

MACROPHAGES AS A KEY FACTOR IN THE PROCESS OF TUMOUR FROM THE INITIATION TO THE THERAPY OF CANCER.

Stephanie Stella Mica Avoulou Meyo^{1,a}, Xianjing Li^{a,1}, Nadia Kabir^{a,2}, Muhammad Sohail^{a,2} and Yong Yang*^{a,2}

^aState Key Laboratory of Natural Medicines, Jiangsu Key Laboratory of Drug Discovery for Metabolic Disease, Center for New Drug Safety Evaluation and Research, China
Pharmaceutical University, Nanjing 211198, China.

Article Received on
19 Feb. 2017,

Revised on 13 March 2017,
Accepted on 02 April 2017

DOI: 10.20959/wjpr20175-8182

*Corresponding Author

Prof. Yong Yang

State Key Laboratory of
Natural Medicines, Jiangsu
Key Laboratory of Drug
Discovery for Metabolic
Disease, Center for New
Drug Safety Evaluation and
Research, China
Pharmaceutical University,
Nanjing 211198, China.

ABSTRACT

The tumor environment is a complex ecological system made up of cells and serves as support to the tumor cells from their implantation to the state of malignancy. Among all the cells of immunity (innate and adaptive) macrophages are those most present on the site of the tumor, playing a predominant role in all stages where they intensively promote the different aspects of tumor initiation, growth and development. Macrophages are composed of two subpopulations M1 (anti-tumor, responsible of the inflammation response and pathogen clearance) and M2 (pro-tumor and anti-inflammatory activities) both depending of diverse external factors. Clinical studies and experimental models have shown that tumor associated macrophages (TAMs) are more related to the M2, giving a poor clinical prognostic by promoting the growth of the tumor and assessing a favorable microenvironment to the malignancy. Thus, this ability to promote

tumor proliferation makes these TAMs a key tool targets for curative cancer solutions via translational approaches, but more research need to be done to ensure a better prognosis.

KEYWORDS: Macrophages M1/M2, tumor associated macrophages, macrophage polarization, Cancer, immunotherapy.

1-INTRODUCTION

Macrophages are innate immune cells that play a broad role in host defense and the maintenance of tissue homeostasis.^[1] Tissue-resident and inflammatory macrophages

originate from circulating bone marrow-derived monocytic precursors.^[2] The complex microenvironment of the tumor involves since its initiation, a large number of malignant and non-malignant cells as well as cancer cells with Stroma and some of the host cells Where they all interact to promote the growth of the tumor in a synchronic way. The tumoral environment is composed of fibroblasts, endothelial cells and complex inflammatory infiltrates, containing different cells of the haematopoietic system (neutrophils, monocytes / macrophages). Monocytes are known to originate in the bone marrow from a common myeloid progenitor that is shared with neutrophils, and they are then released into the peripheral blood, where they circulate for several days before entering tissues and replenishing the tissue macrophage populations.^[3] Tumor associated macrophages (TAMs) constitute a major part of the leukocytes infiltrate in human cancer. Substantial evidence indicates that macrophages rather than being tumoricidal as suggested after their activation *in vitro*.^[4] adopt a pro-tumoral phenotype *in vivo* both in the primary and metastatic sites.^[5] For example lung cancer macrophages are polarized to a pro-tumoral phenotype at the time of tumor initiation.^[6] These precursor cells extravasate into target tissues where they differentiate into mature macrophages and polarize into diverse subsets that have different phenotypes in response to micro-environmental challenges.^[7] These activities include suppression of T cell responses.^[8,9] In addition, macrophages promote many important features of tumor progression including angiogenesis, tumor cell invasion, motility and intravasation as well as at the metastatic site, stimulation of tumor cell extravasations and persistent growth.^[9] Each of these activities is delivered by an identifiable sub-population of macrophages.^[9] Furthermore, it has been estimated that 80% of studies that have tried to relate TAM density to prognosis in any type of cancer have found a negative correlation while less than 10% have found a positive correlation.^[10, 11] The inhibition of tumor and metastasis progression by the ablation of macrophages as shown by several experiments and data proving that the presence of immune cells is very important for the tumor development which can lead to a malignant state or more deeply their acquisition of a malignant phenotype. Which could therefore be a potential target in the search for a curative solution of cancer. Thus, the purpose of this review will discuss the function of diverse macrophage subpopulations showing how the tumor microenvironment takes advantage of macrophage plasticity to mold an immunosuppressive population and their dynamic interplay with tumor cells that confer these pro-tumoral activities and give particular emphasis to the immunoregulatory role of these cells.

2- MACROPHAGES

Macrophages are monocyte-derived myeloid cells that develop from a common myeloid progenitor cell residing within the bone marrow of adult mammals ^[12]. This will also give rise to other myeloid cells, specifically neutrophils, eosinophils, basophiles and dendritic cells, all of which will be exposed to cytokines during their maturation. Like others, the monocytes are derived from this same progenitor site, this during a cascade of events involving granulocyte-macrophage colony stimulating factor (GM-CSF), granulocyte colony stimulating factor (G-CSF) and macrophage colony stimulating factor (M-CSF), leading to a differentiation of hematopoietic stem cells into monoblasts, pro-monocytes and finally into monocytes. Mature monocytes leave the nest to the bloodstream for about 1-3 days and finally enter the different tissues and become macrophages for a steady-state renewal process or due to chemo attractants produced in the framework of a local inflammatory treatment. Prior to entering tissues and differentiating into macrophages, circulating blood monocytes are known to be heterogeneous with at least two general populations identified. ^[13,14,15] Among which CD14^{hi}, CD16⁻ and CD14⁺, CD16⁺ in humans, Ly6C^{high} (GR1⁺) with Ly6C^{low} (GR1⁻) in mice. These phenotypes are respectively divided into two groups: inflammatory (CD14^{hi}, CD16⁻ / Ly6C^{high}) with the ability to migrate to the lesion or infected site mediated by the ligand C-C chemokine CC (CCR) 2-chemokine CC and to spread chronic inflammatory diseases and the group of resident who patrol the vascular system, regulate the inflammatory system And stay in the tissues. The gene expression of each macrophage population is related to the specific microenvironment of tissue in which they reside.

Essential markers of the state of health, macrophages derived from innate immunity and are known to be heterogeneous. There are two polarization states of the macrophages: Type 1 activated classically (M1) and type 2 (M2) “fig.1” The macrophages with M1 phenotype is adapted to attract and activate cells of the adaptive immune system in response of the danger signals delivered by bacterial products or interferon (IFN γ). Important features of M1 macrophages are the expression of nitric oxide synthase (iNOS), reactive oxygen species (ROS) and the production of natural killer (NK) and type 1 T-cell stimulating cytokine IL-12. Beside M2 macrophages can develop in response to for instance IL-4 or IL-13. ^[16,17] M2 play a role in parasite clearance and wound healing where they also polarize T-cells to Th2 and dampen immune responses ^[18]. Even though less effective than dendritic cells, macrophages are antigen-presenting cells (APC) that express human leukocyte antigen (HLA) class I and class II molecules and co-stimulate / inhibit molecules to give instructions to T cells.

Macrophages display great plasticity and can adapt to a large number of activation states ranging between the M1 or M2 phenotype depending on the mix of signals in their direct microenvironment. In addition, fully polarized M1 and M2 macrophages can be redirected *in vitro* towards the opposite functional phenotype by treatment of the cells with cytokines.^[19]

3- THE CONCEPT OF TUMOR ASSOCIATED MACROPHAGES

Macrophages play a prominent role in the stroma and leukocyte compartment in malignancy, distinct macrophage subsets in cancer have been described.^[20,21,22,23] Since macrophages in human cancer cannot be classified into activated classical macrophages of type M1 or alternatively activated as M2, they are called macrophages associated with tumors (TAM). Depending on their activation state, TAMs can present both pro-tumoral and anti-tumoral functions. Macrophages cell have an extreme functional plasticity enabling them to integrate and respond to different stimuli^[24]. After establishing the mechanism of tumor progression, TAMs often express an M2-like phenotype, promote tumor growth through angiogenesis, tissue remodeling, suppression of adaptive immunity^[25, 26] and promote metastasis “fig.2”

4-MECHANISMS OF MACROPHAGE RECRUITMENT AND MACROPHAGE POLARIZATION IN TUMOR PROGRESSION

Macrophages are among the first immune cells to infiltrate the pre-invasive tumorous lesions and persist during the development into invasive cancer.^[27] As has been shown in previous studies, cancer is associated with inflammation that leads to the recruitment of cells derived from bone marrow^[28, 29]. Certain growth factors such as colony stimulating factor 1 (CSF-1), vascular epithelial growth factor (VEGF), but also monocyte, chemotactic protein 1 (MCP-1), many other chemokines CCL and other molecules induce chemotaxis of monocytes in the tumor microenvironment. An explicit study in three different murine tumor models (the BALB / C4T1 mammary tumor model, the BALB / c mammary adenocarcinoma TS / A model and the C57BL / 6 3LL lung carcinoma model) the inflammatory Ly6Chi monocyte subset was shown to be the TAM subsets in the tumor^[30]. With regard to the specific tumor, the molecules are secreted by the tumor cells themselves or the adjacent stromal cells. In cutaneous carcinogenesis, for example, a carcinogenic application induces a proliferation of fibroblasts which secrete monocyte chemottractant protein-1(MCP-1), resulting in chemotaxis of the macrophages. Neutralization of MCP-1 almost completely blocks this accumulation of macrophages in the tumor.^[31] Vascular endothelial growth factor (VEGF), beside its strong angiogenic role, recruits monocytes into the tumor microenvironment and

blockade of VEGF by bevacizumab results not only in reduced vessel density, but also in reduced TAM infiltration.^[32] Once on the site of the tumor, monocytes differentiate into macrophages under the influence of macrophage colony stimulating factor (M-CSF) and others factors derived from tumor cells result in the polarization of macrophages. In general the induction of a TAM related phenotype has been brought in context with tumor cell derived mediators such as M-CSF^[33], IL-4, IL-10, IL-6.^[34,35] transforming growth factor β (TGF- β 1), prostaglandin E2 (PGE2)^[36,37], hyaluron fragments.^[32], and the leukemia inhibitory factor (LIF)^[38]. In cervical cancer, monocytes are skewed from a dendritic (DC) towards a macrophage phenotype by the production of PGE2 and IL-6 from tumor cells.^[39,40]

Similarly, TAMs are derived from circulating monocytes or tissue-resident macrophages.^[41] Cytokines, chemokines, extracellular matrix (ECM) components, and hypoxia are the multiple microenvironmental signals responsible for the mobilization of macrophages in tumor tissues.

4.1 SOLUBLE FACTORS MEDIATING MACROPHAGE MOBILIZATION INTO TUMORS

Recruitment of macrophages at the tumor site is mainly regulated by cytokines, chemokines and growth factors that are derived from tumor and stromal cells of its microenvironment. Monocytes/macrophages migrate toward inflamed tissues under the influence of CCL2 (MCP-1).^[42,43] Experimental studies have revealed the CCL2-dependent infiltration of macrophages into tumors.^[44] For example, systemic administration of CCL2-neutralizing antibodies in tumor-bearing mice or the short-hairpin RNA knockdown of CCR2 in the cancer cell lines significantly reduced tumor growth along with reduced TAM recruitment^[45]. Along with the chemokines, several cytokines such as colony-stimulating factor-1 (CSF-1) and endothelial monocyte-activating polypeptide II (EMAPII) have been implicated in the recruitment of monocytes into tumors.^[45, 46] Indeed, an elevated CSF-1 level correlates with marked macrophage infiltration in human metastatic breast cancer.^[48] Xenotransplantation experiments have further demonstrated that tumor cells transected with CSF-1 gene exhibited an increase in TAM infiltration.^[49] Some growth factors as well as vascular endothelial growth factor (VEGF), endothelin 2, and platelet-derived growth factor (PDGF) have also been reported to promote monocyte/macrophage recruitment.^[50, 51] Within the dense tumor microenvironment, the orchestrated actions of these soluble factors can synergistically speed

up the mobilization of macrophages and the change of these cells to TAMs, which lead to promote alterations in the tumor microenvironment.

4.2 ROLES OF EXTRACELLULAR MATRIX (ECM) COMPONENTS AND THEIR FRAGMENTS IN MACROPHAGE RECRUITMENT

Strands of the structures in and are believed to play essential roles in the formation of ECM during an inflammatory reaction.^[52] After binding to high molecular weight hyaluronic acid (HA), vesicant also cooperatively improves the adhesion of leukocytes to these cables, suggesting that the per cellular HA complex provides suitable support for macrophage recruitment. Our recent study demonstrated the preferential engagement of immunosuppressive M2 macrophages in a tumor microenvironment. Extracellular Matrix (ECM) serves as a structural support for the infiltration of innate immune cells and hyaluronic acid (HA) which is his major component. De la Motte and colleagues have shown that cable-type ECM structures have been formed after extensive HA deposition on smooth mucosal muscle cells treated with poly (I: C); these structures have been implicated in adhesion and recruitment of monocytes through association with the CD44 HA receptor^[53]. Numerous HA-binding partners, such as the inter- α -inhibitor heavy chain (I α I), the tumor necrosis factor (TNF), stimulated gene 6 (TSG-6) and the vesicant HA-binding proteoglycan, are located along the rich in HA and vesicant.^[54]

It has been shown that certain ECM molecules and their proteolytic fragments acted as inflammatory stimuli for the recruitment of innate immune cells thus leading to pro-inflammatory action. For example, elastin fragments generated by macrophage-derived matrix metalloproteinase (MMP)-9/12 exhibit a monocyte chemo tactic activity.^[55] On other hand, TAM-derived oncofetal fibronectin not only promotes cancer cell invasion but also stimulates monocyte migration.^[56] Soluble biglycan and its fragments act on macrophages to produce both TNF- α and macrophage inflammatory protein-2 (MIP-2) in a manner that is dependent on TLR2 and TLR4, and thereby play a positive role in macrophage recruitment and activation.^[57] Oligosaccharides generated by hyaluronidase-catalyzed digestion of high-molecular-weight HA also utilize both TLR2 and TLR4 to stimulate inflammatory gene expression in macrophages and act as an endogenous danger signal.^[58] Tumor-derived HA fragments have also been shown to promote the development of immunosuppressive M2 macrophages by triggering a transient early activation of monocytes.^[59]

4.3 MACROPHAGE RECRUITMENT INTO HYPOXIC AREAS

Several pieces of evidence have shown that advanced solid tumors exhibit hypoxic areas within the tumor mass and that a high number of TAMs accumulate in the avascular areas of a wide range of human tumors.^[60] VEGF-A,^[61] endothelin-2,^[62] and EMAPII.^[63] were involved in the Hypoxia induced by the recruitment of macrophages. The suppression of VEGF-A derived from the myeloid has reduced vascularization in solid tumors.^[64] When TAM reaches hypoxic areas, hypoxia directs them to a pro-tumorigenic phenotype by altering gene expression profiles. Hypoxia-inducible factor (HIF)-1 α is a key transcription factor that regulates hypoxia-induced gene expression^[65, 66]. HIF-1 α also induces CXCL12 expression in direct proportion to the reduced oxygen tension at hypoxic sites.^[67]

5- MACROPHAGES IN CANCER INITIATION AND PROMOTION

There is a reason to believe that inflammation is the cause of many cancers. Mantovani and colleagues have called this the seventh hallmark of cancer and reviewed its characteristics as well as the epidemiological and infectious disease literature that supports this hypothesis^[68]. Direct experimental support demonstrates the role of inflammation in cancer initiation, for example in a mouse model of lung cancer, bronchial exposure with H.influenzae lysate results in inflammation in the lung and an increase in tumorigenesis.^[69] Myeloid-specific ablation of integrin α V also results in an ulcerative colitis that induces colonic tumors.^[70] Ablation in myeloid cells of *stat3*, a transcription factor which function suppresses inflammatory responses because it is a major target of immunosuppressive cytokine IL-10.^[71] causes inflammation in the colon. This is associated with abundant expression of TNF α and IL-6 by macrophages and results in a chronic colitis and invasive colonic adenocarcinoma.^[72] The causality of the inflammation in carcinogenesis in these studies comes from experiments in which suppression of the bacterial flora by antibiotic treatment reduces the inflammation and inhibits tumorigenesis.^[73, 74, 75] These data show that the immune system is normally in equilibrium but once the negative controls of immune responses are compromised, a persistent inflammatory response to normally normal organisms results. This inflammation in turn creates a tumor that settles gradually.

The inflammatory state in myeloid cells is controlled by the transcriptional factors NF- κ B and *stat3* that work in opposition to one another.^[76-71] NF- κ B is a central transducer signals that cause inflammation downstream of TLR activation. In the inflammatory responses associated with cancer initiation, NF- κ B signaling is essential for the inflammatory

phenotype.^[77] Inhibition of this activity through ablation of I κ B kinase α (IKK α) in myeloid cells in mouse models of intestinal cancer reduces inflammation and inhibits tumor progression.^[78]

The type of inflammation associated with increased cancer risk because of chronic infection or persistent irritation is often called “smoldering inflammation”.^[79] Activated macrophages are central to this type of immune response and work in concert with other immune cells.^[80]

NO reacts with peroxidate to give nitrosoperoxy carbonate, and this reaction is a key of the chemistry of inflammation. This highly reactive compound and other products cause mutations in the adjacent epithelial cells^[81, 82]. In addition, there is evidence that the inflammatory microenvironment also promotes genetic instability within the developing tumor epithelial cells^[83]. The mutations are fixed after replication of the epithelial cells in all case, one which is stimulated by growth factors synthesized by the infiltrating or resident immune cells which include macrophages. These growth-promoting effects on tumors are caused by the production of IL-6 in hepatocellular carcinoma (HCC).^[84; 85] and TNF α .^[86] and IL-6 in colitis associated cancers.^[87] Interestingly, IL-6 synthesis in Kupffer cells in response to inflammation-induced liver damage is gender dependent with males who have increased risk of HCC having elevated levels. IL-6 is also required for the increased risk of HCC in female mouse models.^[88] However, once initiated and as the tumors progress toward malignancy, the macrophage phenotype changes from the “inflammatory” type to one that promote tissue formation during development.^[89, 90] In established tumors, NF- κ B signaling is inhibited by the constitutive expression of p50 homodimers that negatively regulate NF- κ B and the macrophages display the M2/trophic phenotype which reduce iNOS and TNF α expression.^[91] The blocking of the NF- κ B function by the inhibition of IKK α in the cultured macrophages reduces the expression signature of the inflammatory gene and pushes the cells to the trophic / M2 type. This transition from stimulated to inhibited NF κ B function between initiation and established tumor stages is poorly understood. The type “activated / trophic” macrophage is also found in cancers that appear in the apparent absence of inflammation, such as breast cancer, in which macrophages are recruited in benign tumors in large numbers as well as the tumor transition to malignancy.

6-IMMUNOSUPPRESSIVE PROPERTIES OF TAMs

Immunosuppressive functions of TAMs appear to be conflicting to the inflammatory tumor-microenvironment and the inflammatory properties of macrophages. Even though TAMs

produce several chemokines that are associated with inflammation in immunity against pathogens, under sterile inflammatory conditions in cancer these molecules exert a growth promoting influence on tumor cells. This effect might be due to *STAT3* activation in TAMs opposing *STAT1* driven Th-1 anti-tumor responses.^[92; 93] Expression of MHC class II molecules on TAMs is actively down regulated by tumor cell derived TGF- β 1, IL-10 and PGE2^[94]. The direct inhibition of immune responses by TAM has also been described.^[95] IL-10 and TNF- α in the tumor microenvironment induce the expression of programmed death ligand 1 (PD-L1) also called B7-H1 on the membrane of macrophages associated with tumors. Although naive T lymphocytes can be stimulated via PD-L1, his most important role is the inhibition of PD-1 receptor activated T lymphocytes^[96]. Allavena and Mantovani have demonstrated an additional indirect mechanism of creating an anti-inflammatory tumor medium, namely the recruitment of other non-inflammatory immunity cells into the microenvironment of tumors^[97]. In the same sense, CCL17 and CCL22 produced by TAMs are involved to primarily attract Th2 and Treg which are non-cytotoxic cells.^[98-99] Macrophage-derived CCL18 recruits rather naïve T-lymphocytes that are primed under immune regulation conditions^[100]. In a murine model of colorectal cancer, it was found that F4 / 80 + TAM secretes large amounts of CCL20 attracting CCR6⁺ cells on the tumor side.^[101]

7-MACROPHAGES AND THE METASTATIC SITE

Cancer is considered to be a systemic disease, and primary tumors secrete factors that influence metastatic outcome at remote sites. When the tumor cells escape from the primary site, they pass through the lymphatic and / or circulatory system and eventually some establish at distant sites to create metastases. These sites vary depending on the cancer, for example in the breast; they mainly go to the bone then the lung and the brain. This process is essential to understand that 95% of deaths from solid tumors in developed countries are due to metastasis.

Monocytes/macrophages are essential metastasis promoters acting both to prepare sites and also to promote the extravasation, the survival and the persistent growth of metastatic cells^[102, 103]. These niches are populated by CD11b⁺, VEGFR1⁺ myeloid cells whose recruitment is promoted by Lysyl Oxidase (LOX) and S110A and whose ablation inhibits the formation of these sites.^[104] Several other factors have been shown to be important for pre-metastatic niche formation, most recently, tumor derived exosomes that program the myeloid cells to be pro-

tumoral and pro-angiogenic through activation of the receptor tyrosine kinase MET.^[105] Platelets are also key elements in the formation of niches, which presumably deposit fibrin in the target tissues to attract the myeloid cells. Thus, pre-metastatic niche formation is blocked by anticoagulants. Gil-Bernabe and colleagues found that tumor-derived tissue factor (coagulation factor III or CD142) stimulated clot formation and enhanced subsequent tumor cell survival at the metastatic site by recruiting CD11b⁺/CD68⁺/F4/80⁺/CX3CR1⁺ macrophages.^[106] LOX (lysyl oxidase), a copper-dependent amine oxidase, maintain the ECM network by cross-linking collagen and elastin. LOX secreted from tumor cells forms the cross-links of collagen IV in the basement membranes at the pre-metastatic sites. CD11b⁺ myeloid cells then adhere to the cross-linked collagen IV and produce MMP-2. The collagen IV peptide cleaved by MMP-2 then enhances the further recruitment of CD11b⁺ cells as a chemoattractant.^[107] By increasing the extracellular activity, matrix remodeling and the creation of the pre-metastatic niche will be effective with this positive feeding loop.

Hiratsuka and colleagues established that primary tumors stimulate the expression of S100A8 and S100A9 proteins in the lung by secreting VEGF-A, TNF- α and TGF- β . The S100A8 and S100A9 proteins both generate the enrollment of macrophage antigen 1 positive myeloid cells (Mac1) into the pre-metastatic microenvironment. Neutralization of these factors with specific antibodies blocked the infiltration of Mac1⁺ myeloid cells and the migration of cancer cells from primary tumors to the lung.^[108] telling that the S100A8 and S100A9 proteins could play a critical role in the creation of the pre-metastatic niche. They also found that S100A8 and S100A9 induced the expression of serum amyloid A3 in alveolar macrophages as well as in endothelial cells.^[109] Broadly speaking, mononuclear phagocytes appear not only to set up preferred sites for seeding metastatic cells but also to improve extravasation of tumor cells, the formation and future growth of metastatic lesions.

8-MACROPHAGES AS ATTRACTIVE TARGETS FOR THERAPEUTIC INTERVENTION AGAINST CANCER

Evidence of many researches has demonstrated that TAM accumulation is associated with poor clinical prognosis and resistance to cancer therapy^[110,111]. However, preclinical data and sustained studies on the activity of macrophages in a solid tumor demonstrate that it significantly decreases by inhibiting CSF-1R by blocking the monoclonal antibodies. Some human monoclonal antibody in the example of RG7155 strongly inhibits the dimerization of CSF-1R. The clinical activity of RG7155 was evaluated in patients with diffuse-type giant

cell tumors and showed a marked reduction in the population of CSF-1R⁺, CD163⁺ macrophages in tumor tissues.^[112] PLX3397, a potent CSF-1R tyrosine kinase inhibitor, improved the efficacy of immunotherapy by decreasing macrophage infiltration and activating tumor-infiltrating lymphocytes.^[113] Other studies have also demonstrated that targeting inhibition of macrophage recruitment, conversion of pro-tumorigenic M2 to antitumor M1 phenotype and suppression of TAM survival is another possible option for intra-tumor manipulation of macrophages. In spite of the fact that the reduction of the presence of macrophages associated with tumors is the main objective of these clinical studies, it should be noted that they will not all be eradicated, but rather in the sense of reprogramming them to a phenotypic anti-Tumors where they will support T-cell responses. As noted above, chemo attractants derived from the tumor and microenvironment are facilitators in the recruitment of macrophages into tumors. Thus, the inhibition of this step could become a more effective anti-cancer therapy in breast cancer animal models, the regulation of macrophages via chemotaxis is the prominent strategy to develop anti-tumorigenic therapeutic agents.^[114] For example, after treatments with the receptor antagonist met-CCL5, both the number of infiltrating macrophages and the size of tumors were significantly decreased.^[115] Pharmacological drugs, such as zoledronic acid combined with sorafenib, control the population of macrophages in a tumor microenvironment and enhance antitumor effects.^[116] Several other pharmacological drugs are believed to block macrophage infiltration and show anti-tumoral effects, including thalidomide, linomide, pentoxifylline and genistein.^[117]

The conversion of macrophages of type M2 into macrophages in M1 to induce pro-inflammatory responses could be considered as the best therapeutic means. In addition, some studies have revealed that TLRs are potent candidates for enhancing the response to macrophages polarized by M1. Another recent study shows that an enzyme, SHIP1 is a crucial phosphatase that plays an important role in the conversion of the M1/M2 macrophage paradigm.^[118] Moreover, the accumulated results show that macrophages are prime mediators which deliver therapeutic inducers to the environment of the tumor. Moreover, a plant-derived diterpenoid (paclitaxel) stimulates macrophages to express high levels of NO, TNF- α , and IL-1 β that enhance tumor cell cytotoxicity as well as restore IL-12 production by macrophages.^[119] Recently, it was reported that an anti-PD-L1 antibody, which blocks the PD-1/PD-L1 pathway, can improve macrophage-mediated T cell activation and has progressed to clinical trials.^[120]

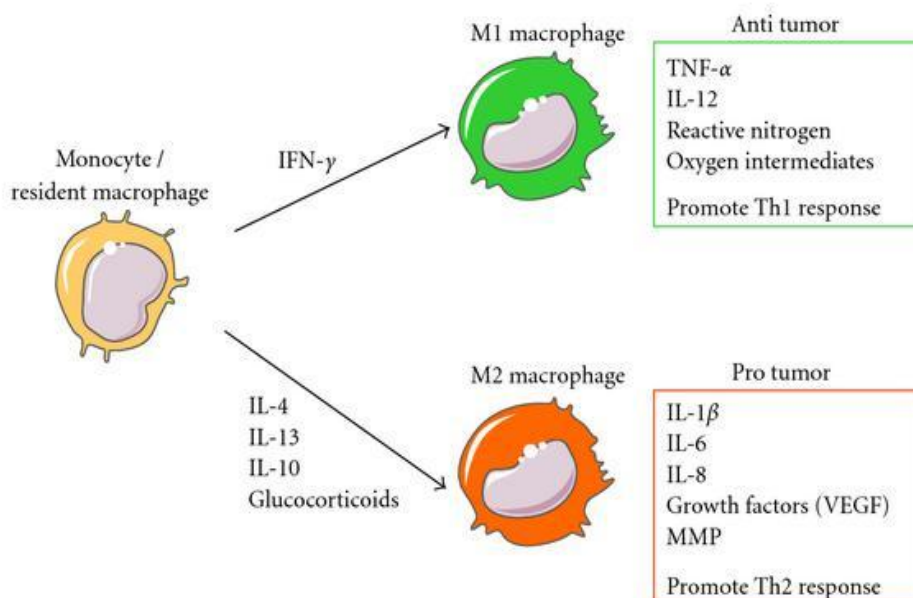


Fig.1 Macrophage polarization. The macrophages will polarize into two distinct populations under the effect of several micro environmental factors (interferon, interleukin 4, 13, 10 and glucocorticoids) classically activated macrophages or M1 with an anti-tumor function and alternatively activated macrophages M2 responsible of the survival and expansion of the tumor. By the promoting the t helper cells (th1 et th2).

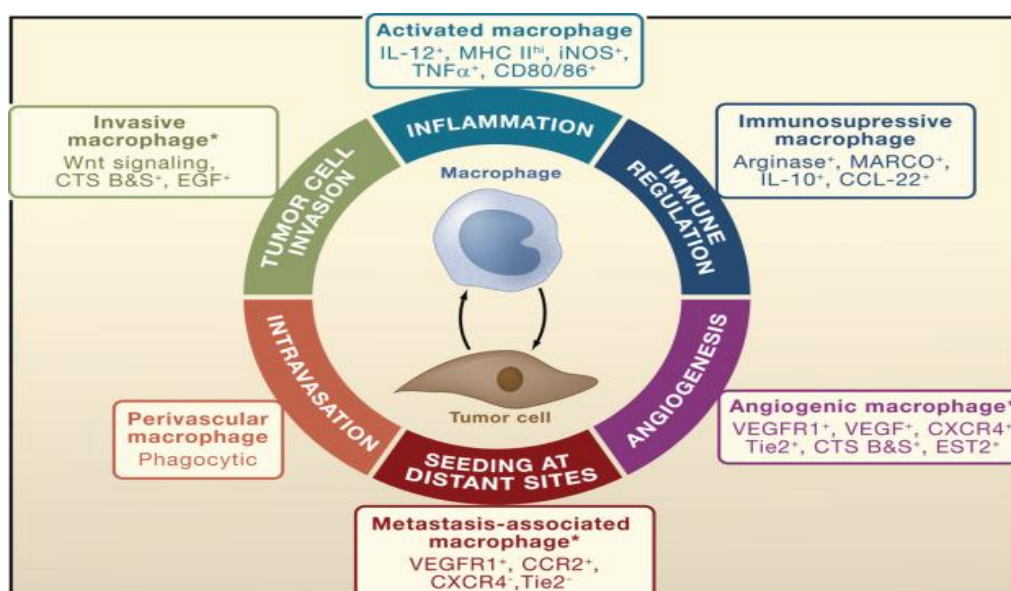


Fig.2 Summary of the function of TAMs in the tumor.

9-CONCLUSION

Macrophages play an essential role from the implantation to the treatment of cancer. Their heterogeneity and plasticity induce the tumor growth, suppress local immunity or attack the

tumor cells and maintain tumor immunity. Macrophages with the support of the microenvironment polarized into substantial sub-population M1 and M2 with a dual effect pro-tumoral and anti-tumoral responsible of the presence of TAM. TAMs are usually associated with an advanced poor prognostic because of his pro-tumoral function. Overall, these finding enhance the fact that macrophages or precisely TAMs are biomarkers for a better understanding of the tumor physiology and therapy. Targeting TAMs or re-polarization can be a good strategy in cancer therapy. Therefore, the ablation of or re-differentiation of macrophages within the tumor microenvironment will become an important tool of combination therapies designed to cure cancer. Strategies based on blocking the tumor-promoting activities of TAMs, and exploitation of macrophage anti-tumor effects or functions will allow the development of promising novel cancer treatment.

10- CONFLICT OF INTEREST

We have no conflict of interest to claim.

11- REFERENCES

1. Gordon, S.; Martinez, F.O.; Alternative activation of macrophages: Mechanism and functions. *Immunity*. 2010; 32: 593–604.
2. Davies, L.C.; Jenkins, S.J.; Allen, J.E.; et al.; Tissue-resident macrophages. *Nat. Immunol.* 2013; 14: 986–995.
3. Volkman, A.; & Gowans, J. L.; The origin of macrophages from human bone marrow in the rat. *Br. J. Exp. Pathol.* 1965; 46: 62–70
4. Fidler, I.J.; Macrophage therapy of cancer metastasis. *Ciba Found. Symp.* 1988; 141: 211–222.
5. Biswas, SK.; Allavena, P.; Mantovani, A.; Tumor-associated macrophages: functional diversity, clinical significance, and open questions. *Seminars in immunopathology*. 2013; 35: 585–600.
6. Redente, EF.; Dwyer-Nield, LD.; Merrick ,DT. et al.; Tumor progression stage and anatomical site regulate tumor-associated macrophage and bone marrow-derived monocyte polarization. *Am. J. Pathol.* 2010; 176: 2972–2985.
7. Coussens, LM.; Zitvogel, L.; Palucka, AK.; Neutralizing tumor-promoting chronic inflammation: a magic bullet? *Science*. 2013; 339: 286–291.
8. Murray, P.J.; Wynn, T.A.; Protective and pathogenic functions of macrophage subsets. *Nat. Rev. Immunol.* 2011; 11: 723–737.

9. Bingle, L.; Brown, NJ.; Lewis, CE.; The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. *J Pathol.* 2002; 196: 254-265
10. Lewis, CE.; Pollard, JW.; Distinct role of macrophages in different tumor microenvironments. *Cancer Res.* 2006; 66: 605-612.
11. Qian, BZ.; Pollard, JW.; Macrophage diversity enhances tumor progression and metastasis. *Cell.* 2010; 141:39–51.
12. Gordon, S.; Taylor, PR.; Monocyte and macrophage heterogeneity. *Nat Rev immunol.* 2005; 5: 953–964.
13. Shepard, JL.; Zon, LI.; Developmental derivation of embryonic and adult macrophages. *Curr Opin Hematol.* 2000; 7: 3–8.
14. Passlick, B.; Flieger, D.; Ziegler-Heitbrock, HW.; Identification and characterization of a novel monocyte subpopulation in human peripheral blood. *Blood* . 1989; 74: 2527–2534.
15. Sunderkotter, C.; Nikolic, T.; Dillon, MJ.; et al.; Subpopulations of mouse blood monocytes differ in maturation stage and inflammatory response. *J. Immunol.* 2004; 172 : 4410–4417.
16. Gordon, S.; Martinez, FO.; et al. Alternative activation of macrophages: mechanism and functions. *Immunity.* 2010; 32: 593-604.
17. Mantovani, A.; Sica, A.; et al. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr Opin Immunol.* 2010; 22: 231-237.
18. Kreider, T.; Anthony, RM.; Gause, WC.; et al.; Alternatively activated macrophages in helminth infections. *Curr Opin Immunol.* 2007; 19: 448-453
19. Stout, RD.; Jiang, C.; Matta, B.; et al.; Macrophages sequentially change their functional phenotype in response to changes in microenvironmental influences. *J Immunol.* 2005; 175: 342-349.
20. Mantovani, A.; Allavena, P.; Sica, A.; et al.; Cancer-related inflammation *Nature.* 2008; 454(7203): 436-44.
21. Pollard, JW.; Trophic macrophages in development and disease. *Nat Rev Immunol.* 2009; 9 (4): 259-70.
22. Grebennikov, SI.; Gretel, FR.; Karin, M.; Immunity, inflammation, and cancer. *Cell.* 2010; 140(6): 883-99.
23. Mantovani, A.; Garlanda, C.; Allavena, P.; Molecular pathways and targets in cancer-related inflammation. *Ann Med.* 2010a; 42(3): 161-170.

24. Stout, RD.; Watkins, SK.; Suttles, J.; Functional plasticity of macrophages: in situ reprogramming of tumor-associated macrophages. *J Leukoc Biol.* 2009; 86(5): 1105-1109.
25. Mantovani, A.; Sozzani, S.; Locati, M.; et al.; Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* (2002); 23: 549–555
26. Mantovani, A.; Sica, A.; Allavena, P et al.; Tumor-associated macrophages and the related myeloid-derived suppressor cells as a paradigm of the diversity of macrophage activation. *Human Immunol.* 2009; 70: 325–330
27. Clark, CE.; Hingorani, SR.; Mick, R.; et al.; .Dynamics of the immune reaction to pancreatic cancer from inception to invasion. *Cancer Res.* 2007; 67(19): 9518-27.
28. Coussens, LM. ; Werb, Z. ; et al. ; Inflammation and cancer. *Cell.* 2002 ; 19-26, 420 (6917): 860-7.
29. Hanahan, D. ; Weinberg, RA. ; et al. ; Cancer Hallmarks: The Next Generation. *Cell.* 2011; 144(5): 646-674.
30. Pipeleers, D.; De Baetselier, P.; Van, Ginderachter.; et al.; tumor microenvironments contain functionally distinct subsets of macrophages derived from From Ly6C (high) monocytes. *Cancer Res.* 2010; 70(14): 5728-39.
31. Zhang, J.; Chen, L.; Xiao, M.; et al.; FSP1+ fibroblasts promote skin carcinogenesis by maintaining MCP-1-mediated macrophage infiltration and chronic inflammation. *Am J Pathol.* 2011a; 178(1): 382-90.
32. Roland, CL.; Dineen, SP.; Lynn. KD.; Inhibition of vascular endothelial growth factor reduces angiogenesis and modulates immune cell infiltration of orthotopic breast cancer xenografts. *Mol Cancer Ther.* 2009; 8(7): 1761-71.
33. Duluc, D.; Delneste. Y.; Tan, F.; et al.; Tumor-associated leukemia inhibitory factor and IL-6 skew monocyte differentiation into tumor-associated macrophage-like cells. *Blood.* 2007; 110(13): 4319-30.
34. Song, L.; Asgharzadeh, S.; Salo, J.; et al.; Valpha24-invariant NKT cells mediate antitumor activity via killing of tumor-associated macrophages. *JCI.* 2009; 119(6): 1524-36.
35. Lewis, CE.; Pollard. JW.; et al. Distinct role of macrophages in different tumor microenvironments. *Cancer Res.* 2006; 66(2): 605-12.
36. Heusinkveld, M.; de Vos van Steenwijk, PJ.; Goedemans, R.; et al.; M2 macrophages induced by prostaglandin E2 and IL-6 from cervical carcinoma are switched to activated M1 macrophages by CD4+ Th1 cells. *J Immunol.* 2011; 187(3): 1157-65.

37. Kuang, DM.; Wu, Y.; Chen, N.; et al.; Tumor-derived hyaluronan induces formation of immunosuppressive macrophages through transient early activation of monocytes. *Blood*. 2007; 110(2): 587-95
38. Duluc, D.; Delneste, Y.; Tan, F.; et al.; Tumor-associated leukemia inhibitory factor and IL-6 skew monocyte differentiation into tumor-associated macrophage-like cells. *Blood*. 2007; 110(13): 4319-30.
39. Heusinkveld, M.; Fox Steenwijk, PJ.; van der Burg, SH.; M2 macrophages induced by prostaglandin E2 and IL-6 from cervical carcinoma are switched to M1 macrophages activated by CD4 + Th1 cells. *J Immunol*. 2011; 187(3): 1157-65.
40. Kim, SW.; Kim, JS.; Papadopoulos, J.; et al.; Consistent interactions between tumor cell IL-6 and macrophage TNF- α Enhance the growth of human prostate cancer cells in the bone of nude mouse. *Int Immunopharmacol*. 2011; 11(7): 862-72.
41. Dalton, H.J.; Armaiz-Pena, G.N.; et al .Monocyte subpopulations in angiogenesis. *Cancer Res*. 2014; 74: 1287–1293.
42. Roca, H.; Varsos, Z.S.; Sud, S.; et al.; CCL2 and interleukin-6 promotes survival of human CD11b+ peripheral blood mononuclear cells and induce M2-type macrophage polarization. *J. Biol. Chem*. 2009; 284: 34342–34354.
43. Zhang, J.; Lu, Y.; Pienta, K.J.; Multiple roles of chemokine (C-C motif) ligand 2 in promoting prostate cancer growth. *J. Natl. Cancer Inst*. 2010; 102: 522–528.
44. Qian, B.Z.; Li, J.; Zhang, H.; et al.; CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. *Nature* 2011; 475: 222–225.
45. Mizutani, K.; Sud, S.; McGregor, N.A.; et al.; The chemokine CCL2 increases prostate tumor growth and bone metastasis through macrophage and osteoclast recruitment. *Neoplasia* 2009; 11: 1235–1242.
46. Lin, E.Y.; Nguyen, A.V.; Russell, R.G.; et al.; Colony-stimulating factor 1 promotes progression of mammary tumors to malignancy. *J. Exp. Med*. 2001; 193: 727–740.
47. Kao, J.; Houck, K.; Fan, Y; et al.; Characterization of a novel tumor-derived cytokine. Endothelial-monocyte activating polypeptide II. *J. Biol. Chem*. 1994; 269: 25106–25119.
48. Scholl, S.M.; Pallud, C.; Beuvon, F.; et al.; Anti-colony-stimulating factor-1 antibody staining in primary breast adenocarcinomas correlates with marked inflammatory cell infiltrates and prognosis. *J. Natl.Cancer Inst*. 1994; 86: 120–126.
49. Dorsch, M.; Hock, H.; Kunzendorf, U.; et al.; Macrophage colony-stimulating factor gene transfer into tumor cells induces macrophage infiltration but not tumor suppression. *Eur. J. Immunol*. 1993; 23: 186–190.

50. Balkwill, F.; Cancer and the chemokine network. *Nat. Rev. Cancer* 2004; *4*: 540–550
51. Solinas, G.; Germano, G.; Mantovani, A.; et al.; Tumor-associated macrophages (TAM) as major players of the cancer-related inflammation. *J. Leukoc. Biol.* 2009; *86*: 1065–1073.
52. Day, A.J.; Prestwich, G.D.; Hyaluronan-binding proteins: Tying up the giant. *J. Biol. Chem.* 2002; *277*: 4585–4588.
53. De La Motte, C.A.; Hascall, V.C.; Calabro, A.; et al.; Mononuclear leukocytes preferentially bind via CD44 to hyaluronan on human intestinal mucosal smooth muscle cells after virus infection or treatment with poly(I·C). *J. Biol. Chem.* 1999; *274*: 30747–30755
54. Kobayashi, N.; Miyoshi, S.; Mikami, T; *et al.* Hyaluronan deficiency in tumor stroma impairs macrophage trafficking and tumor neovascularization. *Cancer Res.* 2010; *70*, 7073–7083.
55. Houghton, A.M.; Quintero, P.A.; Perkins, D.L.; et al.; Elastin fragments drive disease progression in a murine model of emphysema. *J. Clin. Invest.* 2006; *116*: 753–759.
56. Solinas, G.; Schiarea, S.; Liguori, M; *et al.* Tumor-conditioned macrophages secrete migration-stimulating factor: A new marker for M2-polarization, influencing tumor cell motility. *J. Immunol.* 2010; *185*: 642–652.
57. Schaefer, L.; Babelova, A.; Kiss, E; *et al.* The matrix component biglycan is pro-inflammatory and signals through Toll-like receptors 4 and 2 in macrophages. *J. Clin. Invest.* 2005; *115*: 2223–2233
58. Scheibner, K.A.; Lutz, M.A.; Boodoo, S.; et al.; Hyaluronan fragments act as an endogenous danger signal by engaging TLR2. *J. Immunol.* 2006; *177*: 1272–1281.
59. Kuang, D.M.; Wu, Y.; Chen, N.; et al.; Tumor-derived hyaluronan induces formation of immunosuppressive macrophages through transient early activation of monocytes. *Blood.* 2007; *110*: 587–595.
60. Murdoch, C.; Giannoudis, A.; Lewis, C.E. Mechanisms regulating the recruitment of macrophages into hypoxic areas of tumors and other ischemic tissues. *Blood.* 2004; *104*: 2224–2234.
61. Lewis, J.S.; Landers, R.J.; Underwood, J.C.; et al.; Expression of vascular endothelial growth factor by macrophages is up-regulated in poorly vascularized areas of breast carcinomas. *J. Pathol.* 2000; *192*: 150–158.

62. Grimshaw, M.J.; Wilson, J.L.; Balkwill, F.R. Endothelin-2 is a macrophage chemoattractant: Implications for macrophage distribution in tumors. *Eur. J. Immunol.* 2002; 32: 2393–2400.
63. Matschurat, S.; Knies, U.E.; Person, V.; Regulation of EMAP II by hypoxia. *Am. J. Pathol.* 2003; 162: 93–103.
64. Korsisaari, N.; Kasman, I.M.; Forrest, W.F. et al. Inhibition of VEGF-A prevents the angiogenic switch and results in increased survival of Apc^{+/min} mice. *Proc. Natl. Acad. Sci. USA* 2007; 104: 10625–10630.
65. Burke, B.; Tang, N.; Corke, K.P.; Expression of HIF-1 α by human macrophages: Implications for the use of macrophages in hypoxia-regulated cancer gene therapy. *J. Pathol.* 2002; 196: 204–212.
66. Talks, K.L.; Turley, H.; Gatter, K.C.; et al. The expression and distribution of the hypoxia-inducible factors HIF-1 α and HIF-2 α in normal human tissues, cancers, and tumor-associated macrophages. *Am. J. Pathol.* 2000; 157: 411–421.
67. Ceradini, D.J.; Kulkarni, A.R.; Callaghan, M.J.; et al.; Progenitor cell trafficking is regulated by hypoxic gradients through HIF-1 induction of SDF-1. *Nat. Med.* 2004; 10, 858–864.
68. Sica, A.; Saccani, A.; Bottazzi, B.; *et al.* Defective expression of the monocyte chemotactic protein-1 receptor CCR2 in macrophages associated with human ovarian carcinoma. *J. Immunol.* 2000; 164: 733–738.
69. Mantovani, A.; Sica, A.; Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr. Opin. Imm* 2010; 22,237:237
70. Moghaddam, S.J., Li, H., Cho, S.N., et al. Promotion of lung carcinogenesis by chronic obstructive pulmonary disease-like airway inflammation in a K-ras-induced mouse model. *Am. J. Respir. Cell Mol. Biol.* 2009; 40: 443–453.
71. Lacy-Hulbert, A., Smith, A.M., et al. Ulcerative colitis and autoimmunity induced by loss of myeloid alpha β integrins. *Proc. Natl. Acad. Sci.* 2007 USA; 104: 15823–15828.
72. Yu, H., Kortylewski, M., and Pardoll, D. Crosstalk between cancer and immune cells: role of STAT3 in the tumour microenvironment. *Nat. Rev. Immunol.* 2007; 7: 41–51.
73. Deng, L., Zhou, J.F., Sellers, R.S., et al.; A novel mouse model on inflammatory bowel disease links mammalian target of rapamycin-dependent hyperproliferation of colonic epithelium to inflammation-associated tumorigenesis. *Am. J. Pathol.* 2007; 176: 952–967.
74. Enzler, T., Gillessen, S., Manis, J.P.; et al.; Deficiencies of GM-CSF and interferon gamma link inflammation and cancer. *J. Exp. Med.* 2003; 197: 1213–1219.

75. Berg, D.J., Davidson, N. et al.; Enterocolitis and colon cancer in interleukin-10-deficient mice are associated with aberrant cytokine production and CD4(+) TH1-like responses. *J. Clin. Invest.* 1996; 98: 1010–1020.
76. Karin, M., Greten, F.R.; NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat. Rev. Immunol.* 2005; 5: 749–759.
77. Greten, F.R., Eckmann, L., Greten, T.F., et al.; IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell* 2004; 118: 285–296.
78. Balkwill, F., Charles, K.A., Mantovani, A.; Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell.* 2005; 7: 211–217.
79. Meira, L.B., Bugni, J.M., et al.; DNA damage induced by chronic inflammation contributes to colon carcinogenesis in mice. *J. Clin. Invest.* 2008; 118: 2516–2525.
80. Pang, B., Zhou, X., Yu, H. et al.; Lipid peroxidation dominates the chemistry of DNA adduct formation in a mouse model of inflammation. *Carcinogenesis* 2007; 28, 1807–1813.
81. Colotta, F., Allavena, P., et al.; Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis* 2009; 30: 1073–1081.
82. Lin, W.W., Karin, M.; A cytokine-mediated link between innate immunity, inflammation, and cancer. *J. Clin. Invest.* 2007; 117: 1175–1183.
83. Naugler, W.E., Sakurai, et al.; Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science.* 2007; 317: 121–124.
84. Karin, M., Lawrence, T., Nizet, V.; Innate immunity gone awry: linking microbial infections to chronic inflammation and cancer. *Cell* 2006; 124: 823–835.
85. Grivennikov, S., Karin, E., et al. IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell* 2009; 15: 103–113.
86. Naugler, W.E., Sakurai, et al. Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science* 2007; 317: 121–124.
87. Pollard, J.W.; Tumour-educated macrophages promote tumour progression and metastasis. *Nat. Rev. Cancer* 2004; 4: 71–78.
88. Pollard, J.W.; Trophic macrophages in development and disease. *Nat. Rev. Immunol.* 2009; 9: 259–270.
89. Saccani, A.; Schioppa, et al.; p50 nuclear factor-kappaB overexpression in tumor-associated macrophages inhibits M1 inflammatory responses and antitumor resistance. *Cancer Res.* 2006; 66: 11432–11440.

90. Sica, A.; Saccani, A.; Bottazzi, B.; et al.; Immuno Autocrine production of IL-10 mediates defective IL-12 production and NF-kappa B activation in tumor-associated macrophages. *J. cancer.aacrjournals* 2000; 164(2): 762-7.
91. Allavena, P.; Mantovani, A.; Immunology in the clinic review series; focus on cancer: tumour-associated macrophages: undisputed stars of the inflammatory tumour microenvironment. *Clin Exp Immunol.* 2012; 167(2): 195-205.
92. Yu, H.; Pardoll, D.; Jove, R.; STATs in cancer inflammation and immunity: a leading role for STAT3. *Nat Rev Cancer.* 2009; 9(11): 798-809.
93. Lewis, CE.; Pollard, JW.; Distinct role of macrophages in different tumor microenvironments. *Cancer Res.* 2006; 66(2): 605-12.
94. Kryczek, I.; Zou, L.; Rodriguez, P.; et al.; B7-H4 expression identifies a novel suppressive macrophage population in human ovarian carcinoma. *J Exp Med.* 2006; 203(4): 871-81.
95. Kuang, DM.; Zhao, Q.; Peng, C.; et al.; Activated monocytes in peritumoral stroma of hepatocellular carcinoma foster immune privilege and disease progression through PD-L1. *J Exp Med.* 2009; 206(6):1327-37.
96. Mantovani, A.; Savino, B.; Locati, M.; et al.; The chemokine system in cancer biology and therapy. *Cytokine Growth Factor Rev.* 2010; 21(1): 27-39.
97. Schutysse, E.; Struyf, S.; Proost, P.; et al.; Identification of biologically active chemokine isoforms from ascitic fluid and elevated levels of CCL18/pulmonary and activation-regulated chemokine in ovarian carcinoma. *J Biol Chem* 2002; 277(27): 24584-93.
98. Liu, J.; Zhang, N.; Li, Q.; Tumor-associated macrophages recruit CCR6+ regulatory T cells and promote the development of colorectal cancer via enhancing CCL20 production in mice. *PLoS One.* 2011; 6(4): e19495.
99. Joyce, JA.; Pollard, JW.; Microenvironmental regulation of metastasis. *Nature reviews. Cancer.* 2009; 9:239–252
100. Qian, B.; Deng, Y.; Im, JH.; et al.; A distinct macrophage population mediates metastatic breast cancer cell extravasation, establishment and growth. *PloS one.* 2009; 4: 6562.
101. Psaila, B.; Lyden, D.; The metastatic niche: adapting the foreign soil. *Nature reviews. Cancer.* 2009; 9:285–293
102. Peinado, H.; Aleckovic, M.; Lavotshkin, S.; et al.; Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. *Nat. Med.* 2012; 18: 883–891.

103. Gil-Bernabe, A.M.; Ferjancic, S.; *et al.*; Recruitment of monocytes/macrophages by tissue factor-mediated coagulation is essential for metastatic cell survival and premetastatic niche establishment in mice. *Blood* 2012; *119*: 3164–3175
104. Erler, J.T.; Bennewith, K.L.; *et al.* Hypoxia-induced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche. *Cancer Cell* 2009; *15*, 35–44.
105. Hiratsuka, S.; Watanabe, A.; Aburatani, H.; *et al.* Tumour-mediated upregulation of chemoattractants and recruitment of myeloid cells predetermines lung metastasis. *Nat. Cell Biol.* 2006; *8*: 1369–1375
106. Hiratsuka, S.; Watanabe, A.; Sakurai *et al.* The S100A8-serum amyloid A3-TLR4 paracrine cascade establishes a pre-metastatic phase. *Nat. Cell Biol.* 2008; *10*: 1349–1355
107. Gazzaniga, S.; Bravo, A.I.; Guglielmotti, A.; Targeting tumor-associated macrophages and inhibition of MCP-1 reduce angiogenesis and tumor growth in a human melanoma xenograft. *J. Invest. Dermatol.* 2007; *127*: 2031–2041.
108. Fischer, C.; Jonckx, B.; Mazzone, M.; *et al.* Anti-PlGF inhibits growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. *Cell* 2007; *131*: 463–475.
109. Zhang, W.; Zhu, X.D.; Sun, H.C.; *et al.* Depletion of tumor-associated macrophages enhances the effect of sorafenib in metastatic liver cancer models by antimetastatic and antiangiogenic effects. *Clin. Cancer Res.* 2010; *16*: 3420–3430.
110. Dineen, S.P.; Lynn, K.D.; *et al.* Vascular endothelial growth factor receptor 2 mediates macrophage infiltration into orthotopic pancreatic tumors in mice. *Cancer Res.* 2008; *68*, 4340–4346.
111. Ries, C.H.; Cannarile, M.A.; Hoves, S.; *et al.*; Targeting tumor-associated macrophages with Anti-CSF-1R antibody reveals a strategy for cancer therapy. *Cancer Cell* 2014; *25*, 846–859.
112. Mok, S.; Koya, R.C.; Tsui, C.; *et al.*; Inhibition of CSF-1 receptor improves the antitumor efficacy of adoptive cell transfer immunotherapy. *Cancer Res.* 2014; *74*, 153–161.
113. Niu, M.; Valdes, S.; Naguib, YW.; *et al.*; Tumor associated macrophage-mediated targeted therapy of triple negative breast cancer. *Mol Pharm* 2016; *13*: 1833–1842.
114. Stock, M.K.; Hammerich, L.; Do, O.N.; *et al.*; HEMet-CCL5 modifies monocyte subpopulations during liver fibrosis regression. *Int J Clin Exp Pathol* 2013; *6*: 678–685.

115. Hiroshima, Y.; Maawy, A.; Hassanein, MK.; et al.; The tumor-educated-macrophage increase of malignancy of human pancreatic cancer is prevented by zoledronic acid PLoS One 2014; 9:e103-382.
116. Joseph, IB.; Isaacs, JT.; et al.; Macrophage role in the anti-prostate cancer response to one class of antiangiogenic agents. J Nat Cancer Inst 1998; 90:1648–1653.
117. Solinas, G.; Germano, G.; Mantovani, A.; et al.; Tumor associated macrophages (TAM) as major players of the cancer related inflammation. J Leukoc Biol 2009; 86:1065–1073.
118. Mullins, DW.; Burger, CJ.; Elgert, KD.; Paclitaxel enhances macrophage IL-12 production in tumor-bearing hosts through nitric oxide. J Immunol 1999; 162: 6811–6818.
119. Haser, sahin.; Marie-luise, B.; Hermann, E.; Therapeutic potential of chemokine receptor antagonists for liver disease. E. review of clinical pharmacology 2010; 503-513
120. Athina Vadalouca, MD.; Efklidis Raptis, MD.; Eleni Moka, MD.; et al.; Pharmacological Treatment of Neuropathic Cancer Pain: A Comprehensive Review of the Current Literature. Pain.P 2012; 12, 219–251.
121. Cao, W.; Peters, JH.; Nieman, D.; et al.; Macrophage subtype predicts lymph node metastasis in oesophageal adenocarcinoma and promotes cancer cell invasion in vitro. Br J Cancer 2015; 113:738–746.
122. Davies, L.C.; Jenkins, S.J.; et al. Tissue-resident macrophages. Nat. Immunol. 2013; 14, 986–995.
123. Bin-Zhi, Qian.; Jeffrey, W.; et al.; Macrophage Diversity Enhances Tumor Progression and Metastasis.cell.2010; 14;39-51