

**ANTICANCER ACTIVITY AND ANTI DIABETIC ACTIVITY OF
STREPTOMYCES SPECIES JKCM1*****M. Guravaiah**

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Article Received on
20 Feb. 2018,

Revised on 11 March 2018,
Accepted on 01 April 2018,

DOI: 10.20959/wjpr20187-12640

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ABSTRACT

The Actinomycetales, especially the genus *Streptomyces*, produce a number of antibiotics and other bioactive natural products. Measurement of cell viability and proliferation forms the basis for numerous in vitro assays of a cell population's response to external factors. MTT Assay is a colorimetric assay that measures the reduction of yellow 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) by mitochondrial succinate dehydrogenase. HepG2, Human Liver cancer cells were cultured in DMEM containing 4.5 g/L D-glucose with 10% heat-inactivated FBS at 37°C, 5% CO₂ atmosphere.

KEYWORDS: Actinomycetales, cell viability, colorimetric assay and MTT Assay.

INTRODUCTION

The members of the order Actinomycetales, especially the genus *Streptomyces*, produce a number of antibiotics and other bioactive natural products. The genomic analysis of some *Streptomyces* strains revealed the presence of biosynthetic gene clusters for about 30 secondary metabolites (Bentley et al., 2002; Ikeda et al., 2003; Ohnishi et al., 2008).

Metabolic disorders (i.e., any of the diseases or disorders that disrupt normal metabolism) are common pathologies, and especially diabete (Kaur J.,2014). In 2013, it was estimated that over 382 million people throughout the world have diabetes and this number is expected to increase up to 500 million in 2030 (Barde S.R et al.,2015) when it is expected that this disease will be the 7th leading cause of death (Mathers C.D et al., 2006).

ANTI CANCER ACTIVITY

Measurement of cell viability and proliferation forms the basis for numerous in vitro assays of a cell population's response to external factors. The MTT Cell Proliferation Assay measures the cell proliferation rate and conversely, when metabolic events lead to apoptosis or necrosis, the reduction in cell viability.

MATERIALS AND METHODS

DMEM (Dulbecco's modified Eagles medium), MTT[3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide], trypsin, EDTA Phosphate Buffered Saline (PBS) and were purchased from Sigma Chemicals Co. (St. Louis, MO) and Fetal Bovine Serum (FBS) were purchased from Gibco. 25 cm² and 75 cm² flask and 96 well plated purchased from eppendorf India.

Maintenance of Cell Line

The MCF-7 breast adenocarcinoma cancer cell line were purchased from NCCS, Pune and the cells were maintained in MEM supplemented with 10% FBS and the antibiotics penicillin/streptomycin (0.5 mL⁻¹), in atmosphere of 5% CO₂/95% air at 37 °C.

Preparation of Test Compound

For MTT assay, Each Test compounds were weighed separately and dissolved in DMSO. With media make up the final concentration to 1 mg/ ml and the cells were treated with series of concentrations from 10 to 100 µg/ ml.

MCF-7 CELL VIABILITY BY MTT ASSAY

Principle

MTT Assay is a colorimetric assay that measures the reduction of yellow 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) by mitochondrial succinate dehydrogenase. The assay depends both on the number of cells present and on the assumption that dead cells or their products do not reduce tetrazolium. The MTT enters the cells and passes into the mitochondria where it is reduced to an insoluble, dark purple coloured formazan crystals. The cells are then solubilized with a DMSO and the released, solubilized formazan reagent is measured spectrophotometrically at 570 nm.

Procedure

Cell viability was evaluated by the MTT Assay with three independent experiments with six concentrations of compounds in triplicates. MCF- 7 cells were trypsinized and perform the thetryphan blue assay to know viable cells in cell suspension. Cells were counted by haemocytometer and seeded at density of 5.0×10^3 cells / well in 100 μ l media in 96 well plate culture medium and incubated overnight at 37°C. After incubation, take off the old media and add fresh media 100 μ l with different concentrations of test compound in represntive wells in 96 plates. After 48 hrs., Discard the drug solution and add the fresh medic with MTT solution ($0.5 \text{ mg} / \text{mL}^{-1}$) was added to each well and plates were incubated at 37°C for 3 hrs. At the end of incubation time, precipitates are formed as a result of the reduction of the MTT salt to chromophore formazan crystals by the cells with metabolically active mitochondria. The optical density of solubilized crystals in DMSO was measured at 570 nm on a microplate reader. The percentage growth inhibition was calculated using the following formula and concentration of test drug needed to inhibit cell growth by 50% values is generated from the dose-response curves for each cell line using with origin software.

$$\% \text{ Inhibition} = \frac{100 (\text{Control} - \text{Treatment})}{\text{Control}}$$

1. Hela Cell line: *Streptomyces species* JKCM1.

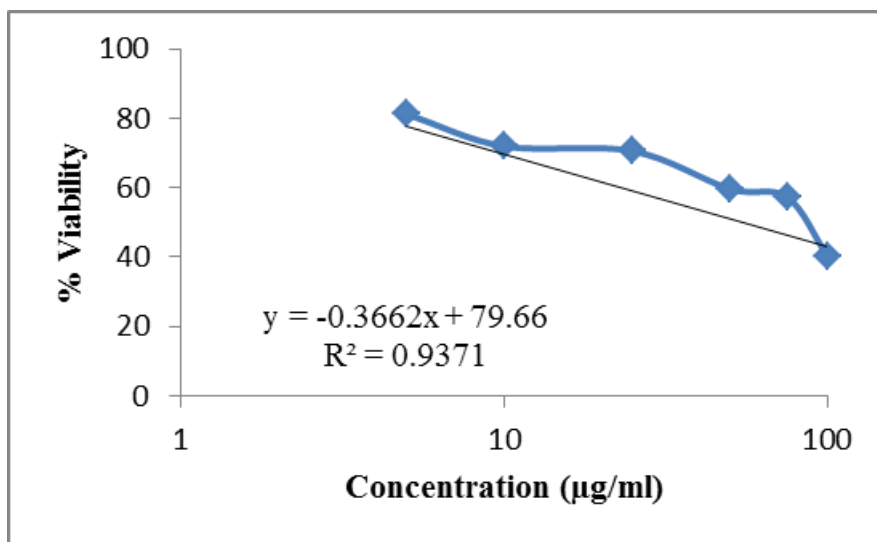
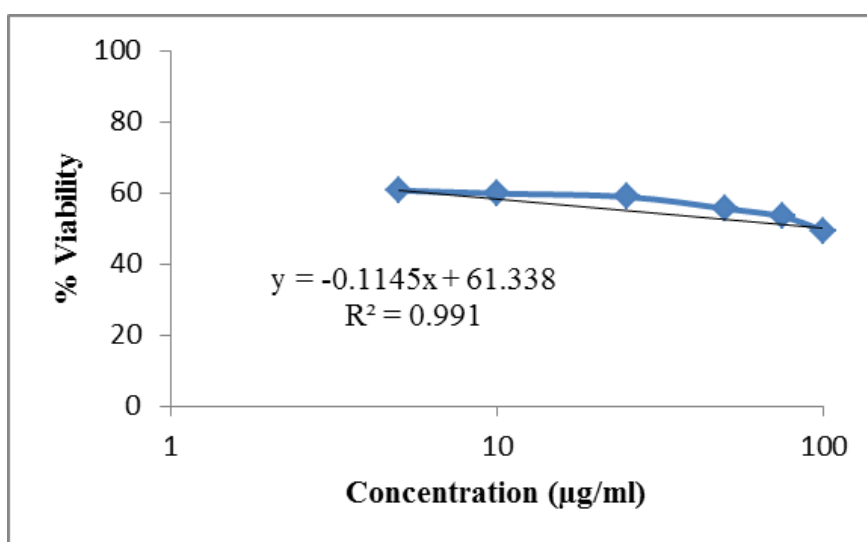


Figure 1: Cytotoxic effect of the *Streptomyces species* JKCM1 Hela Cell Line.

Table 1: Cytotoxic Properties of Streptomyces species JKCM1 on Hela Cell Line.

Concentration($\mu\text{g/ml}$)	Absorbance at 570nm			Average	Average-Blank	% Viability	IC ₅₀ ($\mu\text{g/ml}$)
100	0.841	0.843	0.845	0.843	0.84	40.287	
75	1.196	1.198	1.201	1.198	1.195	57.314	
50	1.244	1.246	1.248	1.246	1.243	59.616	
25	1.472	1.474	1.478	1.474	1.471	70.551	
10	1.501	1.503	1.505	1.503	1.5	71.942	
5	1.693	1.696	1.699	1.696	1.693	81.199	
Untreated	2.088	2.089	2.088	2.088	2.085	100	
Blank	0.003	0.004	0.003	0.003	0	0	

2. HT - 29 Cell line: Streptomyces species JKCM1.**Figure 2: Cytotoxic effect of the Streptomyces species JKCM1 HT29 Cell Line.****Table 2: Cytotoxic Properties of Streptomyces species JKCM1 on HT29 Cell Line.**

Concentration($\mu\text{g/ml}$)	Absorbance at 570nm			Average	Average-Blank	% Viability	IC ₅₀ ($\mu\text{g/ml}$)
100	0.998	1.01	1.03	1.012	1.008	49.411	
75	1.09	1.092	1.094	1.092	1.088	53.333	
50	1.135	1.139	1.14	1.138	1.134	55.588	
25	1.204	1.206	1.207	1.205	1.201	58.872	
10	1.223	1.225	1.227	1.225	1.221	59.852	
5	1.239	1.241	1.243	1.241	1.237	60.637	
Untreated	2.044	2.043	2.045	2.044	2.04	100	
Blank	0.004	0.005	0.004	0.004	0		

ANTI DIABETIC ACTIVITY

HepG2, Human Liver cancer cells were cultured in DMEM containing 4.5 g/L D-glucose with 10% heat-inactivated FBS at 37°C, 5% CO₂ atmosphere. The cells were seeded into 96-well plate with six wells left as blank wells and let growing to confluence; then cells were

fully differentiated in DMEM with 2% FBS for 5 days. Before tests, the medium was replaced by RPMI1640 (2 g/L glucose) supplemented with 0.2% BSA. The medium was removed after 2 h, and the same medium containing *Streptomyces species* JKCM1 (0.5, 5 and 10 µg/mL), GA (0.05, 0.5 and 5 µM), metformin (0.01 mM), and DMSO in absence or presence of insulin (1 µmol/L) was added to all wells including the blank. The glucose in the medium was determined by the glucose-oxidase method after 48 h treatment. The amount of glucose uptake by muscle cells was calculated by using the following formula:

Glucose uptake = [Glucose concentration of blank wells] [Glucose concentration of cell plated wells]

RESULTS

Table: Glucose uptake in HepG2 cells after 48 h incubation in media with glucose (2 g/L).

Treatment	Concentration	Glucose consumption (mg/100 mL)	
		Absence of insulin	Presence of insulin (1µmol/L)
Vehicle control	0.1% DMSO	2.95 ± 0.07	7.17 ± 0.03
Metformin	0.01 mM	12.80 ± 0.06	14.96 ± 0.03
	0.1 mM	10.84 ± 0.09	12.68 ± 0.13
<i>Streptomyces species</i> JKCM1	0.5 µg/mL	20.77 ± 0.07	28.69 ± 0.06
	5 µg/mL	21.05 ± 0.08	29.79 ± 0.05
	10 µg/mL	22.98 ± 0.07	30.96 ± 0.04
Gallic acid	0.05 µM	18.04 ± 0.05	22.96 ± 0.09
	0.5 µM	15.96 ± 0.07	21.87 ± 0.10
	5 µM	16.58 ± 0.14	20.83 ± 0.09

CONCLUSION

The anticancer drugs are available for the treatment of cancer but new anticancer drugs are urgently needed due to harmful adverse effects of the existing drugs. In our study we reported that our crude metabolite obtained from co-culture of two marine *Streptomyces species* JKCM1 has excellent anticancer activity and anti Diabetic Activity. This extract might be used in infectious and cancer diseases and further research will be done for the isolation of pure compounds.

REFERENCE

1. Bentley SD, Chater KF, Cerdano-Tarraga AM, Challis GL, Thomson NR, James KD, Harris DE, Quil MA, Kieser H, Harper D, Bateman A, Brown S. Complete genome

- sequence of the model actinomycete *Streptomyces coelicolor* A3(2). *Nature*, 2002; 417: 141-147.
2. Ikeda H, Ishikawa J, Hanamoto A, Shinose M, Kikuchi H, Shiba T, Sakaki Y, Hattori M, Omura S. Complete genome sequence and comparative analysis of the industrial microorganism *Streptomyces avermitilis*. *Nat Biotechnol*, 2003; 21: 526-531.
 3. Ohnishi Y, Ishikawa J, Hara H, Suzuki H, Ikenoya M, Ikeda H, Yamashita A, Hattori M, Horinouchi S. Genome sequence of the streptomycin-producing microorganism *Streptomyces griseus* IFO 13350. *J Bacteriol*, 2008; 190: 4050-4060.
 4. Kaur J. A comprehensive review on metabolic syndrome. *Cardiol. Res. Pract.*, 2014; 2014: 943162. doi: 10.1155/2014/943162.
 5. Barde S.R., Sakhare R.S., Kanthale S.B., Chandak P.G., Jamkhande P.G. Marine bioactive agents: A short review on new marine antidiabetic compounds. *Asian Pac. J. Trop. Dis.*, 2015; 5: S209–S213. doi: 10.1016/S2222-1808(15)60891-X.
 6. Mathers C.D., Loncar D. Projections of global mortality and burden of disease from 2002 to 2030. *PLoS Med.*, 2006; 3: e442. doi: 10.1371/journal.pmed.0030442.