegeneration a mouse model of Parkinson's disease: Involvement of neuroinflammation and neurotrophins. *Chemico-biological interactions*, 2018 Jan 5; 279: 111-20.
14. Guo B, Zheng C, Cai W, Cheng J, Wang H, Li H, Sun Y, Cui W, Wang Y, Han Y, Lee SM. Multifunction of chrysin in Parkinson's model: anti-neuronal apoptosis, neuroprotection via activation of MEF2D, and inhibition of monoamine oxidase-B. *Journal of agricultural and food chemistry*, 2016 Jul 6; 64(26): 5324-33.
15. Krishnamoorthy A, Sevanan M, Mani S, Balu M, Balaji S, Ramajayan P. Chrysin restores MPTP induced neuroinflammation, oxidative stress and neurotrophic factors in an acute Parkinson's disease mouse model. *Neuroscience letters*, 2019 Sep 14; 709: 134382.
16. Del Fabbro L, Goes AR, Jesse CR, de Gomes MG, Souza LC, Ladd FV, Ladd AA, Arantes RV, Simionato AR, Oliveira MS, Furian AF. Chrysin protects against behavioral, cognitive and neurochemical alterations in a 6-hydroxydopamine model of Parkinson's disease. *Neuroscience Letters*, 2019 Jul 27; 706: 158-63.
17. Guo B, Zheng C, Cai W, Cheng J, Wang H, Li H, Sun Y, Cui W, Wang Y, Han Y, Lee SM. Multifunction of chrysin in Parkinson's model: anti-neuronal apoptosis, neuroprotection via activation of MEF2D, and inhibition of monoamine oxidase-B. *Journal of agricultural and food chemistry*, 2016 Jul 6; 64(26): 5324-33.

18. Sampaio TB, Pinton S, da Rocha JT, Gai BM, Nogueira CW. Involvement of BDNF/TrkB signaling in the effect of diphenyl diselenide on motor function in a Parkinson's disease rat model. *European journal of pharmacology*, 2017 Jan 15; 795: 28-35.
19. Liu Y, Song X, He J, Zheng X, Wu H. Synthetic derivatives of chrysin and their biological activities. *Medicinal Chemistry Research*, 2014 Feb; 23: 555-63.
20. Fan, X., Xie, M., Zhao, F., Li, J., Fan, C., Zheng, H., et al. Daphnetin triggers ROS-induced cell death and induces cytoprotective autophagy by modulating the AMPK/Akt/mTOR pathway in ovarian cancer. *Phytomedicine*, 2021 mar; 221-230.
21. Tu, L., Li, S., Fu, Y., Yao, R., Zhang, Z., Yang, S., et al. The therapeutic effects of daphnetin in collagen-induced arthritis involve its regulation of Th17 cells. *Int. Immunopharmacol.*, 2012; 13(4): 417-423.
22. Ueno, K., and Saito, N. Daphnetin, isolated from *Daphne odora*. *Acta Crystallogr. Sect. B.*, 1976; 32(3): 946-948.
23. Qi Z, Qi S, Gui L, Shen L, Feng Z. Daphnetin protects oxidative stress-induced neuronal apoptosis via regulation of MAPK signaling and HSP70 expression. *Oncology Letters*, 2016 Sep 1; 12(3): 1959-64.
24. Qi Z, Qi S, Gui L, Shen L, Feng Z. Daphnetin protects oxidative stress-induced neuronal apoptosis via regulation of MAPK signaling and HSP70 expression. *Oncology Letters*, 2016 Sep 1; 12(3): 1959-64.
25. Vinayagam, R., and Xu, B. 7, 8-Dihydroxycoumarin (daphnetin) protects INS-1 pancreatic β -cells against streptozotocin-induced apoptosis. *Phytomedicine*, 2017; 24: 119-126.
26. Wang D, Zhu B, Liu X, Han Q, Ge W, Zhang W, Lu Y, Wu Q, Shi L. Daphnetin ameliorates experimental autoimmune encephalomyelitis through regulating heme oxygenase-1. *Neurochemical research*, 2020 Apr; 45: 872-81.
27. Zhang L, Gu Y, Li H, Cao H, Liu B, Zhang H, Shao F. Daphnetin protects against cisplatin-induced nephrotoxicity by inhibiting inflammatory and oxidative response. *International immunopharmacology*, 2018 Dec 1; 65: 402-7.
28. Xu K, Guo L, Bu H, Wang H. Daphnetin inhibits high glucose-induced extracellular matrix accumulation, oxidative stress and inflammation in human glomerular mesangial cells. *Journal of Pharmacological Sciences*, 2019 Feb 1; 139(2): 91-7.
29. Schuh C, Schieberle P. Characterization of the key aroma compounds in the beverage prepared from Darjeeling black tea: quantitative differences between tea leaves and infusion. *Journal of agricultural and food chemistry*, 2006 Feb 8; 54(3): 916-24.

30. Grelle G, Otto A, Lorenz M, Frank RF, Wanker EE, Bieschke J. Black tea theaflavins inhibit formation of toxic amyloid- β and α -synuclein fibrils. *Biochemistry*, 2011 Dec 13; 50(49): 10624-36.
31. Aizawa T, Yamamoto A, Ueno T. Effect of oral theaflavin administration on body weight, fat, and muscle in healthy subjects: a randomized pilot study. *Bioscience, biotechnology, and biochemistry*, 2017 Feb 1; 81(2): 311-5.
32. Luo Z, Zhao Y, Wang Y, Yang X, Zhao B. Protective effect of theaflavins on neuron against 6-hydroxydopamine-induced apoptosis in SH-SY5Y cells. *Journal of clinical biochemistry and nutrition*, 2012; 50(2): 133-8.
33. Zhang J, Cai S, Li J, Xiong L, Tian L, Liu J, Huang J, Liu Z. Neuroprotective effects of theaflavins against oxidative stress-induced apoptosis in PC12 cells. *Neurochemical research*, 2016 Dec; 41: 3364-72.
34. Grelle G, Otto A, Lorenz M, Frank RF, Wanker EE, Bieschke J. Black tea theaflavins inhibit formation of toxic amyloid- β and α -synuclein fibrils. *Biochemistry*, 2011 Dec 13; 50(49): 10624-36.
35. Anandhan A, Janakiraman U, Manivasagam T. Theaflavin ameliorates behavioral deficits, biochemical indices and monoamine transporters expression against subacute 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP)-induced mouse model of Parkinson's disease. *Neuroscience*, 2012 Aug 30; 218: 257-67.
36. Anandhan A, Tamilselvam K, Radhiga T, Rao S, Essa MM, Manivasagam T. Theaflavin, a black tea polyphenol, protects nigral dopaminergic neurons against chronic MPTP/probenecid induced Parkinson's disease. *Brain Research*, 2012 Jan 18; 1433: 104-13.
37. Szuster-Ciesielska A, Tustanowska-Stachura A, Slotwinska M, Marmurowska-Michalowska H, Kandefer-Szerszeń M. In vitro immunoregulatory effects of antidepressants in healthy volunteers. *Polish journal of pharmacology*, 2003 May 1; 55(3): 353-62.
38. O'Neill EJ, Termini D, Albano A, Tsiani E. Anti-cancer properties of theaflavins. *Molecules*, 2021 Feb 13; 26(4): 987.
39. Anandhan A, Essa MM, Manivasagam T. Therapeutic attenuation of neuroinflammation and apoptosis by black tea theaflavin in chronic MPTP/probenecid model of Parkinson's disease. *Neurotoxicity Research*, 2013 Feb; 23: 166-73.

40. Sirk TW, Friedman M, Brown EF. Molecular binding of black tea theaflavins to biological membranes: relationship to bioactivities. *Journal of agricultural and food chemistry*, 2011 Apr 27; 59(8): 3780-7.
41. Dreger H, Lorenz M, Kehrer A, Baumann G, Stangl K, Stangl V. Characteristics of catechin-and theaflavin-mediated cardioprotection. *Experimental Biology and Medicine*, 2008 Apr; 233(4): 427-33.
42. Zu M, Yang F, Zhou W, Liu A, Du G, Zheng L. In vitro anti-influenza virus and anti-inflammatory activities of theaflavin derivatives. *Antiviral Research*, 2012 Jun 1; 94(3): 217-24.
43. Singh R, Akhtar N, Haqqi TM. Green tea polyphenol epigallocatechin-3-gallate: Inflammation and arthritis. *Life sciences*, 2010 Jun 19; 86(25-26): 907-18.
44. Sharma VK, Bhattacharya A, Kumar A, Sharma HK. Health benefits of tea consumption. *Tropical Journal of Pharmaceutical Research*, 2007; 6(3): 785-92.
45. Lambert JD, Elias RJ. The antioxidant and pro-oxidant activities of green tea polyphenols: a role in cancer prevention. *Archives of biochemistry and biophysics*, 2010 Sep 1; 501(1): 65-72.
46. Lambert JD, Elias RJ. The antioxidant and pro-oxidant activities of green tea polyphenols: a role in cancer prevention. *Archives of biochemistry and biophysics*, 2010 Sep 1; 501(1): 65-72.
47. Mandel SA, Avramovich-Tirosh Y, Reznichenko L, Zheng H, Weinreb O, Amit T, Youdim MB. Multifunctional activities of green tea catechins in neuroprotection. *Neurosignals*, 2005 Jun 10; 14(1-2): 46-60.
48. Unno K, Takabayashi F, Kishido T, Oku N. Suppressive effect of green tea catechins on morphologic and functional regression of the brain in aged mice with accelerated senescence (SAMP10). *Experimental Gerontology*, 2004 Jul 1; 39(7): 1027-34.
49. Unno K, Takabayashi F, Yoshida H, Choba D, Fukutomi R, Kikunaga N, Kishido T, Oku N, Hoshino M. Daily consumption of green tea catechin delays memory regression in aged mice. *Biogerontology*, 2007 Apr; 8: 89-95.
50. Schaffer S, Halliwell B. Do polyphenols enter the brain and does it matter? Some theoretical and practical considerations. *Genes & nutrition*, 2012 Apr; 7: 99-109.
51. Choi JY, Park CS, Kim DJ, Cho MH, Jin BK, Pie JE, Chung WG. Prevention of nitric oxide-mediated 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine-induced Parkinson's disease in mice by tea phenolic epigallocatechin 3-gallate. *Neurotoxicology*, 2002 Sep 1; 23(3): 367-74.

52. Ritchie K, Lovestone S. The dementias. *The Lancet*, 2002 Nov 30; 360(9347): 1759-66.
53. Hayat K, Iqbal H, Malik U, Bilal U, Mushtaq S. Tea and its consumption: benefits and risks. *Critical reviews in food science and nutrition*, 2015 Jun 7; 55(7): 939-54.
54. Mattson MP. Apoptosis in neurodegenerative disorders. *Nature reviews Molecular cell biology*, 2000 Nov 1; 1(2): 120-30.
55. Sadrzadeh SH, Saffari Y. Iron and brain disorders. *Pathology Patterns Reviews*, 2004 May 1; 121(suppl_1): S64-70.
56. Tseng HC, Wang MH, Chang KC, Soung HS, Fang CH, Lin YW, Li KY, Yang CC, Tsai CC. Protective effect of (-) epigallocatechin-3-gallate on rotenone-induced parkinsonism-like symptoms in rats. *Neurotoxicity Research*, 2020 Mar; 37: 669-82.
57. Levites Y, Weinreb O, Maor G, Youdim MB, Mandel S. Green tea polyphenol (-)-epigallocatechin-3-gallate prevents N-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine-induced dopaminergic neurodegeneration. *Journal of neurochemistry*, 2001 Sep 1; 78(5): 1073-82.
58. Wang L, Xu S, Xu X, Chan P. (-)-Epigallocatechin-3-gallate protects SH-SY5Y cells against 6-OHDA-induced cell death through STAT3 activation. *Journal of Alzheimer's Disease*, 2009 Jan 1; 17(2): 295-304.
59. Wang L, Xu S, Xu X, Chan P. (-)-Epigallocatechin-3-gallate protects SH-SY5Y cells against 6-OHDA-induced cell death through STAT3 activation. *Journal of Alzheimer's Disease*, 2009 Jan 1; 17(2): 295-304.
60. Zhou T, Zhu M, Liang Z. (-)-Epigallocatechin-3-gallate modulates peripheral immunity in the MPTP-induced mouse model of Parkinson's disease. *Molecular medicine reports*, 2018 Apr 1; 17(4): 4883-8.
61. Zhou T, Zhu M, Liang Z. (-)-Epigallocatechin-3-gallate modulates peripheral immunity in the MPTP-induced mouse model of Parkinson's disease. *Molecular medicine reports*, 2018 Apr 1; 17(4): 4883-8.
62. Hou RR, Chen JZ, Chen H, Kang XG, Li MG, Wang BR. Neuroprotective effects of (-)-epigallocatechin-3-gallate (EGCG) on paraquat-induced apoptosis in PC12 cells. *Cell biology international.*, 2008 Jan; 32(1): 22-30.
63. Kang KS, Wen Y, Yamabe N, Fukui M, Bishop SC, Zhu BT. Dual beneficial effects of (-)-epigallocatechin-3-gallate on levodopa methylation and hippocampal neurodegeneration: in vitro and in vivo studies. *PloS one*, 2010 Aug 5; 5(8): e11951.

64. Ehrnhoefer DE, Bieschke J, Boeddrich A, Herbst M, Masino L, Lurz R, Engemann S, Pastore A, Wanker EE. EGCG redirects amyloidogenic polypeptides into unstructured, off-pathway oligomers. *Nature structural & molecular biology*, 2008 Jun; 15(6): 558-66.
65. Xu Y, Zhang Y, Quan Z, Wong W, Guo J, Zhang R, Yang Q, Dai R, McGeer PL, Qing H. Epigallocatechin gallate (EGCG) inhibits alpha-synuclein aggregation: A potential agent for Parkinson's disease. *Neurochemical research*, 2016 Oct; 41: 2788-96.
66. Cheng B, Gong H, Xiao H, Petersen RB, Zheng L, Huang K. Inhibiting toxic aggregation of amyloidogenic proteins: a therapeutic strategy for protein misfolding diseases. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 2013 Oct 1; 1830(10): 4860-71.
67. Lu J, Fu T, Qian Y, Zhang Q, Zhu H, Pan L, Guo L, Zhang M. Distribution of α -asarone in brain following three different routes of administration in rats. *European Journal of Pharmaceutical Sciences*, 2014 Oct 15; 63: 63-70.
68. Fang YQ, Shi C, Liu L, Fang RM. Analysis of transformation and excretion of β -asarone in rabbits with GC-MS. *European journal of drug metabolism and pharmacokinetics*, 2012 Sep; 37: 187-90.
69. Liu L, Fang YQ. Analysis of the distribution of β -asarone in rat hippocampus, brainstem, cortex and cerebellum with gas chromatography-mass spectrometry (GC-MS). *J Med Plants Res.*, 2011 May 4; 5(9): 1728-34.
70. Hei X, Xie M, Xu J, Li J, Liu T. β -asarone exerts antioxidative effects on H₂O₂-stimulated PC12 cells by activating Nrf2/HO-1 pathway. *Neurochemical Research*, 2020 Aug; 45: 1953-61.
71. Kim BW, Koppula S, Kumar H, Park JY, Kim IW, More SV, Kim IS, Han SD, Kim SK, Yoon SH, Choi DK. α -Asarone attenuates microglia-mediated neuroinflammation by inhibiting NF kappa B activation and mitigates MPTP-induced behavioral deficits in a mouse model of Parkinson's disease. *Neuropharmacology*, 2015 Oct 1; 97: 46-57.
72. Zhang S, Gui XH, Huang LP, Deng MZ, Fang RM, Ke XH, He YP, Li L, Fang YQ. Neuroprotective effects of β -asarone against 6-hydroxy dopamine-induced parkinsonism via JNK/Bcl-2/Beclin-1 pathway. *Molecular neurobiology*, 2016 Jan; 53: 83-94.
73. Kuma A, Hatano M, Matsui M, Yamamoto A, Nakaya H, Yoshimori T, Ohsumi Y, Tokuhisa T, Mizushima N. The role of autophagy during the early neonatal starvation period. *Nature*, 2004 Dec 23; 432(7020): 1032-6.
74. Pan T, Kondo S, Le W, Jankovic J. The role of autophagy-lysosome pathway in neurodegeneration associated with Parkinson's disease. *Brain.*, 2008 Aug 1; 131(8): 1969-78.

75. Jheng JR, Ho JY, Horng JT. ER stress, autophagy, and RNA viruses. *Frontiers in microbiology*, 2014 Aug 5; 5: 388.
76. Ning B, Deng M, Zhang Q, Wang N, Fang Y. β -Asarone inhibits IRE1/XBP1 endoplasmic reticulum stress pathway in 6-OHDA-induced parkinsonian rats. *Neurochemical research*, 2016 Aug; 41: 2097-101.
77. Wei G, Chen YB, Chen DF, Lai XP, Liu DH, Deng RD, Zhou JH, Zhang SX, Li YW, Lii H, Liu LF. β -Asarone inhibits neuronal apoptosis via the CaMKII/CREB/Bcl-2 signaling pathway in an in vitro model and A β PP/PS1 mice. *Journal of Alzheimer's Disease*, 2013 Jan 1; 33(3): 863-80.
78. Huang L, Deng M, He Y, Lu S, Liu S, Fang Y. β -asarone increases MEF2D and TH levels and reduces α -synuclein level in 6-OHDA-induced rats via regulating the HSP70/MAPK/MEF2D/Beclin-1 pathway: Chaperone-mediated autophagy activation, macroautophagy inhibition and HSP70 up-expression. *Behavioural Brain Research*, 2016 Oct 15; 313: 370-9.
79. Venkatesan R, Karuppiiah PS, Arumugam G, Balamuthu K. β -Asarone exhibits antifungal activity by inhibiting ergosterol biosynthesis in *Aspergillus niger* ATCC 16888. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 2019 Mar 4; 89: 173-84.
80. McGaw LJ, Jäger AK, Van Staden J, Eloff JN. Isolation of β -asarone, an antibacterial and anthelmintic compound, from *Acorus calamus* in South Africa. *South African Journal of Botany*, 2002 Feb 1; 68(1): 31-5.
81. Xiao B, Huang X, Wang Q, Wu Y. Beta-Asarone Alleviates Myocardial Ischemia–Reperfusion Injury by Inhibiting Inflammatory Response and NLRP3 Inflammasome Mediated Pyroptosis. *Biological and Pharmaceutical Bulletin*, 2020 Jul 1; 43(7): 1046-51.
82. Shenvi S, Diwakar L, Reddy GC. Nitro derivatives of naturally occurring β -asarone and their anticancer activity. *International journal of medicinal chemistry*, 2014; 2014(1): 835485.
83. Hung KC, Huang HJ, Wang YT, Lin AM. Baicalein attenuates α -synuclein aggregation, inflammasome activation and autophagy in the MPP⁺-treated nigrostriatal dopaminergic system in vivo. *Journal of ethnopharmacology*, 2016 Dec 24; 194: 522-9.
84. Im HI, Joo WS, Nam E, Lee ES, Hwang YJ, Kim YS. Baicalein prevents 6-hydroxydopamine-induced dopaminergic dysfunction and lipid peroxidation in mice. *Journal of pharmacological sciences*, 2005; 98(2): 185-9.

85. Hu Q, Uversky VN, Huang M, Kang H, Xu F, Liu X, Lian L, Liang Q, Jiang H, Liu A, Zhang C. Baicalein inhibits α -synuclein oligomer formation and prevents progression of α -synuclein accumulation in a rotenone mouse model of Parkinson's disease. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 2016 Oct 1; 1862(10): 1883-90.
86. Mu X, He G, Cheng Y, Li X, Xu B, Du G. Baicalein exerts neuroprotective effects in 6-hydroxydopamine-induced experimental parkinsonism in vivo and in vitro. *Pharmacology Biochemistry and Behavior*, 2009 Jun 1; 92(4): 642-8.
87. Si L, Lai Y. Pharmacological mechanisms by which baicalin ameliorates cardiovascular disease. *Frontiers in Pharmacology*, 2024 Aug 9; 15: 1415971.
88. Zhao Q, Chen XY, Martin C. *Scutellaria baicalensis*, the golden herb from the garden of Chinese medicinal plants. *Science bulletin*, 2016 Sep; 61: 1391-8.
89. Lee W, Ku SK, Bae JS. Anti-inflammatory effects of Baicalin, Baicalein, and Wogonin in vitro and in vivo. *Inflammation*, 2015 Feb; 38: 110-25.
90. Shieh DE, Liu LT, Lin CC. Antioxidant and free radical scavenging effects of baicalein, baicalin and wogonin. *Anticancer research*, 2000 Sep 1; 20(5A): 2861-5.
91. Peng-Fei L, Fu-Gen H, Bin-Bin D, Tian-Sheng D, Xiang-Lin H, Ming-Qin Z. Purification and antioxidant activities of baicalin isolated from the root of huangqin (*Scutellaria baicalensis* gcorisi). *Journal of food science and technology*, 2013 Jun; 50: 615-9.
92. Song Z, He C, Yu W, Yang M, Li Z, Li P, Zhu X, Xiao C, Cheng S. Baicalin Attenuated A β 1-42-Induced Apoptosis in SH-SY5Y Cells by Inhibiting the Ras-ERK Signaling Pathway. *BioMed Research International*, 2022; 2022(1): 9491755.
93. Ju XN, Mu WN, Liu YT, Wang MH, Kong F, Sun C, Zhou QB. Baicalin protects against thrombin induced cell injury in SH-SY5Y cells. *International Journal of Clinical and Experimental Pathology*, 2015; 8(11): 14021.
94. Zhou QB, Ju XN, Wang XY, Wang MH, Kong F, Sun C, Bi JZ. Pretreatment with baicalin attenuates hypoxia and glucose deprivation-induced injury in SH-SY5Y cells. *Chinese journal of integrative medicine*, 2016 Mar; 22: 201-6.
95. Xin L, Gao J, Lin H, Qu Y, Shang C, Wang Y, Lu Y, Cui X. Regulatory mechanisms of baicalin in cardiovascular diseases: a review. *Frontiers in pharmacology*, 2020 Nov 2; 11: 583200.
96. Zhang Y, Liao P, Zhu ME, Li W, Hu D, Guan S, Chen L. Baicalin attenuates cardiac dysfunction and myocardial remodeling in a chronic pressure-overload mice model. *Cellular Physiology and Biochemistry*, 2017 Feb 16; 41(3): 849-64.

97. Zhao T, Tang H, Xie L, Zheng Y, Ma Z, Sun Q, Li X. Betulin: a review of its traditional uses, botany, phytochemistry, pharmacology and toxicology. *Journal of Pharmacy and Pharmacology*, 2019 Sep; 71(9): 1353-69.
98. Zhu W, Jin Z, Yu J, Liang J, Yang Q, Li F, Shi X, Zhu X, Zhang X. Betulin ameliorates experimental inflammatory bowel disease through polarization of macrophages to an M2 phenotype. *International Immunopharmacology*, 2016 Jun 1; 35: 119-26.
99. Zhang P, Xu D, Wang S, Fu H, Wang K, Zou Y, Sun A, Ge J. Inhibition of aldehyde dehydrogenase 2 activity enhances antimycin-induced rat cardiomyocytes apoptosis through activation of MAPK signaling pathway. *Biomedicine & Pharmacotherapy*, 2011 Dec 1; 65(8): 590-3.
100. Amiri S, Dastghaib S, Ahmadi M, Mehrbod P, Khadem F, Behrouj H, Aghanoori MR, Machaj F, Ghamsari M, Rosik J, Hudecki A. Betulin and its derivatives as novel compounds with different pharmacological effects. *Biotechnology advances*, 2020 Jan 1; 38: 107409.
101. Alakurtti S, Mäkelä T, Koskimies S, Yli-Kauhaluoma J. Pharmacological properties of the ubiquitous natural product betulin. *European journal of pharmaceutical sciences*, 2006 Sep 1; 29(1): 1-3.
102. Ekman R, Eckerman C. Aliphatic carboxylic acids from suberin in birch outer bark by hydrolysis, methanolysis, and alkali fusion. *Paperi ja puu.*, 1985; 67(4): 255-73.
103. Opattova A, Cente M, Novak M, Filipcik P. The ubiquitin proteasome system as a potential therapeutic target for treatment of neurodegenerative diseases. *Gen Physiol Biophys*, 2015 Oct 1; 34(4): 337-52.
104. Tsai CW, Tsai RT, Liu SP, Chen CS, Tsai MC, Chien SH, Hung HS, Lin SZ, Shyu WC, Fu RH. Neuroprotective effects of betulin in pharmacological and transgenic *Caenorhabditis elegans* models of Parkinson's disease. *Cell transplantation*, 2017 Dec; 26(12): 1903-18.
105. Lu X, Yang S, Lu Q, Zhang Y, Cha Z, Huang W, Li T. Betulin ameliorates neuronal apoptosis and oxidative injury via DJ-1/Akt/Nrf2 signaling pathway after subarachnoid hemorrhage. *CNS Neuroscience & Therapeutics*, 2024 Sep; 30(9): e70019.
106. Lu X, Yang S.
107. Lu Q, Zhang Y, Cha Z, Huang W, Li T. Betulin ameliorates neuronal apoptosis and oxidative injury via DJ-1/Akt/Nrf2 signaling pathway after subarachnoid hemorrhage. *CNS Neuroscience & Therapeutics*, 2024 Sep; 30(9): e70019.

108. Abrishamdar M, Farbood Y, Sarkaki A, Rashno M, Badavi M. Evaluation of Betulinic Acid Effects on Pain, Anxiety, Catalepsy, and Oxidative Stress in Animal Model of Parkinson's Disease.
109. Farzan M, Farzan M, Shahrani M, Navabi SP, Vardanjani HR, Amini-Khoei H, Shabani S. Neuroprotective properties of Betulin, Betulinic acid, and Ursolic acid as triterpenoids derivatives: A comprehensive review of mechanistic studies. *Nutritional neuroscience*, 2024 Mar 3; 27(3): 223-40.
110. Wu Q, Li H, Qiu J, Feng H. Betulin protects mice from bacterial pneumonia and acute lung injury. *Microbial pathogenesis*, 2014 Oct 1; 75: 21-8.
111. Haque S, Nawrot DA, Alakurtti S, Ghemtio L, Yli-Kauhaluoma J, Tammela P. Screening and characterisation of antimicrobial properties of semisynthetic betulin derivatives. *PLoS One.*, 2014 Jul 17; 9(7): e102696.
112. Sousa MC, Varandas R, Santos RC, Santos-Rosa M, Alves V, Salvador JA. Antileishmanial activity of semisynthetic lupane triterpenoids betulin and betulinic acid derivatives: synergistic effects with miltefosine. *PloS one*, 2014 Mar 18; 9(3): e89939.
113. Alcazar W, López AS, Alakurtti S, Tuononen ML, Yli-Kauhaluoma J, Ponte-Sucré A. Betulin derivatives impair *Leishmania braziliensis* viability and host–parasite interaction. *Bioorganic & Medicinal Chemistry*, 2014 Nov 1; 22(21): 6220-6.
114. Guo MY, Li WY, Zhang Z, Qiu C, Li C, Deng G. Betulin suppresses *S. aureus*-induced mammary gland inflammatory injury by regulating PPAR- γ in mice. *International Immunopharmacology*, 2015 Dec 1; 29(2): 824-31.
115. Zhang SY, Zhao QF, Fang NN, Yu JG. Betulin inhibits pro-inflammatory cytokines expression through activation STAT3 signaling pathway in human cardiac cells. *Eur Rev Med Pharmacol Sci.*, 2015 Jan 1; 19(3): 455-60.
116. Reutrakul V, Anantachoke N, Pohmakotr M, Jaipetch T, Yoosook C, Kasisit J, Napaswa C, Panthong A, Santisuk T, Prabpai S, Kongsaree P. Anti-HIV-1 and Anti-Inflammatory Lupanes from the Leaves, Twigs, and Resin of *Garcinia hanburyi*. *Planta medica*, 2010 Mar; 76(04): 368-71.
117. Yang SJ, Liu MC, Xiang HM, Zhao Q, Xue W, Yang S. Synthesis and in vitro antitumor evaluation of betulin acid ester derivatives as novel apoptosis inducers. *European Journal of Medicinal Chemistry*, 2015 Sep 18; 102: 249-55.
118. Yim NH, Jung YP, Kim A, Kim T, Ma JY. Induction of apoptotic cell death by betulin in multidrug-resistant human renal carcinoma cells. *Oncology reports*, 2015 Aug 1; 34(2): 1058-64.

119. Härmä V, Haavikko R, Virtanen J, Ahonen I, Schukov HP, Alakurtti S, Purev E, Rischer H, Yli-Kauhaluoma J, Moreira VM, Nees M. Optimization of invasion-specific effects of betulin derivatives on prostate cancer cells through lead development. PLoS One, 2015 May 12; 10(5): e0126111.
120. Seervi M, Lotankar S, Barbar S, Sathaye S. Assessment of cytochrome P450 inhibition and induction potential of lupeol and betulin in rat liver microsomes. Drug Metabolism and Personalized Therapy, 2016 Jun 1; 31(2): 115-22.