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SUCCESSFUL EVALUATION OF ANTIMICROBIAL ACTIVITY OF GREEN TEA AGAINST VARIOUS PATHOGENIC ORGANISMS

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ABSTRACT

Green tea, a beverage consumed around the world, has progressed from casual beverage to medicinal powerhouse through the centuries. Green tea, made from the leaves of the *Camellia sinensis* plant. In this investigation attempt was made to check antimicrobial activity of green tea against eleven different pathogenic bacteria; *E.coli, S.aureus, Shigella, S.paratyphi A, S.paratyphi B, S.pyogen, S.typhimurium, P.mirabilis, P.vulgaris, C.diptheria, K.pneumoniae* etc. The method used to check antimicrobial activity was Kirby bauer method. Two

types of extracts were used to assessed antimicrobial activity that is Boiled and overnight mixed extract. Green tea is showing good antimicrobial activity against all pathogens.

Index Terms: Antimicrobial activity, Kirby bauer method, Green tea.

I. INTRODUCTION

Tea is a very popular drink world-wide. It is produced from the plant *Camellia sinensis*, which is grown in at least 30 countries, and grows best in certain tropical and subtropical regions Green tea, a beverage consumed around the world, has progressed from casual beverage to medicinal powerhouse through the centuries. There are several different types of tea available on the market, including green, black, white, herbal, and oolong. Green tea, made from the leaves of the Camellia sinensis plant, is unfermented; the freshly plucked tealeaf is steam blasted in perforated drums or cooked in iron pans, denaturing its oxidizing enzymes.

Green tea has been shown to have antimicrobial effects against a variety of gram positive and gram negative bacteria. These antimicrobial effects will be discussed in more detail later in this paper.

II. MATERIAL AND METHODS

A. Sample

Green tea were purchased from local market, Ratnagiri, Maharashtra, India. Unprocessed green tea were selected for study as it may contain natural ingredients.

B. Storage

Sample were stored in dry place and away from sunlight, so as to avoid physicochemical properties of green tea.

C. Test sample preparation

Test samples were prepared for analysis by two methods.1 gm of Green tea were added to 10 ml of sterile saline. This mixture were kept for overnight incubation and second sample were prepared by boiling the same mixture. Both the supernatant were used as sample for further analysis.

D. Cultures used

Total 11 samples were selected for study. Pure cultures were obtained from culture collection laboratory of Biological Sciences Department, Gogate Jogalekar College, Ratangiri, Maharashtra, India. While selecting cultures care was taken that pathogens which causes infection to all physiological tracts of human body were selected. Namely, *E.coli, S.aureus, Shigella, S.paratyphi A, S.paratyphi B, S.pyogen, S.typhimurium, P.mirabilis, P.vulgaris, C.diptheria, K.pneumoniae* were selected.

E. Testing for antimicrobial activity by agar well diffusion method

Petriplates containing 20ml Muller Hinton medium were seeded with 24hr culture of bacterial strains. (*E.coli, S.aureus, Shigella, S.paratyphi A, S.paratyphi B, S.pyogen, S.typhimurium, P.mirabilis, P.vulgaris, C.diptheria, K.pneumoniae*). Wells were cut and 20 µl of the both the samples were added. The plates were then incubated at 37°C for 24 hours. The antibacterial activity was assayed by measuring the diameter of the inhibition zone formed around the well.

F. Minimum Inhibitory Concentration (MIC)

Various concentrations of Green tea samples were Prepared using suitable stock and diluents (as mentioned in the following tables). Then 0.1 ml of respective cultures were added to each tube and all tubes were incubated at appropriate temperature (37°C).

	St. Nutrient Broth containing 20%			
Stock	green tea (both overnight and boiled			
	samples were prepared separately)			
Diluent	St. Nutrient Broth			
Range	2-20% green tea at intervals of 2%			
Total volume	5 ml			
Culture	24 hrs old culture of <i>E.coli</i> , <i>S.aureus</i> ,			
	Shigella, S.paratyphi A, S.paratyphi			
	B, S.pyogen, S.typhimurium,			
	P.mirabilis, P.vulgaris, C.diptheria,			
	K.pneumoniae Optical density			
	adjusted to 0.1 at 530nm)			
Incubation	37°C /24 hrs			
period and time	37 C/24 III8			

After performing MIC, each respective sample were plated to Sterile Nutrient agar plate. And further analysed for growth of organisms. CFU (colony forming Unit) were counted for each concentration.

III. RESULT AND DISCUSSION

A. Results of agar well diffusion method

CULTURES USED	SET 1		SET 2		
	ZONE OF INHIBITION (MM)				
	BOILED	OVERNIGHT	BOILED	OVERNIGHT	
	GREEN TEA	GREEN TEA	GREEN TEA	GREEN TEA	
	(100MG/ML)	(100MG/ML)	(100MG/ML)	(100MG/ML)	
E.coli	23mm	20mm	19mm	20mm	
S.aureus	30mm	-	26mm	-	
Shigella	20mm	15mm	20mm	16mm	
S.paratyphi A	15mm	1	17mm	-	
S.pyogen	18mm	15mm	20mm	18mm	
S.typhimurium	-	20mm	-	21mm	
P.mirabilis	-	•	-	-	
P.vulgaris	26mm	22mm	28mm	19mm	
C.diptheria	24mm	20mm	20mm	17mm	
K.pneumoniae	-	20mm	-	16mm	
S.paratyphi B	-	17mm	-	16mm	

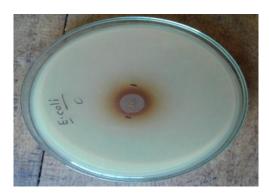


Fig. 1: Petriplate showing zone of inhibition with *E.coli*.



Fig. 2: Petriplate showing zone of inhibition with *S. aureus*.



Fig. 3: Petriplate showing zone of inhibition with Shigella.



Fig. 4: Petriplate showing zone of inhibition with S.paratyphi A.



Fig. 5: Petriplate showing zone of inhibition with *S.pyogen*.



Fig. 6: Petriplate showing zone of inhibition with *S.typhimurium*.



Fig. 7: Petriplate showing zone of inhibition with *P.mirabilis*.



Fig. 8: Petriplate showing zone of inhibition with *P.vulgaris*.



Fig. 9: Petriplate showing zone of inhibition with *C.diptheria*.



Fig. 10: Petriplate showing zone of inhibition with K.pneumoniae.



Fig. 11: Petriplate showing zone of inhibition with S.paratyphi B.

IV. DISCUSSION

In conclusion, this experiment has shown that green tea in general shows antimicrobial properties. Almost all of the organisms tested showing susceptibility for green tea. All these bacterial species selected comprises all physiological tracts as per the infections are concerned. Hence, we can make conclusion that green tea is effective for overall health of an individual.

V. REFERENCES

- M. Radji, R. A. Agustama, B. Elya, And C. R.Tjampakasari, "Antimicrobial Activity of Green Tea Extract Against Isolates Of Methicillin-Resistant Staphylococcus aureus And Multi-Drug Resistant Pseudomonas aeruginosa," Asian Pacific Journal of Tropical Biomedicine, 2013; 3(8): 663-667.
- 2. Peter W. Taylor, Jeremy M.T., Hamilton-Miller, And Paul D. Stapleton, "Antimicrobial Properties Of Green Tea Catechins," Europe Pmc Funders Group Food Sci Technol Bull. 2009 October 19. 2005; 2: 71–81.
- 3. S. Archana And J. Abraham, "Comparative Analysis of Antimicrobial Activity of Leaf Extracts From Fresh Green Tea, Commercial Green Tea And Black Tea On Pathogens," *Journal Of Applied Pharmaceutical Science*, 2011; 01(08): 149-152.
- 4. M. Rafael, A. bernaelya, c. Rianatjampakasari, "Antimicrobial Activity of Green Tea Extract Against Isolates of Methicillin–Resistant Staphylococcus aureus And Multi–Drug Resistant Pseudomonas aeruginosa,", Asian Pacific Journal of Tropical Biomedicine, August 2013; 3(8): 663-667.
- 5. J. Gopal, M. Muthu,, D. Paul, D. Hwan Kim & S. Chun "Bactericidal activity of green tea extracts: the importance of catechin containing nano particles," Scientific Reports |6:19710, DOI: 10.1038/srep19710