

EFFICIENCY OF THE DECOMPOSITION PROCESS OF *AGARICUS BISPORUS* IN THE MYCOREMEDIATION OF REFINERY WASTEWATER: ROMI STREAM CASE STUDY

Ugya A.Y.*¹ and Imam T.S.¹

¹Biological Sciences Department, Bayero University Kano, Kano, Nigeria.

Article Received on
01 Dec. 2016,

Revised on 21 Dec. 2016,
Accepted on 11 Jan. 2017

DOI: 10.20959/wjpr20172-7549

***Corresponding Author**

Ugya A.Y.

Biological Sciences
Department, Bayero
University Kano, Kano,
Nigeria.

ABSTRACT

This study involves the use of the decomposition of *Agaricus bisporus* in the mycoremediation of refinery waste water. Kaduna Refining and Petrochemical Company release its waste water into Romi stream which is affecting the quality of the stream water and its Biota. Several technologies have been employed to treat this water but all to no avail. Mycology which is an emerging technique in the treatment of waste water was employed in this study owing to the fact that the decomposition of this organism is associated with bacteria's that have the potentials of degrading pollutants. This study recorded increase in pH and Dissolved Oxygen (DO) but recorded high reduction efficiency

for Sulphate, Phosphate, Nitrate, Alkalinity, Electrical Conductivity (EC), Biological Oxygen Demand (BOD), Chemical Oxygen Demand (COD) and heavy metals (Manganese (Mn), Silver (Ag), Lead (Pb), Mercury (Hg) and Zinc (Zn). These studies thereby show that the decomposition process of *Agaricus bisporus* is effective in the remediation of wastewater from Kaduna Refining and Petrochemical Company.

KEYWORDS: Wastewater, Remediation, Fungi, Refinery wastewater, Flora and Fauna.

INTRODUCTION

The world's ever increasing population and her progressive adoption of an industrial- based lifestyle has inevitably led to an increased anthropogenic impact on the biosphere (Asamudo *et al.*, 2005; Aliyu *et al.*, 2016). Since the beginning of the industrial revolution, water pollution by toxic metals has accelerated dramatically. According to Nriagu (1996) about 90% of the anthropogenic emissions of heavy metals have occurred since 1900 AD; it is now well recognized that human activities lead to a substantial accumulation of heavy metals in

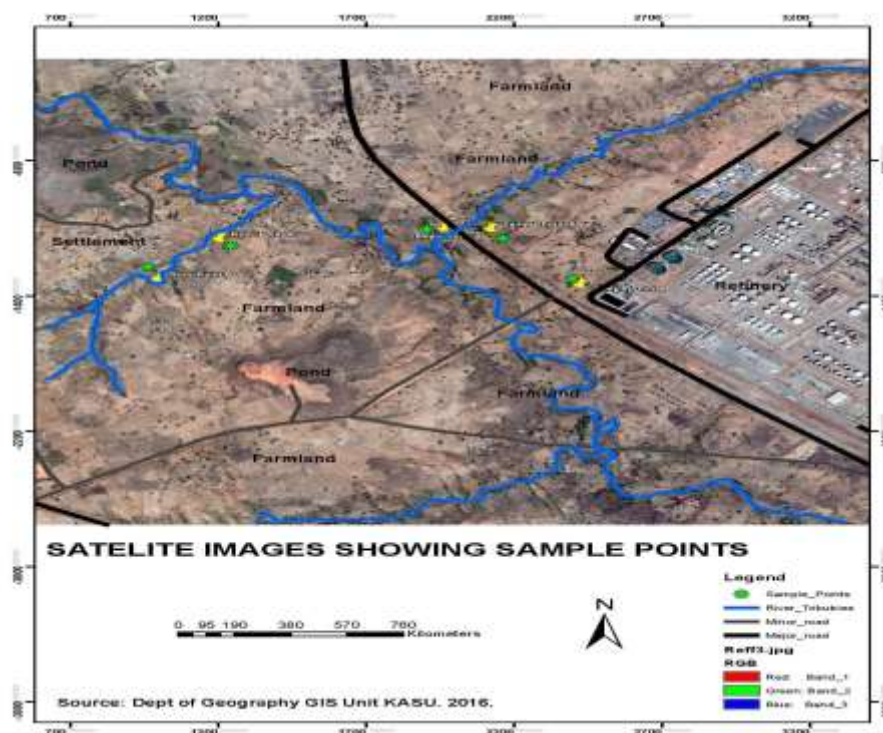
water on a global scale. Man's exposure to heavy metals comes from industrial activities like mining, smelting, refining and manufacturing processes (Nriagu, 1996). A number of chemicals, heavy metals and other industries in the coastal areas have resulted in significant discharge of industrial effluents into the coastal water bodies. These toxic substances are released into the environment and contribute to a variety of toxic effects on living organisms in food chain (Dembitsky, 2003) by bioaccumulation and biomagnification. Heavy metals, such as cadmium, copper, lead; chromium, zinc and nickel are important environmental pollutants, particularly in areas with high anthropogenic pressure (United States Environmental Protection Agency, 1997). Water bodies has been traditionally the site for disposal for most of the heavy metal wastes which needs to be treated (Bio-Wise, 2003; Aboulroos *et al.*, 2006). In refining of refinery products opportunities exist for the release of other pollutants such as oil and grease, phenol, sulphate, suspended solids, dissolved solids, nitrates, e.t.c (Asamudo *et al.*, 2005; Nayyef and Amal, 2012; Ji *et al.*, 2007; Patel and Kanungo, 2010) in to the ecosystem. These pollutants are produce in an effort to improve human standard of living but ironically their unplanned intrusion into the environment can reverse the same standard of living by impacting negatively on the environment (Asamudo *et al.*, 2005; Subhashini *et al.*, 2013; Xiaomei *et al.*, 2004). Refinery effluents can seep into aquifers and pollutes the underground water or where it is discharge without proper treatment into water bodies, the pollutants cannot be confined within specific boundries (Nayyef and Amal, 2012; Asamudo *et al.*, 2005). They can therefore affect aquatic lifes in enormous ways (Ugya and Ahmad, 2016).

Many technologies have been developed to treat this waste water and restore environmental quality; However their costs are high and most of them are difficult to use under field conditions, hence in such a condition there is an urgent need to study natural, simple, and cost-effective techniques for control of pollution from industrial effluents (Ji *et al.*, 2007). Viewing this fact mycoremediation was assumed to be very useful, as it is an innovative, eco-friendly and efficient technology in which natural properties of fungi are used in engineered system to remediate hazardous wastes water (Ugya *et al.*, 2016). The present study thereby shows the efficiency of the decomposition process of *Agaricus bisporus* in the remediation of refinery waste water from Kaduna Refinery and Petrochemical Company, since the waste water has become a menance to the flora and fauna of Romi.

MATERIAL AND METHOD

The Study Area

Romi stream is one of the tributary of River Kaduna; it is located in the southern part of Kaduna metropolis between latitude 10° to 11° north and longitude 7° to 8°East, River Romi follow a course of about 16.4 km and also receives waste water from Kaduna Refining and Petrochemical Company.



Collection of Samples

Sampling Points

Water samples were collected in five different points (point A, point B, point C, point D and point E); The point E samples were collected at about 14 km away from the point of discharge of the refinery effluents into the Romi stream, while the point A samples were point of discharge of the refinery effluents into the stream. The point B samples were collected at about 4 km away from the point of discharge of the refinery effluents into the stream. The point C and D samples were collected at about 8km and 10km respectively away from the point of discharge of the refinery effluents into the stream. Water samples were collected using Grab method according to APHA (1998); Samples were collected into clean one litre plastic bottles and stored in an ice box of 4°C and was taken to the KEPA and KRPC laboratory within twenty-four hours for analysis. Water samples was collected by lowering pre cleaned plastic bottles into the bottom of the water body, 30 cm deep and allowed to over flow before withdrawing. Water samples were collected from March to October, 2016.

Heavy Metal Determination in water before Treatment

Heavy metals in water samples were determined by Atomic Absorption Spectroscopy method (AAS). According to the procedure, 10ml of HNO₃ (Nitric acid) was diluted with 500ml of distilled water. The Atomic Absorption spectrophotometer had already been calculated according to wavelength of different heavy metal parameter. An aliquot of the stock solution was used as blank by aspirating for 5minutes to enable flushing of burner system and to auto-zero the instrument before measurement.

Experimental Set-up

Experiments were set up to investigate mycoremediatory efficiency by the decomposition of *Agaricus bisporus* in water collected from various points of Romi stream.

The experiments were conducted for 21 days and repeated for the period of 8 month (March-October, 2016).

Experimental Setup

To access the mycoremediation capacity of *Agaricus bisporus* in the removal of heavy metals from refinery waste water, an offsite culture experiment was conducted in General Biology laboratory of Kaduna State University (KASU).

The experiment was grouped into five Set, each set consist of troughs each containing five liters of waste water from each station , the experiments in each group was divided into three sets.

Set 1: *Agaricus bisporus* was inoculated in waste water of different concentration 100%
Water from point A

Set 2: *Agaricus bisporus* was inoculated in waste water of different concentration 100%
Water from point B

Set 3: *Agaricus bisporus* was inoculated in waste water of different concentration 100%
Water from point C

Set 4: *Agaricus bisporus* was inoculated in waste water of different concentration 100%
Water from point D

Set 5: *Agaricus bisporus* was inoculated in waste water of different concentration 100%
Water from point E.

Control: 100% water without *Agaricus bisporus* (control) (Ugya *et al.*, 2015a;).

After 21 days the water samples were then filtered and heavy metal and physicochemical parameters analysis was performed on the water sample using the techniques employed by Ugya and Imam (2015) and Ugya *et al.*, (2015b).

Reduction Percentage

The pollutant removal efficiency of the decomposition process of *Agaricus bisporus* was determined as reduction percentage. The initial and final concentrations of the physiochemical parameters of water were use in the following formula:

$$\text{Reduction Percentage} = \frac{B-A}{B} \times \frac{100}{1}$$

Where A= initial concentration

B= final concentration (APHA 2005; Ugya, 2015)

RESULT AND DISCUSSION

Result obtain in this studies shows that EC, BOD, COD where significantly reduced Fig (4) while the pH was increased from 6.78 – 7.32 across the points. The increase in pH shows that the decomposition of mushroom has aided the conversion of acidic water to neutral water. Similar results were reported using different organism as performed in work by researchers

such as Dipu *et al.*, (2011) Ji *et al.*, (2014), Wang *et al.*, (2015), Ugya *et al.*, (2015c), Ugya *et al.*, (2015d), Xu *et al.*, (2015). The high EC, BOD and COD reduction could be attributed to the degradation of pollutants by the decomposition process of *Agaricus bisporus* which result in the reduction of dissolved CO₂ (Ajayan *et al.*, 2015; Fernandez *et al.*, 2016).

The high phosphorus, sulphate, nitrate and alkaline Fig (5) reduction of the waste water could be attributed to the use of this nutrient by the microorganisms responsible for the decomposition of the *Agaricus bispous*. Ajayan *et al* (2015) phosphate is used in microorganism cells mainly for production of phospholipids, adenosine triphosphates (ATP) and nucleic acid. Hence these nutrients must have been used by the microorganisms responsible for the decomposition of mushroom for growth. Similar results have be reported by Pavasant *et al.*, (2007), Khalaf, (2008), Ertugrul *et al* (2008), Chu *et al* (2009), Cheriaa *et al* (2009), Zhou *et al.*, (2014), Fernandez *et al.*, (2016), Wang *et al.*, (2016) using order organisms.

The high heavy metal (Mn, Ag, Hg, Pb and Zn) Fig (3) removal could be associated with the degradatory role associated with microorganisms responsible for the process of decomposition of Mushroom (Bhaita 2011). Microorganism are useful in the bioremediation of metal contaminated sites due to their ability to tolerate those metals (Malik 2004) and their small cell size; microorganism exhibit a large surface area-to-volume ratio, which is promptly available for contact with the surrounding environment; and their functionally rich cell wall groups can easily interact with cations in solution (Roy *et al.* 1993). They can also secrete enzymes that will degrade heavy metals effectively (Ueshima *et al.*, 2008; Wu *et al.*, 2011; Prandini *et al.*, 2016; Ugya *et al.*, 2016a; Ugya and Imam., 2016).

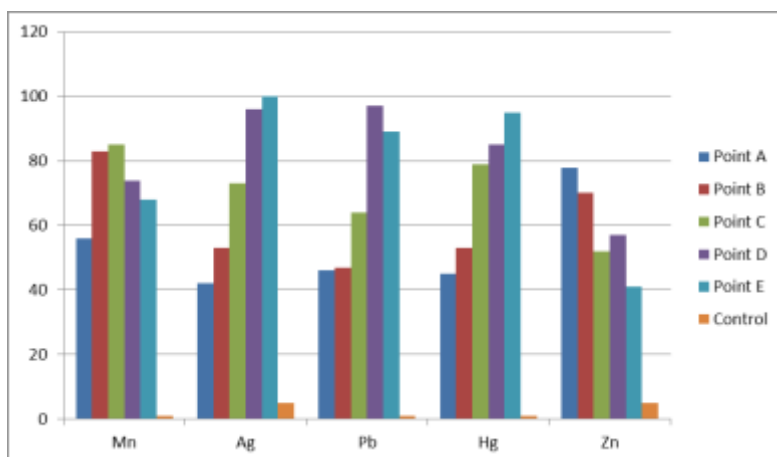


Figure 3: Mean % Reduction Percentage of heavy metals

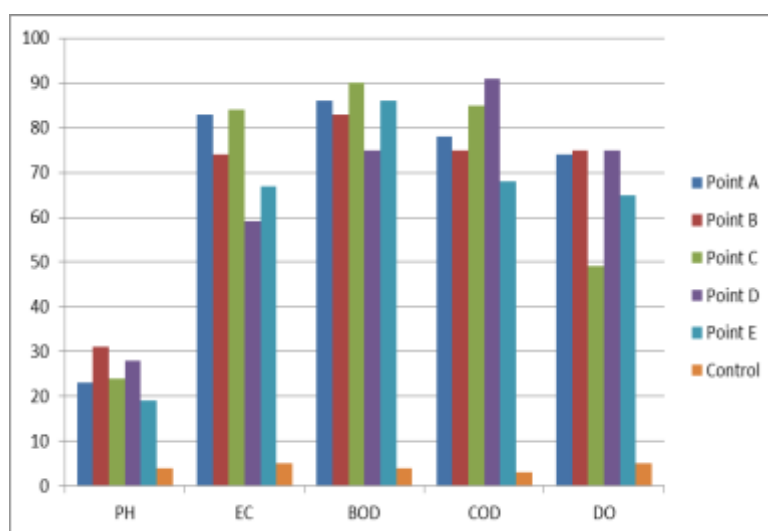


Figure 4: Mean Physico-chemical Parameter Removal Efficiency

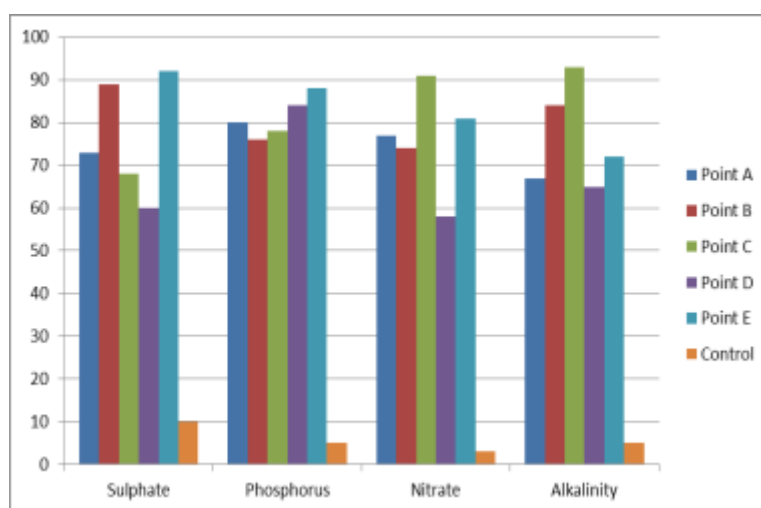


Figure 5: Mean Nutrient Removal efficiency

CONCLUSION

Water quality study of Romi Stream has brought to the fore some important concerns that were muted by research works Like Suzy *et al.*, (2016), which indicated the presence of several heavy metals in high concentration to cause contamination to biotic species of flora and fauna that, abound in the stream. Other parameters monitored such as the oxygen characteristics of the water in terms of COD, BOD and DO, Sulphate, Nitrate, Phosphorus are all indicating toxicity above the threshold that can be purified by the stream. These studies shows that efficiency of the decomposition process of *Agaricus bisporus* in the treatment of the Kaduna Refinery waste water there by reducing the toxicity on the flora and fauna since it is able to remove and degrade pollutants present in the stream to a significant level in all point.

REFERENCES

1. Aboulroos, S.A, Helal, M.I.D and Kamel, M.M. (2006). Remediation Of Pb And Cd Polluted Soils Using In Situ Immobilization And Phytoextraction Techniques. *Soil Sediment Contam.* 15: 199-215.
2. Aliyu, Y1.,Toma, I.M. and Ugya, A.Y. (2016) Effects of Very Low Frequency Electromagnetic Method (VLFEM) and Physicochemical Change of Zango Abattoir, *Bayero Journal of Pure and Applied Sciences*, 9(1): 32 – 38.
3. Asamudo, N.U., A.S. Daba and Ezeronye, O.U. (2005). Bioremediation of Textile Effluent Using Phanerochaete Chrysosporium., *African Journal of Biotechnology*. 4(13): 1548- 1553.
4. American Public Health Association. 2005. Standard methods for the examination of water and wastewater. Washington (DC): APHA.
5. Ajayana, K.V., Selvarajua, M., Unnikannan, P. and Palliyath S. (2015) Phycoremediation of Tannery Wastewater Using Microalgae Scenedesmus Species. *International Journal of Phytoremediation*, 17: 10, 907-916.
6. Bhaita, S.C. (2011). “*Environmental Pollution and Control in Chemical Process Industries*”. 2nd Edition, Kanna Publishers India pp 1273.
7. BIO-WISE (2003). Contaminated Land Remediation: A Review Of Biological Technology, London. DTI.
8. Cheriaa, J., Bettaieb, F., Denden, I., Bakhrouf, A., 2009. Characterization of new algae isolated from textile wastewater plant. *J. Food Agric. Environ.* 7(3–4): 700–704.
9. Chu, W.L., See, Y.C., Phang, S.M., 2009. Use of immobilised *Chlorella vulgaris* for the

- removal of colour from textile dyes. *J. Appl. Phycol.* 21(6): 641–648.
10. Dembitsky, V. (2003). Natural Occurrence of Arsenic Compounds in Plants, Lichens, Fungi, Algal Species and Microorganisms. *Plant Sci.* 165: 1177-1192.
 11. Ertugrul, S., Bakir, M., Donmez, G., 2008. Treatment of dye-rich wastewater by an immobilized thermophilic cyanobacterial strain: *Phormidium* sp. *Ecol. Eng.* 32(3): 244–248.
 12. Fernández, I., Berenguel, M., Guzmán, J.L., Acien, F.G., de Andrade, G.A., Pagano, D.J., 2016. Hierarchical control for microalgae biomass production in photobiorreactors. *Control Eng. Pract.* 54: 246–255.
 13. Franchino, M., Comino, E., Bona, F., Riggio, V.A., 2013. Growth of three microalgae strains and nutrient removal from an agro-zootechnical digestate. *Chemosphere*, 92(6): 738–744.
 14. Gentili, F.G., 2014. Microalgal biomass and lipid production in mixed municipal, dairy, pulp and paper wastewater together with added flue gases. *Bioresour. Technol.* 169: 27–32.
 15. Ji, F., Liu, Y., Hao, R., Li, G., Zhou, Y., Dong, R., 2014. Biomass production and nutrients removal by a new microalgae strain *Desmodesmus* sp. in anaerobic digestion wastewater. *Bioresour. Technol.* 161: 200–207.
 16. Ji, M.K., Yun, H.S., Park, Y.T., Kabra, A.N., Oh, I.H., Choi, J., 2015. Mixotrophic cultivation of a microalga *Scenedesmus obliquus* in municipal wastewater supplemented with food wastewater and flue gas CO₂ for biomass production. *J. Environ. Manage.* 159, 115–120.
 17. Ji, G.D. Sun, T. H. and Ni, R. J. (2007). Surface Flow Constructed Wetland for Heavy Oil – Produced Water Treatment. *Bio. Techno*; 98: 436-441.
 18. Khalaf, M.A., 2008. Biosorption of reactive dye from textile wastewater by nonviable biomass of *Aspergillus niger* and *Spirogyra* sp. *Bioresour. Technol.* 99(14): 6631–6634.
 19. Loutseti, S., Danielidis, D.B., Economou-Amilli, A., Katsaros, C., Santas, R., Santas, P., 2009. The application of a micro-algal/bacterial biofilter for the detoxification of copper and cadmium metal wastes. *Bioresour. Technol.* 100(7): 2099–2105.
 20. Malik A. 2004. Metal bioremediation through growing cells. *Environ Inter*, 30: 261–278.
 21. Marungrueng, K., Pavasant, P., 2007. High performance biosorbent (*Caulerpa lentillifera*) for basic dye removal. *Bioresour. Technol.* 98(8): 1567–1572.
 22. Nayyef, M. Azeez and Amal A. Sabbar (2012). Efficiency of *Lemna minor* L. In The Phytoremediation of Waste Water Pollutants from Basrah Oil Refinery. *Journal of*

- Applied Biotechnology in Environmental Sanitation*. 2012; 1(4): 163-172.
23. Nriagu, J.O. (1996). Toxic Metal Pollution in Africa. *Science*, 223: 272.
24. Prandini, J.M., da Silva, M.L., Mezzari, M.P., Pirolli, M., Michelon, W., Soares, H.M., 2016. Enhancement of nutrient removal from swine wastewater digestate coupled to biogas purification by microalgae *Scenedesmus* spp. *Bioresour. Technol.* 202: 67–75.
25. Patel, D.K. and Kanungo, V.K. (2010). Phytoremediation Potential of Duckweed (*Lemna minor* L.: A Tiny Aquatic Plant) In The Removal Of Pollutants From Domestic Wastewater With Special Reference To Nutrients. *The Bio Sci.* 2010; 5(3): 355- 358.
- a. Panico, A. Basco, G. Lanzano, F. Pirozzi, F. Santucci de Magistris, G. Fabbrocino, E. Salzano (2016) Evaluating the structural priorities for the seismic vulnerability of civilian and industrial wastewater treatment plants. *Safety Sci.* (2016), <http://dx.doi.org/10.1016/j.ssci.2015.12.030>.
26. Roy D, Greenlaw PN, Shane BS. 1993 Adsorption of heavy metals by green algae and ground rice hulls. *J Environ Sci Health Part A.* 28: 37–50.
27. Subhashini V, Swamy A.V.V.S And Hema K.R. (2013). Phytoremediation Emerging And Technology For The Uptake Of Cadmium The Contaminated Soil By Plant Species. *Int Journal Of Environ*, 2003; (4): 0976-4402.
28. Suzy K.Z, Ezra A, G., A. Abdulhameed and Nayyaya A.J (2016) Study of Heavy Metal and Physicochemical Attribute of River Romi, Kaduna, Nigeria. *International Journal of Fisheries and Aquatic Studies*, 4(3): 518-522.
29. Sun, J., Hu, Y.Y., Li, W.J., Zhang, Y.P., Chen, J., Deng, F., 2015. Sequential decolorization of azo dye and mineralization of decolorization liquid coupled with bioelectricity generation using a pH self-neutralized photobioelectrochemical system operated with polarity reversion. *J. Hazard. Mater.* 289: 108–117.
30. United States Environmental Protection Agency (USEPA) (1997). Cleaning up the Nation's Waste Sites: Markets and Technology Trends. EPA/542/R-96/005. Office of Solid Waste and Emergency Response, Washington, DC.
31. Ugya, A.Y, Imam, T.S and Tahir, S.M (2016). The Role of Phytoremediation in Remediation of Industrial Waste. *World Journal of Pharmaceutical Research*, 5(12): 1403-1430.
32. Ugya A.Y, Tahir S.M., and Imam T.S (2016b) *Emerging Trend in the Remediation of Pollution*. First Edition, ISBN-978135529177: Lulu Publishers.
33. Ugya A.Y and Imam T.S (2016b) *Wastewater Remediation Using Plants Techniques*. First Edition Lulu Publishers ISBN- 9781365547164.

34. Ugya, A.Y. and T.S. Imam (2015). The efficiency of *Eicchornia crassipes* in the phytoremediation of waste water from Kaduna Refinery and petrochemical company. *IOSR J. Environ. Sci. Toxicol. Food Technol.*, 9: 43-47.
35. Ugya, A.Y., (2015). The efficiency of *Lemna minor* L. in the phytoremediation of Romi stream: A case study of Kaduna refinery and petrochemical company polluted stream. *J. Applied Biol. Biotechnol.*, 3: 11-14.
36. Ugya, A.Y., S.M. Tahir and T.S. Imam, (2015a). The efficiency of *Pistia stratiotes* in the phytoremediation of Romi stream: A case study of Kaduna refinery and petrochemical company polluted stream. *Int. J. Health Sci. Res.*, 5: 492-497.
37. Ugya, A.Y., T.S. Imam and A.S. Hassan, (2015c). The use of *Ecchornia crassipes* to remove some heavy metals from Romi stream: A case study of Kaduna Refinery and Petrochemical company polluted stream. *IOSR J. Pharm. Biol. Sci.*, 10: 43-46.
38. Ugya, A.Y., T.S. Imam and S.M. Tahir, (2015d). The use of *Pistia stratiotes* to remove some heavy metals from Romi stream: A case study of Kaduna Refinery and Petrochemical company polluted stream. *IOSR J. Environ. Sci. Toxicol. Food Technol.*, 9: 48-51.
39. Ugya, A.Y., I.M., Toma. and Abba A. (2015b) Comparative Studies on the Efficiency of *Lemna minor* L., *Eicchornia crassipes* and *Pistia stratiotes* in the phytoremediation of Refinery Waste Water. *Sciences World Journal*, 10(3).
40. Ueshima, M., Ginn, B.R., Haack, E.A., Szymailowski, J.E.S., Fein, F.B., 2008. Cd adsorption onto *Pseudomonas putida* in the presence and absence of extracellular polymeric substances. *Geochim. Cosmochim. Acta*, 72(24): 5885–5895.
41. Van Den Hende, S., Carre, E., Cocaud, E., Beelen, V., Boon, N., Vervaeren, H., 2014. Treatment of industrial wastewaters by microalgal bacterial flocs in sequencing batch reactors. *Bioresour. Technol.* 161: 245–254.
42. Vijayaraghavan, K., Yun, Y.S., 2008. Bacterial biosorbents and biosorption. *Biotechnol. Adv.* 26(3): 266–291.
43. Wang, M., Yang, H., Ergas, S.J., van der Steen, P., 2015a. A novel shortcut nitrogen removal process using an algal-bacterial consortium in a photo-sequencing batch reactor (PSBR). *Water Res.* 87: 38–48.
44. Wang, Y., Shih-Hsin Ho, Chieh-Lun Cheng, Wan-Qian Guo, Dillirani Nagarajan , Nan-Qi Ren, Duu-Jong Lee, Jo-Shu Chang (2016) Perspectives on the feasibility of using microalgae for industrial wastewater treatment. *Bioresource Technology*, 222: 485–497.
45. Wu, Y., Hu, Z., Yang, L., Graham, B., Kerr, P.G., 2011. The removal of nutrients from

- non-point source wastewater by a hybrid bioreactor. *Bioresour. Technol.* 102(3): 2419–2426.
46. Xiaomei, Lu., Maleeya Kruatrachue, Prayad Pokethitiyook and Kunaporn Homyok (2004). Removal Of Cadmium And Zinc By Water Hyacinth, *S. Scienceasia*; 30: 93-103.
47. Xiong, J.Q., Kurade, M.B., Abou-Shanab, R.A.I., Ji, M.K., Choi, J., Kim, J.O., Jeon, B.H., 2016. Biodegradation of carbamazepine using freshwater microalgae *Chlamydomonas mexicana* and *Scenedesmus obliquus* and the determination of its metabolic fate. *Bioresour. Technol.* 205: 183–190.
48. Xu, M., Li, P., Tang, T., Hu, Z., 2015. Roles of SRT and HRT of an algal membrane bioreactor system with a tanks-in-series configuration for secondary wastewater effluent polishing. *Ecol. Eng.* 85: 257–264
49. Zhou, D., Niu, S., Xiong, Y., Yang, Y., Dong, S., 2014. Microbial selection pressure is not a prerequisite for granulation: dynamic granulation and microbial community study in a complete mixing bioreactor. *Bioresour. Technol.* 161: 102–108.
50. Ugya A.Y and Ahmad A.M (2016):The significance of Refinery Effluent Remediation in the Conservation of Aquatic Ecosystem: In book: *Biodiversity Conservation in Changing Climate*, First Edition, Chapter: 16, Lenin Media Private Limited, Delhi, India, Editors: Dr M.M Abid Ali Khan, Murtaza Abid, Prof. M.S Naqvi, Dr. Abideen Mustafa Omer, Dr B. Rani, Dr. S.N Haider, pp.16.