

## FORMULATION AND IN VITRO EVALUATION OF MUCO ADHESION AND FLOATING MICROSPHERES OF ETEODOLAC USING IONIC GELATION METHOD

K. Divya Laxmi\*, S. Vineela Sangu, M. A. Rahman and Mohammad Asif

Gyana Jyothi College of Pharmacy, Uppal Bus Depot, Hyderabad-500089, Telangana, India.

Article Received on  
02 Jan. 2019,

Revised on 24 Jan. 2019,  
Accepted on 15 Feb. 2019

DOI: 10.20959/wjpr20193-14339

### \*Corresponding Author

**Dr. K. Divya Laxmi**

Gyana Jyothi College of  
Pharmacy, Uppal Bus Depot,  
Hyderabad-500089,  
Telangana, India.

### ABSTRACT

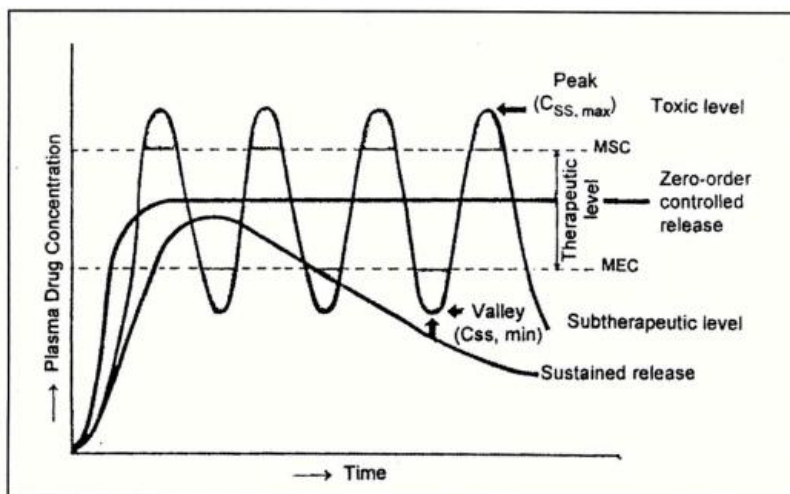
The purpose of this study is to formulation and evaluation of floating and mucoadhesion microsphere of etodolac using ionic gelatin method. The floating and mucoadhesion microspheres were studied for micromeritic properties such as Bulk density, Tapped density, Carrs index, Hausner's ratio, Angle of repose which were found to be within limits. The percentage yield of floating microsphere formulation f1 to f6 and mucoadhesive microspheres m1 to m3 were in range of  $77.14 \pm 0.64$  to  $92.74 \pm 0.74\%$ . The *in vitro* buoyancy of formulation f1 to f6, it was range from  $71.96 \pm 1.04$  to  $82.96 \pm 1.07$ . Among all formulation f6 was found to be highest *in-vitro* buoyancy  $82.96 \pm 1.07$ .

The results also showed that the larger the particle size, the longer the floating time. The entrapment efficiency of floating microspheres f1 to f6 and mucoadhesive microspheres were in the range of  $77.43 \pm 2.72$  to  $98.11 \pm 2.59$ . From the dissolution data of floating microspheres by ionic gelation method. Formulations prepared with sodium alginate alone has shown maximum drug release at 12 hr in the ratio of 1:3. Formulations prepared with sodium alginate along with HPMC K 4M retards the drug release. Among all formulations of floating microspheres F3 was considered as optimised for floating microspheres. From the release kinetics data, it was evident that floating optimised formulation follows zero order release kinetics. From the dissolution data of mucoadhesive microspheres by ionic gelation method m1 formulation has shown maximum drug release at 12 hrs. When increase in the polymer concentration retards the drug release more than 12 hrs. Hence m1 was considered as optimised formulation for mucoadhesive microspheres. From the release kinetics data, it was evident that mucoadhesive optimised formula was followed zero order release kinetics.

**KEYWORDS:** Etodolac, ionic gelation method, floating and mucoadhesion microspheres.

## 1. INTRODUCTION

The oral route of drug administration is the most important method of administering drugs for systemic effects. The parenteral route is not routinely used or not possible to self-administration of medication. The topical route of administration has only recently been employed to deliver drugs to the body for systemic effects.

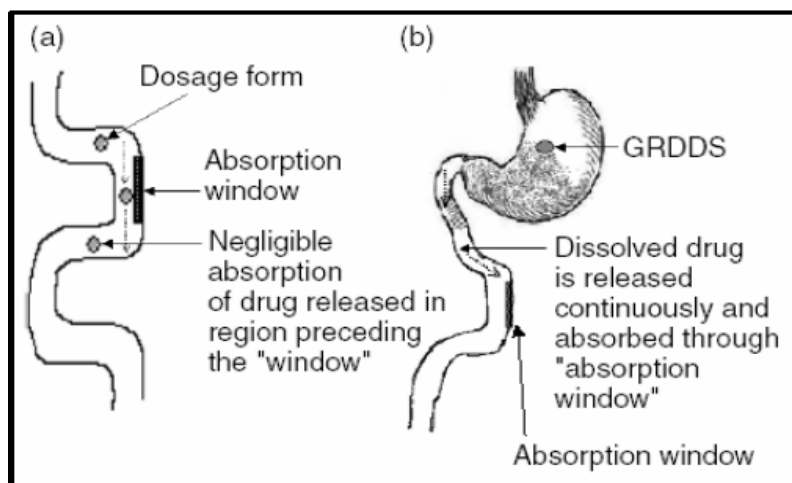


Plasma level profiles following conventional and controlled release dosing controlled release dosage forms suffer from mainly two adversities: The short gastric retention time (GRT) and unpredictable gastric emptying time (GET).<sup>[8,9]</sup> Gastric emptying of dosage forms is an extremely variable process and ability to prolong and control the emptying time is valuable asset for dosage forms, which resides in the stomach for a longer period of time than conventional dosage forms.<sup>[10]</sup>

### Gastroretentive drug delivery system (GRDDS)

Recent scientific and patent literature shows increased interest in academics and industrial research groups regarding the novel dosage forms that can be retained in the stomach for a prolonged and predictable period of time. One of the most feasible approaches for achieving a prolonged and predictable drug delivery profile in the GI tract is to control the gastric residence time (GRT), using gastroretentive drug delivery system (GRDDS) that will provide us with new and important therapeutic options<sup>8</sup>. Gastroretentive system can remain in the gastric region for several hours and hence significantly prolong the gastric residence time of drugs. Prolonged gastric retention improves bioavailability, reduces drug waste, and improves solubility of drugs that are less soluble in a high pH environment. It has

applications also for local drug delivery to the stomach and proximal small intestines. Gastro retention helps to provide better availability of new products with new therapeutic possibilities and substantial benefits for patients.



(a) Conventional drug delivery system (b) GRDDS

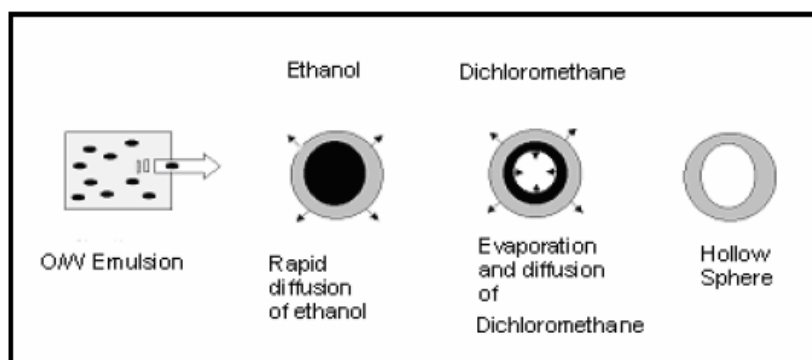
### Classification of FDDS

Based on the mechanism of buoyancy, floating systems can be classified into two distinct categories *viz.* Non-effervescent and effervescent systems.

#### A. Non-Effervescent systems

1. Colloidal gel barrier systems:
2. Micro- porous compartment system:
3. Alginate beads.
4. Hollow Microspheres.

Hollow microspheres (microballoons) were prepared by novel emulsion solvent diffusion method. The ethanol: dichloromethane solution of the drug and an enteric acrylic polymer were poured into an agitated aqueous solution of PVA that was thermally controlled at 40°C. The gas phase was generated in dispersed polymer droplet by evaporation of dichloromethane and formed an internal cavity in microsphere of polymer with drug (Fig1.10).



**Mechanism of microballoon formation by emulsion-solvent diffusion method.**

## B. Effervescent systems

1. Volatile liquid containing systems
2. Gas generating systems.

## 2. METHODOLOGY

### Material and sources

S. No	List of Chemicals	Manufacturing Company
1	Etodolac	Sura Labs
2	Sodium alginate	Merk specialities Pvt Limited, Mumbai
3	Carbopol 934	Merk specialities Pvt Limited, Mumbai
4	Calcium chloride	Merk specialities Pvt Limited, Mumbai
5	Hpmc k 4m	Merk specialities Pvt Limited, Mumbai
6	Sodium bicarbonate	Merk specialities Pvt Limited, Mumbai

### Formulation of floating Microspheres

Formulation code	Drug (mg)	Sodium alginate (mg)	HPMC K 4M	Sodium Bicarbonate (mg)
F1	1000	1000	-	200
F2	1000	2000	-	200
F3	1000	3000	-	200
F4	1000	750	250	200
F5	1000	1500	500	200
F6	1000	2250	750	200

### Formulation of Mucoadhesive Microspheres

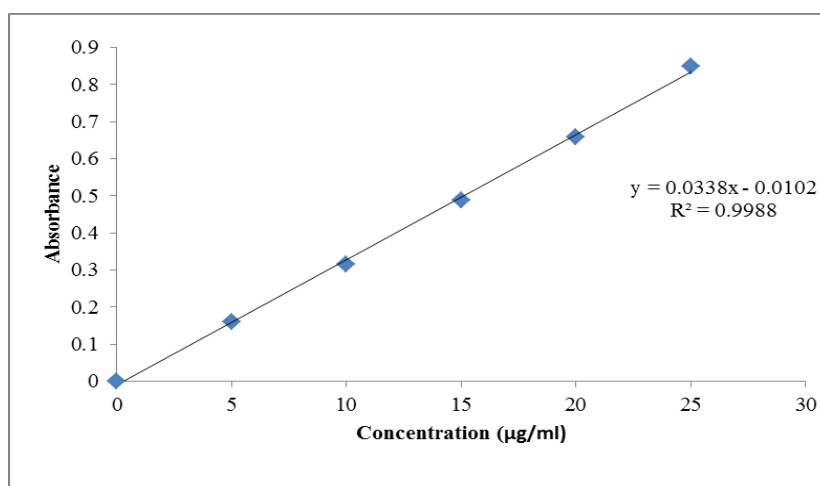
S. No	Formulation code	Drug (mg)	Sodium alginate (mg)	Carbopol 934 (mg)
1	M1	1000	500	500
2	M2	1000	1000	1000
3	M3	1000	1500	1500

### 3. RESULTS AND DISCUSSION

#### Standard calibration curve of Etodolac

#### Standard calibration data of Etodolac in 0.1N HCL at 225nm

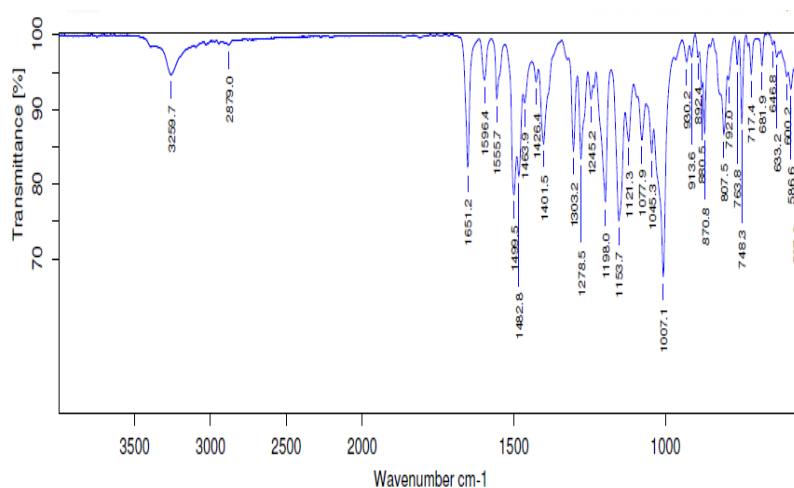
Concentration (µg/ml)	Absorbance
0	0
5	0.161
10	0.316
15	0.488
20	0.658
25	0.850



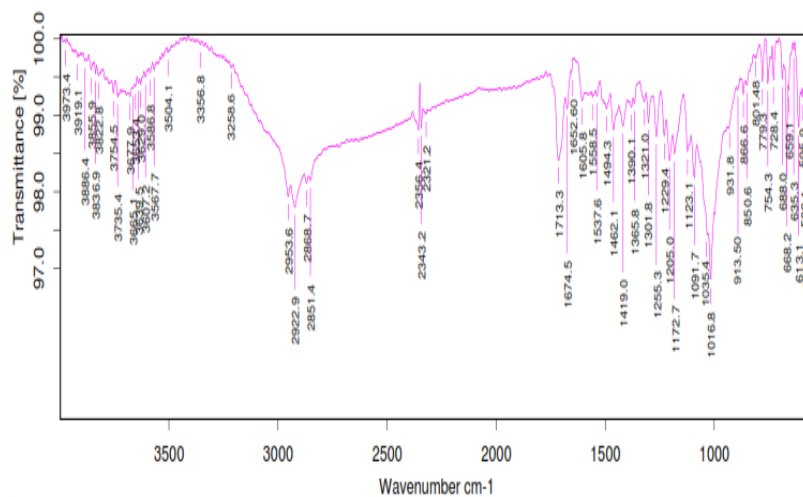
Standard calibration curve of Etodolac in 0.1N HCL

#### Preformulation studies

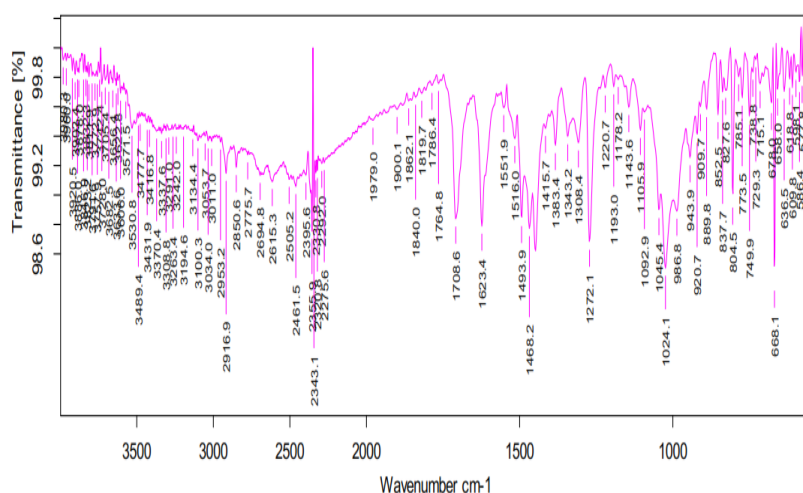
#### Drug- Excipient Compatability studies



FTIR of Etodolac pure drug

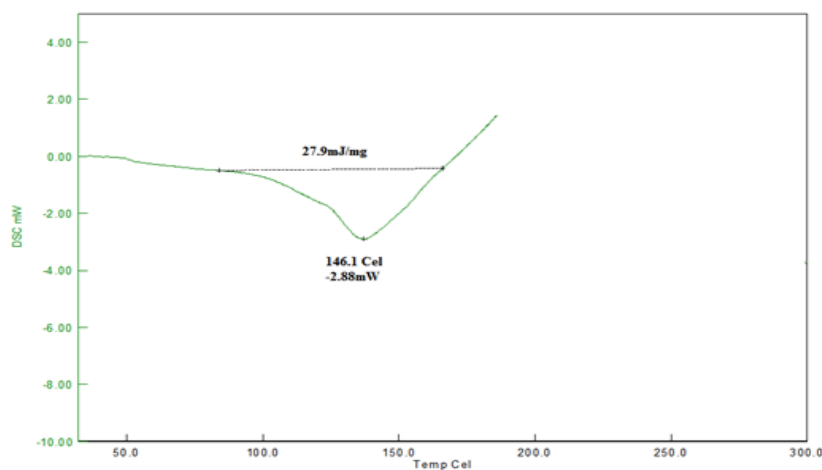


FTIR of Optimized Floating Microspheres F3 formula

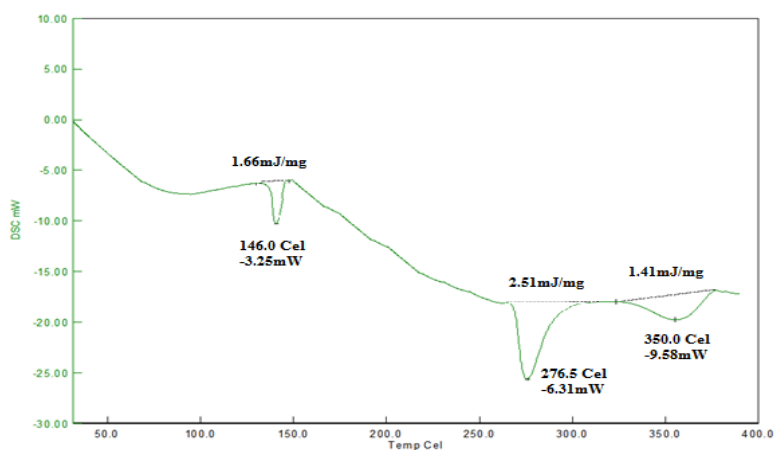


FTIR of Optimized Mucoadhesive Microspheres M1 formula

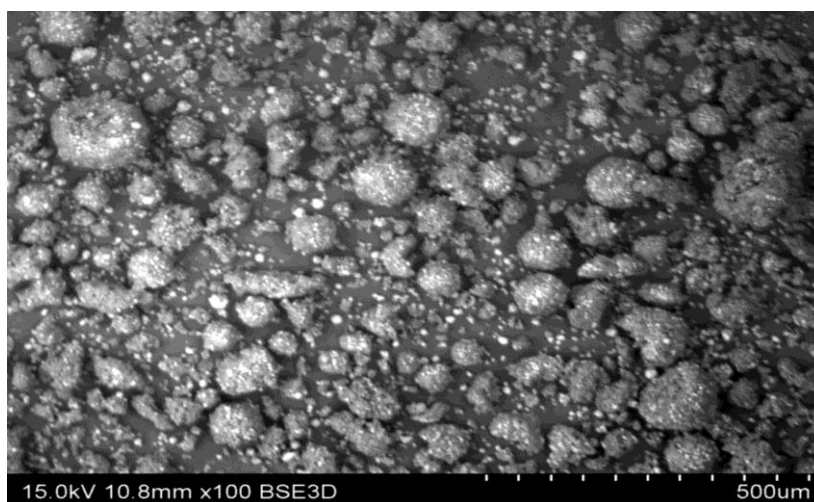
## DSC Studies



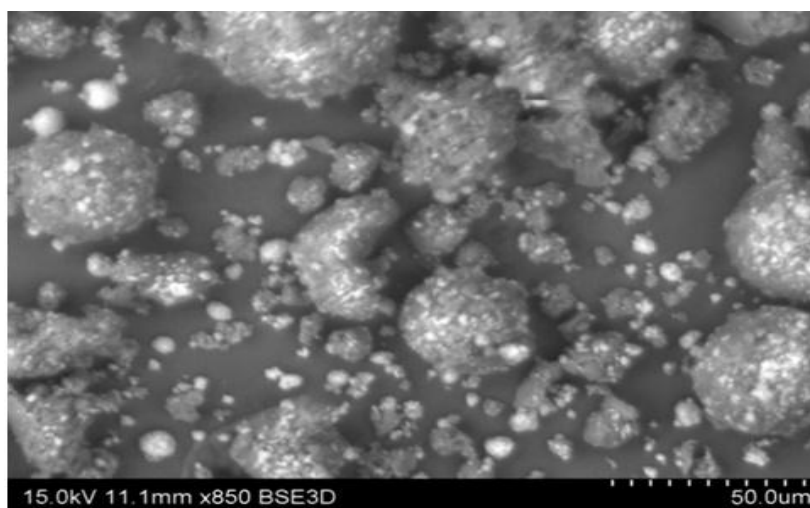
DSC of Etodolac pure drug



DSC of Etodolac + Sodium alginate + HPMC K4M.

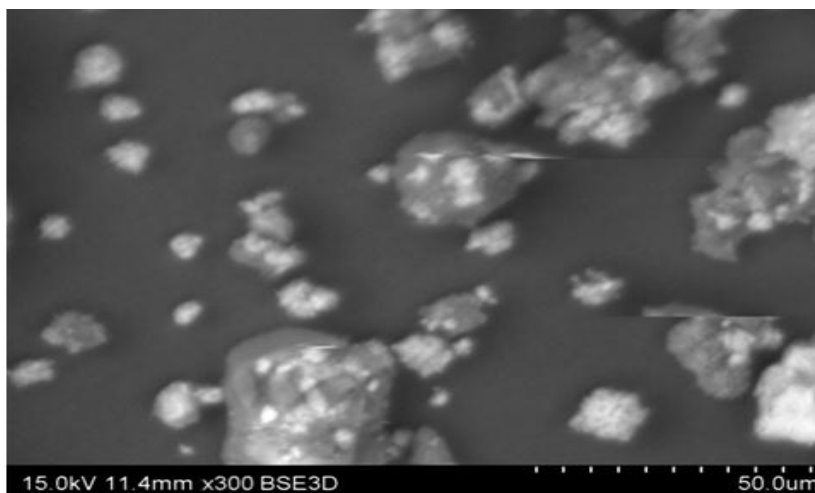


SEM of Etodolac pure drug



SEM of Etodolac Floating Microspheres optimised F3 formulation





SEM of Etodolac Mucoadhesive Microspheres M1 optimised formulation

## Evaluation of Microspheres

## Micrometric Properties

S. No	Formulation code	Mean partical size (μm)	Bulk density (gm. /cm <sup>3</sup> )	Tapped density (gm. /cm <sup>3</sup> )	Hausners ratio	Carr's Index	Angle of Repose(θ)
1	F1	381.55±2.54	0.32±0.010	0.39±0.018	1.21±0.04	11.13±0.11	28.49±1.71
2	F2	455.22±2.52	0.35±0.012	0.40±0.015	1.14±0.05	12.5±0.64	27.72±1.89
3	F3	471.52±2.05	0.40±0.007	0.47±0.014	1.17±0.03	14.8±0.24	30.88±2.78
4	F4	385.15±1.08	0.36±0.014	0.44±0.014	1.22±0.01	18.18±0.33	27.00±1.93
5	F5	451.84±2.07	0.41±0.015	0.47±0.015	1.14±0.02	12.76±0.26	26.02±1.80
6	F6	493.24±2.43	0.40±0.012	0.48±0.021	1.2±0.01	16.66±0.33	26.56±1.43
7	M1	473.9±2.16	0.39±0.018	0.45±0.022	1.15±0.03	13.33±1.5	26.80±1.68
8	M2	482.12±2.21	0.41±0.015	0.48±0.027	1.17±0.01	14.5±0.86	27.11±1.59
9	M3	477.5±2.15	0.44±0.017	0.50±0.015	1.13±0.02	12±0.35	26.56±1.68

All values represented as mean ± standard deviation (n=3)

Percentage yield, *in vitro* buoyancy and incorporation efficiency of floating microspheres of Etodolac

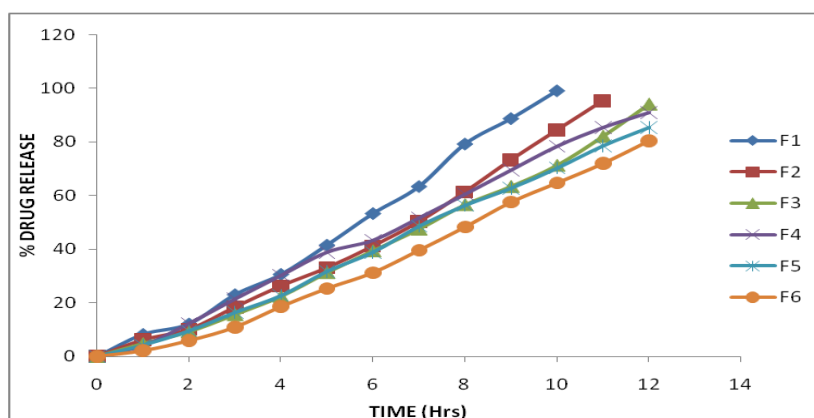
S. No	Formulation code	Percentage yield	<i>In vitro</i> buoyancy (in sec)	Entrapment Efficiency (%)
1	F1	77.14±0.64	71.96±1.04	77.43±2.72
2	F2	82.29±0.69	75.43±2.02	87.34±2.84
3	F3	85.35±0.66	79.32±0.97	87.11±3.01
4	F4	93.08±0.72	73.41±1.03	92.30±2.88
5	F5	80.48±0.65	75.33±1.32	79.76±1.58
6	F6	86.05±0.51	82.96±1.07	91.94±2.17
7	M1	90.17±0.43	-	98.11±2.59
8	M2	92.74±0.74	-	83.91±2.02
9	M3	90.64±0.55	-	90.38±2.34

All values represented as mean ± standard deviation (n=3)



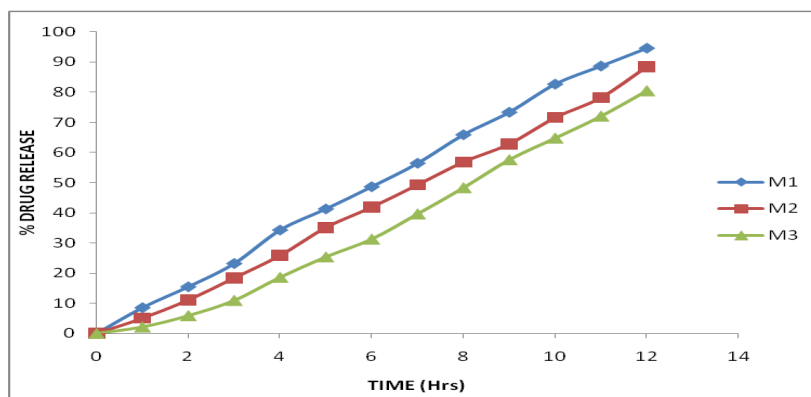
**In Vitro drug release*****In-Vitro* drug release data of Etodolac floating microspheres F1 to F6**

S. No	Cumulative %drug release						
	Time (Hrs)	F1	F2	F3	F4	F5	F6
	0	0	0	0	0	0	0
1	1	8.24 ± 0.98	6.22 ± 1.05	4.83 ± 1.15	4.07 ± 1.28	4.23 ± 1.11	2.16±1.24
2	2	12.14 ± 1.25	10.11 ± 1.12	9.23 ± 2.24	12.32 ± 0.98	9.56 ± 1.64	5.91±1.52
3	3	23.08 ± 2.05	18.42 ± 1.85	15.65 ± 1.08	21.44 ± 2.01	16.43 ± 1.54	10.96±1.47
4	4	30.64 ± 1.56	26.32 ± 2.04	22.42 ± 0.98	30.23 ± 1.41	22.71 ± 2.12	18.65±0.97
5	5	41.55 ± 1.81	33.08 ± 2.17	31.32 ± 1.64	38.86 ± 1.06	31.78 ± 0.95	25.41±2.17
6	6	53.34 ± 2.14	41.15 ± 1.53	39.44 ± 1.55	43.29 ± 1.75	38.92 ± 1.04	31.32±1.61
7	7	63.41 ± 1.74	50.28 ± 1.67	47.54 ± 1.34	51.65 ± 2.11	48.64 ± 2.06	39.68±2.05
8	8	79.27 ± 2.05	61.33 ± 1.74	56.63 ± 1.27	60.46 ± 1.62	56.38 ± 1.26	48.36±1.04
9	9	88.75 ± 1.34	73.28 ± 1.97	63.43 ± 1.31	69.45 ± 1.47	62.81 ± 1.40	57.65±1.67
10	10	99.12 ± 2.08	84.36 ± 2.17	71.32 ± 1.55	78.34 ± 1.09	70.30 ± 1.55	64.84±2.21
11	11		95.34 ± 2.08	82.14 ± 2.43	85.34 ± 1.14	78.64 ± 1.07	72.11±1.33
12	12			94.14 ± 2.11	90.91 ± 2.07	85.48 ± 2.17	80.55±1.41

***In-Vitro* drug release profile of Etodolac Floating microspheres*****In-Vitro* drug release data of Etodolac Mucoadhesive microspheres**

S. No	Time (hr)	Cumulative % Drug Release		
		M1	M2	M3
1	0	0	0	0
2	1	8.56 ± 1.67	5.07± 1.26	2.16±1.24
3	2	15.48 ± 2.15	11.12± 0.98	5.91±1.52
4	3	23.18 ± 1.06	18.41± 1.41	10.96±1.47
5	4	34.29 ± 0.96	25.87± 0.86	18.65±0.97
6	5	41.28 ± 2.04	35.14± 0.78	25.41±2.17
7	6	48.65 ± 1.62	41.88± 1.07	31.32±1.61
8	7	56.43 ± 1.34	49.32± 1.12	39.68±2.05

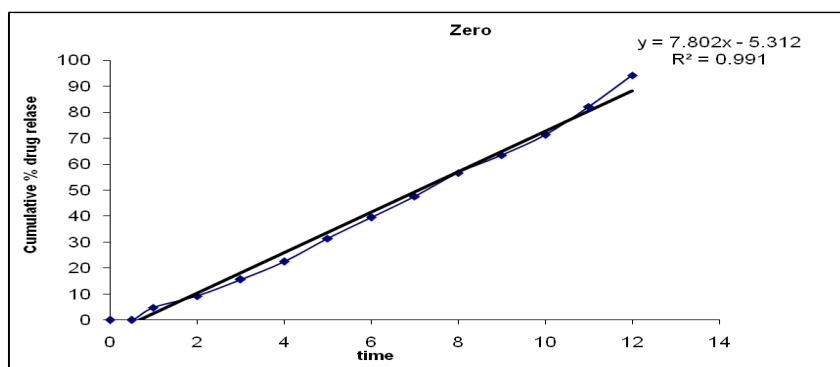
9	8	$65.87 \pm 2.11$	$56.87 \pm 1.16$	$48.36 \pm 1.04$
10	9	$73.34 \pm 1.47$	$62.87 \pm 1.96$	$57.65 \pm 1.67$
11	10	$82.65 \pm 1.32$	$71.59 \pm 0.84$	$64.84 \pm 2.21$
12	11	$88.65 \pm 2.06$	$78.23 \pm 0.39$	$72.11 \pm 1.33$
13	12	$94.55 \pm 0.92$	$88.49 \pm 1.64$	$80.55 \pm 1.41$



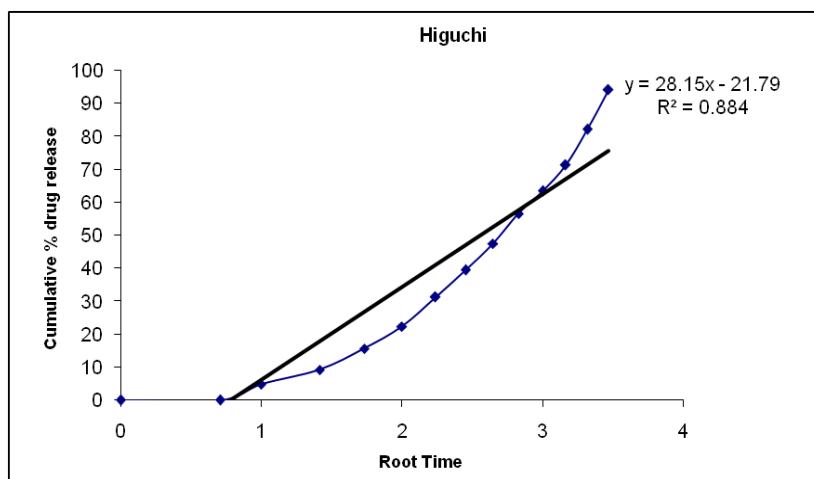
Comparison of *In-Vitro* drug release profile of Etodolac mucoadhesive microspheres

#### Data of Release Kinetics

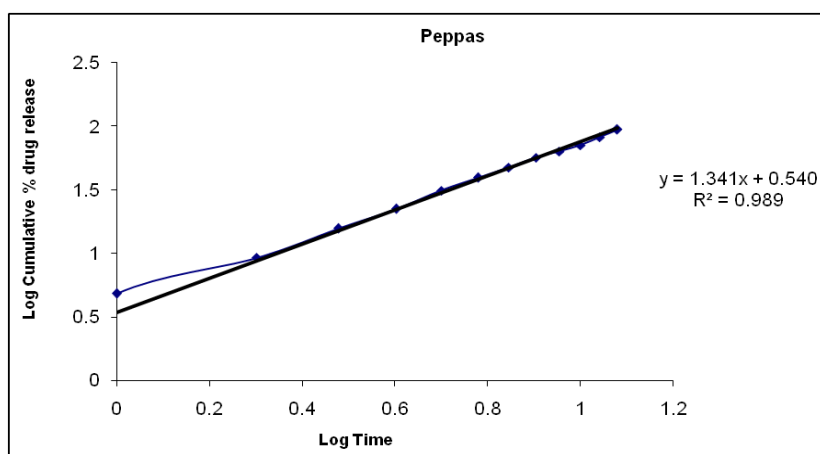
S. No	Cumulative (%) Release Q	Time (T)	Root (T)	Log (%) Release	Log (T)	Log (%) Remain
1	0	0	0			2.000
2	0	0.5	0.707	#NUM!	-0.301	2.000
3	4.83	1	1.000	0.684	0.000	1.979
4	9.23	2	1.414	0.965	0.301	1.958
5	15.65	3	1.732	1.195	0.477	1.926
6	22.42	4	2.000	1.351	0.602	1.890
7	31.32	5	2.236	1.496	0.699	1.837
8	39.44	6	2.449	1.596	0.778	1.782
9	47.54	7	2.646	1.677	0.845	1.720
10	56.63	8	2.828	1.753	0.903	1.637
11	63.43	9	3.000	1.802	0.954	1.563
12	71.32	10	3.162	1.853	1.000	1.458
13	82.14	11	3.317	1.915	1.041	1.252
14	94.14	12	3.464	1.974	1.079	0.768



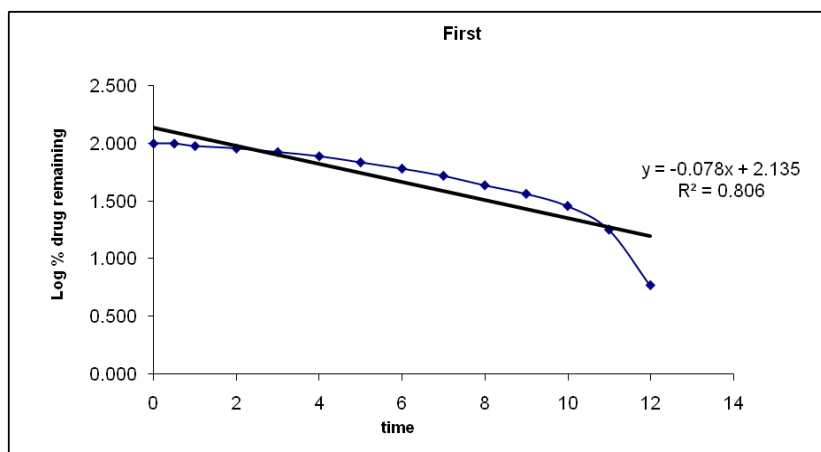
Graph of Zero order release kinetics



Graph of Higuchi release kinetics



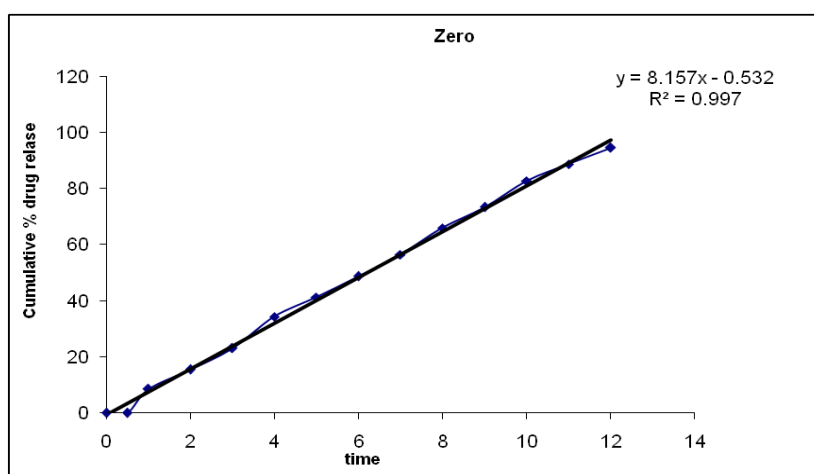
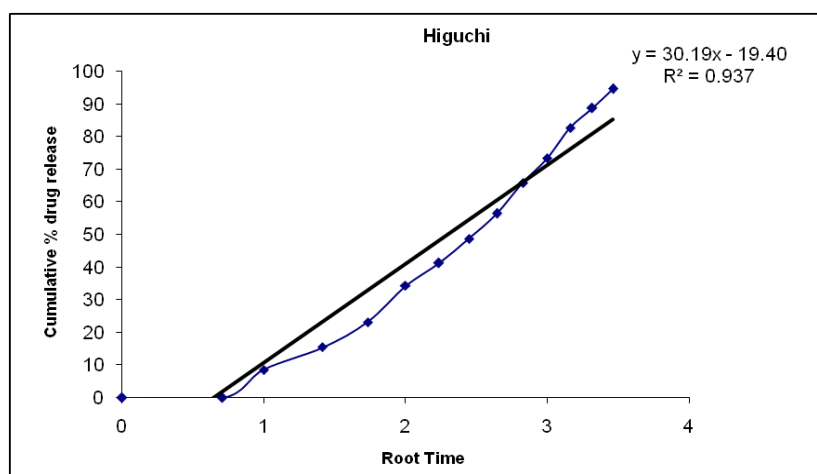
Graph of Peppas release kinetics

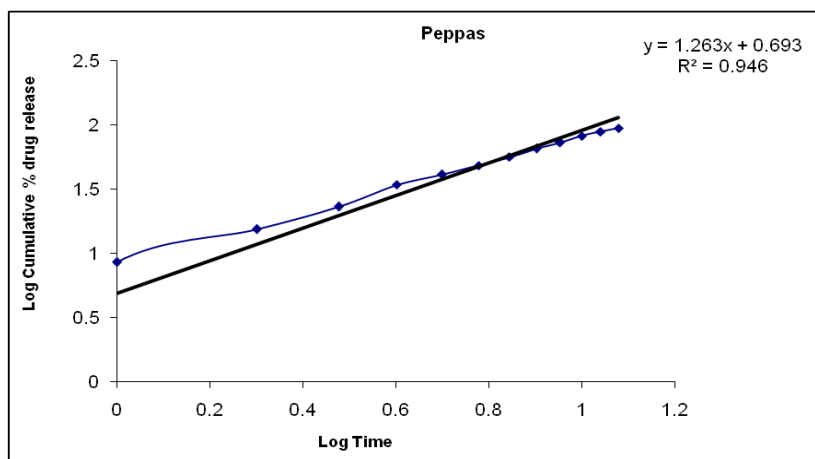


Graph of First order release kinetics

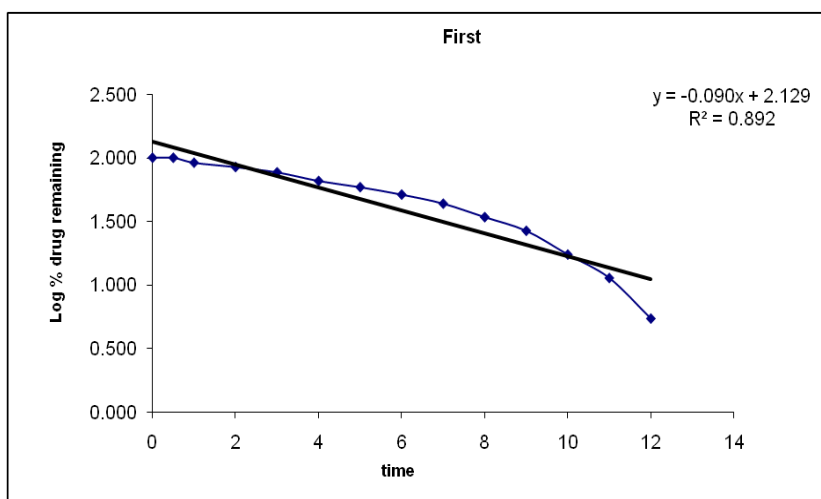
**In Vitro Drug Release Kinetics for Mucoadhesive Microspheres M1 formulation**

S. No	Cumulative (%) Release Q	Time (T)	Root (T)	Log (%) Release	Log (T)	LOG (%) Remain
1	0	0	0			2.000
2	8.56	1	1.000	0.932	0.000	1.961
3	15.48	2	1.414	1.190	0.301	1.927
4	23.18	3	1.732	1.365	0.477	1.885
5	34.29	4	2.000	1.535	0.602	1.818
6	41.28	5	2.236	1.616	0.699	1.769
7	48.65	6	2.449	1.687	0.778	1.711
8	56.43	7	2.646	1.752	0.845	1.639
9	65.87	8	2.828	1.819	0.903	1.533
10	73.34	9	3.000	1.865	0.954	1.426
11	82.65	10	3.162	1.917	1.000	1.239
12	88.65	11	3.317	1.948	1.041	1.055

**Graph of Zero order release kinetics****Graph of Higuchi release kinetics**



Graph of Peppas release kinetics



Graph of First order release kinetics

From the release kinetics data, It was evident that mucoadhesive optimised formula was followed Zero order release kinetics.

## CONCLUSION

The floating and mucoadhesion microspheres were studied for micromeritic properties such as bulk density, tapped density, carrs index, hausners ratio, angle of repose which were found to be within limits. The percentage yield of floating microsphere formulation f1 to f6 and mucoadhesive microspheres m1 to m3 were in range of  $77.14 \pm 0.64$  to  $92.74 \pm 0.74\%$ . The *in vitro* buoyancy of formulation f1 to f6, it was range from  $71.96 \pm 1.04$  to  $82.96 \pm 1.07$ . Among all formulation f6 was found to be highest *in-vitro* buoyancy  $82.96 \pm 1.07$ . The results also showed that the larger the particle size, the longer the floating time. The entrapment efficiency of floating microspheres f1 to f6 and mucoadhesive microspheres were in the range of  $77.43 \pm 2.72$  to  $98.11 \pm 2.59$ . Formulations prepared with sodium alginate alone has

shown maximum drug release at 12 hr in the ratio of 1:3. Formulations prepared with sodium alginate along with HPMC K 4M retards the drug release. Among all formulations of floating microspheres F3 was considered as optimised for floating microspheres. From the release kinetics data, it was evident that floating optimised formulation follows zero order release kinetics. From the dissolution data of mucoadhesive microspheres by ionic gelation method M1 formulation has shown maximum drug release at 12 hrs. When increase in the polymer concentration retards the drug release more than 12 hrs. Hence M1 was considered as optimised formulation for mucoadhesive microspheres. From the release kinetics data, it was evident that mucoadhesive optimised formula was followed zero order release kinetics.

## REFERENCES

1. Banker GS, Anderson NR. Tablets: the theory and practice of industrial pharmacy. 3<sup>rd</sup>. Bombay: Varghese Pub. House; 2003.
2. Chein YW. Novel drug delivery systems. 2<sup>nd</sup> ed. New York: Marcel Dekker, Inc.; 1992.
3. Lee TW, Robinson JR. Remington: the science and practice of pharmacy. 20<sup>th</sup> ed. Pennsylvania: Mack Publishing Company; 2001.
4. Aulton ME. Pharmaceutics: the science of dosage form design. 2<sup>nd</sup> ed. Livingstone C. Elsevier Science Ltd; 2002.
5. Welling PG, Dobrinska. Controlled drug delivery: fundamentals and applications. 2<sup>nd</sup> ed. New York: Marcel Dekker Inc.; 1987.
6. Brahmkar DM, Jaiswal SB. Bio pharmaceutics and pharmacokinetics a treatise. Reprint of 1st edn. Delhi: Vallabh Prakashan; 2003.
7. Anurag Sood, Ramesh Panchagnula. Design of controlled release delivery systems using a modified pharmacokinetic approach: a case study for drugs having a short elimination half-life and a narrow therapeutic index. International journal of pharmaceutics 2003; 261: 27–41.
8. Yeole PG, Khan S, Patel VF. Floating drug delivery systems: need and development. Indian J Pharm Sci., 2005; 67(3): 265-272.
9. Vyas SP, Khar RK. Gastro-retentive system in: controlled drug delivery system: concept & advances. 1st ed. New Delhi: Vallabh Prakashan, 2002.
10. Arora S, Ali J, Ahuja A, Khar RK, Baboota S. Floating drug delivery system. A review. Aaps Pharm Sci Tech, 2005; 6(3): 372-390.
11. Chawla G, Gupta P, Koradia V, Bansal AK. Gastroretention a means to address regional variability in intestinal drug absorption. Pharmaceutical technology, 2003: 50-68.

12. Singh b, ahuja n. Progress in controlled and novel drug delivery system. New delhi: cbs publishers and distributors, 2004.
13. Etyan, ak, eran, l, michel, f, hoffman, a. Expandable gastroretentive dosage forms. J. Cont. Rel., 2003; 909: 143-162.
14. Chatterjee cc. Human physiology, medical allied agency, 11<sup>th</sup> edition. 2001.
15. Garg s, sharma s. Gastroretentive drug delivery systems. Business brief pharmatech 5th ed. 2003: available at: <http://www.touchbriefings.com/cdps/cditem.cfm?Nid-17&cid-5>.
16. Singh bn, kwon hk. Floating drug delivery systems: an approach to oral controlled drug delivery via gastric retention. Journal of controlled release, 2000; 63: 235–249.
17. Amnon hoffmana, david sa, eran lb, sara ea, eytan ka, michael fa. Pharmacokinetic and pharmacodynamics aspects of gastroretentive dosage forms. International journal of pharmaceutics, 2004; 277: 141–153
18. Bardonnnet pl, faivre v, pugh wj, piffaretti jc, falson f. Gastroretentive dosage forms: overview and special case of helicobacter pylori. Journal of controlled release, 2006; 111: 1-18.
19. Av mayavanshi, ss gajjar. Floating drug delivery systems to increase gastric retention of drugs: a review. Research j. Pharm. And tech, 2008; 1(4): 345- 358.
20. Brahma n. Singh, kwon h him. Floating drug delivery system; an approach to oral control drug delivery via gastric retention. Journal of controlled release, 2000; (63): 235-259.
21. Fix ja, cargill r, engle e. Controlled gastric emptying iii. Gastric residence time of a non-disintegrating geometric shape in human volunteers. Pharm res, 1993; 10: 1087-1089.
22. Chowdary kpr, srinivas l. Mucoadhesive drug delivery systems: a review of current status. Indian drugs, 2000; 37: 400-403.
23. Kedzierewicz f, thouvenot p, lemut j, etienne a, hoffman m, maincent p. evaluation of peroral silicone dosage forms in humans by gammascintigraphy. J control rel, 1999; 58: 195-205.
24. Martin a, swarbrick j, cammarata a. Physical pharmacy. 2nd ed. Bombay: varghese publishing company, 1991.