

WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 8.084

Volume 9, Issue 5, 2519-2540.

Review Article

ISSN 2277-7105

A REVIEW ON POTENTIAL PROPERTIES AND THERAPEUTIC APPLICATIONS OF GRAPE SEED EXTRACT

Bharat Kwatra*

Invenzion Labs and Conceptive Farmacia Pvt. Ltd.

Article Received on 19 March 2020.

Revised on 11 April 2020, Accepted on 01 May 2020,

DOI: 10.20959/wjpr20205-17514

*Corresponding Author Prof. Bharat Kwatra Invenzion Labs and Conceptive Farmacia Pvt

Ltd.

ABSTRACT

Grape seeds are rich in proanthocyanidins which have been shown to possess potent free radical scavenging activity. Grape seeds are a complex matrix containing 40% fiber, 16% oil, 11% proteins, and 7% complex phenols such as tannins. Grape seeds are rich sources of flavonoids and contain monomers, dimers, trimers, oligomers, and polymers. The monomeric compounds includes (+)-catechin, (-)-epicatechin, and (-)-epicatechin-3-O-gallate. Studies have reported that grape seeds exhibit a broad spectrum of pharmacological properties against oxidative stress. Their potential health benefits include protection against oxidative damage, and anti-diabetic, anti-

cholesterol, and anti-platelet functions. Recognition of such health benefits of proanthocyanidins has led to the use of grape seeds as a dietary supplement by the consumers.

KEYWORDS: Grape seeds are rich in proanthocyanidins which have been shown to possess potent free radical scavenging activity.

1. INTRODUCTION

Although grape seeds are a relatively inexpensive source of antioxidant compounds^[5], they amount to 38–52% on a dry matter basis.^[6] Grape seeds are treated as waste if extracts are not made and it is estimated that about 10–12 kg of grape seeds in 100 kg of wet residues are produced by the industry.^[1] In recent years, grape seed extract has become increasingly popular on the market as a nutritional supplement, especially in Australia, Korea, Japan and the United States.^[7] This is because grape seeds are rich in phenolic compounds^[8] and have potentially beneficial effects for human health such as protection against peptic ulcers.^[8,9]

Grape seeds have been reported to exhibit scavenge superoxide radicals.^[10] Grape seeds are rich in flavan-3-ol, including proanthocyanidins and catechins.^[8]

They contain high concentration of polyphenol proanthocyanidins, which are the oligomers Antioxidants of flavan-3-ol 2017, 6, 71; doi:10.3390/antiox6030071 www.mdpi.com/journal/antioxidants Antioxidants 2017, 6, 71 2 of 11 units including catechin and epicatechin.^[11] Proanthocyanidins, which belong to condensed tannins, are present as procyanidins and prodelphinidins in grape seeds and skins.^[12] Procyanidins and prodelphinidins are extracted from the seeds and skins during fermentation into wine. [12] Anthocyanins and proanthocyanidins play a significant role in the stability, taste, and color of the red wines.^[13] The simplest proanthocyanidins are dimeric proanthocyanidins which have 4→8 linked monomers. [14] Therefore, the aim of this work was to summarize the phytochemical constituents, pharmacological activities, and industrial applications of grape seeds. We hope that this work would be a valuable reference resource to provide beneficial inspiration for the future studies.

Search Strategy

An electronic literature search was conducted using PubMed, Medline (OvidSP), DOAJ, Researchgate and Google Scholar until September 2019. Additional articles were identified from references in the retrieved articles. Search terms included combinations of the following: "grape", "grape seed", "phytochemical", "antioxidant", "anthocyanins", "proanthocyanidins", and "pharmacological". The search was restricted to articles in English that addressed the phytochemical constituents and pharmacological properties of grape seeds. For the purpose of this review, studies involving human subjects will be given priority.

2. Phytochemical Compounds in Grape Seeds

Grape seeds contain protein (11%), fiber (35%), minerals (3%), and water (7%).^[15] In addition, the lipid content of grape seeds ranges from 7 to 20%.^[15] The oil content of grape seeds is traditionally extraction using mechanical methods or organic solvents.^[4] In mechanical extraction, although the quality of product is superior, the extraction gives a lower yield. While organic solvent extraction gives a higher yield, it requires solvent recovery through distillation and the final product contains traces of residual solvent.^[4] While the supercritical method is regarded as a promising method which can produce similar quantity and better quality of oil yield than mechanical and organic solvent extractions.^[4] Cold-pressing is used to extract the oil from grape seeds without chemical treatment or

heat.^[16] Although the cold pressing usually gives a lower yield than other conventional solvent extraction, it may retain more bioactive components and be safer because there are no solvent residues in the grape seed oil.^[15] Several studies have been conducted on grape seeds, in order to determine their bioactive compounds.^[8,17–21] Grape seed extracts contain a heterogeneous mixture of monomers (5–30%), oligomers (17–63%), and polymers (11–39%) composed of proanthocyanidins.^[22]

Proanthocyanidins are the major compound in grape seed extracts. [23] The red color and astringent taste of grape seed extracts can be attributed to proanthocyanidins. However, higher concentrations of proanthocyanidins may affect the sensory and color properties of the product. [24] Negro et al. [25] reported that the total phenols (gallic acid equivalent (GAE)), total flavonoids (catechin equivalent (CE)), catechin equivalent (CE), and proanthocyanidins (cyanidin equivalent (CyE)) in red grape seed extracts were 8.58 g/100 g dry matter (DM), 8.36 g/100 g DM, 6.41 g/100 g, DM and 5.95 g/100 g DM, respectively. Another study by Ghouila et al. [26] documented that the total phenolic content and total flavonoid content of Ahmeur Bouamer grape seeds were 265.15 mg GAE/g of dry mass and 14.08 mg catechin equivalent (CE)/g of dry mass, respectively. In a study analyzing 26 samples of six white grape varieties and 44 of four red grape varieties from different areas of Castilla-La Mancha, Spain, Rodríguez Montealegre et al. [8] reported that the grape seeds were shown to contain catechin, epicatechin, epicatechin gallate, protocatechuic acid, procyanidin B1, procyanidin B2, procyanidin B3, and procyanidin B4. Small quantities of gallic acid and protocatechuic acid were also present in grape seeds. [8] Similarly, another study by Escribano-Bailon et al. [17] reported that the major constituents in V. vinifera (Tinta del pais) grape seeds were (+)catechin (11%) followed by (-)-epicatechin (10%), (-)-epicatechin-3-O-gallate (9%), epicatechin 3-O-gallate-(4β→8)-catechin (B1-3-O-gallate) (7%), Antioxidants 2017, 6, 71 3 of 11 and epicatechin- $(4\beta \rightarrow 8)$ -epicatechin (dimer B2) (6%).

These results were also similar with previous studies conducted on the polyphenols of grape seeds from other varieties of grape species in Greek islands. [21,27,28] Anastasiadi et al. [21] reported that the total phenolic content in grape seeds ranged from 325 to 812 mg/g gallic acid equivalents. There are 14 dimeric, 11 trimeric procyanidins, and 1 tetrameric procyanidin that have been identified from grape seeds. [18] In a study that investigated the quantities of flavan-3-ols in the seeds of 17 grape cultivars grown in Ontario, Canada using high-performance liquid chromatography (HPLC), Fuleki and Ricardo da Silva [18]

documented the presence of monomers of (+)-catechin, (-)-epicatechin-3-O-gallate, and (-)-epicatechin. Considerable quantities of highly polymerized procyanidins were also found in grape seeds. Similarly, Prieur et al. also reported the presence of catechin, epicatechin-3-O-gallate, and epicatechin were determined by HPLC following the thiolysis degradation, providing evidence that procyanidins are the grape seed tannins. Gabetta et al. documented the presence of monomers, dimers, trimers, tetramers, pentamers, hexamers, heptamer, and their gallates in grape seeds. A study by Freitas et al. obtained several fractions of oligomeric and polymeric procyanidins from grape seeds using gel chromatography. The authors also identified several grape seed flavan-3-ol derivatives and phenol acids including protocatechic, caffeic acids, trimer epi-(4 β -8 or 6)-epi-(4 β -6 or 8)-epi, trimer cat-(4 α -8)-epi-(4 β -8)-epi, and trimer cat-(4 α -8)-epi-(4 β -6)-cat.

Some factors such as the variety of grapevine, viticultural, and environmental factors can influence the concentration of phenolic compounds in grapes.^[8] Epicatechin, (+)-catechin, and gallic acid were the major phenolic compounds detected in the muscadine grape seeds.^[20] Pastrana-Bonilla et al.^[20] reported that the concentration of epicatechin, (+)-catechin, gallic acid, total anthocyanin, and total phenolics in muscadine grape seeds were 1299 mg/100 g of fresh weight (FW), 558 mg/100 g of FW, 6.9 mg/100 g of FW, 4.3 mg/100 g of FW, and 2179 mg/g GAE, respectively.

3. Pharmacological Properties

Proanthocyanidins are present in substantial amounts in grape seeds and have attracted the attention of consumers because of their potential health effects.^[31] In vitro, proanthocyanidins have been shown to exhibit strong antioxidant activity and scavenge reactive oxygen and nitrogen species, modulate immune function and platelet activation, and produce vasorelaxation by inducing nitric oxide (NO) release from endothelium.^[31] In addition, proanthocyanidins also inhibits the progression of atherosclerosis and prevent the increase of low-density lipoprotein (LDL) cholesterol concentration.^[32]

3.1. Anti-Diabetic

Montagut et al.^[33] reported Wister female rats treated with 25 mg grape seed procyanidin extract/kg body weight per day for 30 days had an improved homeostatis model assessment-insulin resistance index accompanied by downregulation of primers Glut4, Irs1, and Pparg2 in mesenteric white adipose tissue (WAT), suggesting that grape seed procyanidin has a positive long term-effect on glucose homeostasis. Another study by Montagut et al.^[34]

demonstrated that the oligomeric structures of grape seed procyanidin extracts activated the insulin receptor by interacting and inducing the autophosphorylation of the insulin receptor in order to stimulate the glucose uptake.

3.2. Antioxidant

Different methods including the 1,1-diphenyl-2-picryhidrazyl (DPPH) method and oxygen radical absorbance capacity (ORAC) have been employed to evaluate the antioxidant capacity of phenolic compounds from grape seeds. Poudel et al. [35] reported that the antioxidant capacity of grape seed extract using DPPH ranged between 17 and 92 mmol Trolox® antioxidant equivalent (TE)/g. While the ORAC method was 42 mmol TE/g. [35] Pastrana-Bonilla et al. [20] demonstrated that the antioxidant capacity was the highest in grape seeds (281 µM Trolox equivalent antioxidant capacity (TEAC)/g Antioxidants 2017, 6, 71 4 of 11 of FW) followed by leaves (236 µM TEAC/g of FW), skins (13 µM TEAC/g of FW), and pulps (2.4 µM TEAC/g of FW), suggesting that the grape seeds extracts are promising antioxidant for dietary supplement. Similarly, two studies [36,37] also confirmed the antioxidant activity of grape seed extract by using β-carotene linoleate, linoleic acid peroxidation, DPPH, and phosphomolybdenum complex methods. Puiggròs et al. [38] reported that grape seed procyanidin extracts modulated the expression of antioxidant systems, suggesting that grape seed procyanidin extracts might improve the cellular redox status via glutathione synthesis pathways. Another study by Guo et al. [39] reported that grape seed procyanidins provided protective effect against the ethanol induced toxicity in mouse brain cells. Vinson et al. [40] reported that supplementation of 50 mg/kg and 100 mg/kg grape seed proanthocyanidin extracts in hamsters reduced the formation of foam cells by 50% and 63%, respectively. The development of foam cells is the indicator of early stages of atherosclerosis. [40] In a randomized, double-blind, placebo-controlled study, Preuss et al. [41] reported that the supplementation of grape seed proanthocyanidin extracts and chromium polynicotinate in hypercholesterolemic participants decreased the concentrations of total cholesterol, oxidized LDL, and LDL significantly after two months.

3.3. Anti-Platelet

Grape seed extracts were shown to exhibit anti-platelet action. ^[42] Olas et al. ^[42] reported that grape seed extracts (5–50 μ g/mL) showed a reduction in platelet aggregation, adhesion, and generation of superoxide anion. In addition, the authors ^[42] also found that grape seed extract was more effective than the pure resveratrol solution in the reduction of platelet processes.

3.4. Anti-Cholesterol

Cetin et al. [43] reported that there were no significant changes in levels of superoxide dismutase and catalase of rats supplemented with grape seed extracts when methotrexate (MTX) was administered. The authors [43] suggested that the supplementation of grape seed extract may be protective against oxidative damage induced by MTX which is used as cytotoxic chemotherapeutic agent. [44] In a 12-week of supplementation containing 0, 200, or 400 mg grape seed extract to 61 healthy subjects with LDL cholesterol level of 100 to 180 mg/dL, Sano et al. [45] reported that the 200 mg and 400 mg group reported a significant decrease in malondialdehyde-modified LDL (MDA-LDL) when compared to the baseline level after 12 weeks (p = 0.008 and 0.009, respectively). MDA-LDL is involved in the pathogenesis of arteriosclerosis. [46] In a study that investigated the effect of supplementing a single high-fat meal with 300 mg of proanthocyanidin-rich grape seed extracts in eight male adults, Natella et al. [47] reported that grape seed extracts reduce postprandial oxidative stress by increasing the plasma antioxidant concentration and preventing the increase of lipid hydroperoxides. Consequently, this improves the resistance to oxidative modification of LDL cholesterol. [47] It is suggested that the polyphenols in grape seed extracts activate serum paraoxonase (PON) which prevents the postprandial increase in lipid peroxides. [47] Serum paraoxonase is an enzyme associated with high-density lipoprotein (HDL) that hydrolyzes lipoprotein peroxides and inhibits LDL oxidation. [48]

3.5. Anti-Inflammation

In a study that evaluated the effect of procyanidin from grape seeds on the inflammatory mediators in rat fed with a high fat diet, Terra et al. [49] reported that rats fed with a high fat diet supplemented with procyanidins from grape seeds (345 mg/kg feed) for 19 weeks had a lower plasma C-reactive protein (CRP) level than rats fed with high-fat diet, suggesting that the decrease in plasma CRP is related to a downregulation of CRP mRNA expression in the liver and mesenteric white adipose tissue (WAT).

In addition, the authors^[49] also reported a decrease in the expression of the proinflammatory cytokine tumour necrosis factor alpha (TNF-α) and interleukin 6 (IL-6) in the mesenteric WAT of rats Antioxidants 2017, 6, 71 5 of 11 fed with a high fat diet supplemented with procyanidins from grape seeds. IL-6 is a stress-induced inflammatory cytokine, which is directly implicated in atherogenesis.^[49] Studies have revealed that the concentrations of proinflammatory mediators are elevated in the conditions such as insulin resistant states of

diabetes and obesity. ^[50] In a study that investigated whether procyanidins play a role in modulating inflammation, Chacon et al. ^[51] reported that macrophage-like (THP-1) cell lines and human adipocytes (SGBS) pre-treated with grape-seed procyanidin extracts had a reduction in IL-6 and monocyte chemoattractant protein (MCP-1) expression after inflammatory stimulus. In addition, the authors ^[51] also reported that the translocation of NF- κ B to the nucleus was partially inhibited in macrophage-like (THP-1) cell lines and human adipocytes (SGBS) pre-treated with grape-seed procyanidin extracts. Terra et al. ^[52] reported that rats fed with grape-seed procyanidin extracts had a reduced body weight and plasmatic systemic markers of TNF- α and CRP.

In addition, grape-seed procyanidin extracts also increased adiponectin expression and decreased IL-6.^[52] Grape-seed procyanidin extracts are also linked to a reduced expression of epidermal growth factor module-containing mucin-like receptor 1 (EMR1) (specific marker of macrophage F4/80), suggesting a reduced macrophage infiltration of WAT.^[52] Therefore, the regular consumption of food containing procyanidins might help to prevent low-grade inflammatory-related diseases in obesity characterized by macrophage accumulation in WAT and abnormal cytokine production.^[52] In a study that investigated the effect of grape seed procyanidins on glucose metabolism in streptozotocin-induced diabetic rats, Pinent et al.^[53] reported that grape seed procyanidins stimulated the glucose uptake in L6e9 myotubes and 3T3-L1 adipocytes in a dose-dependent manner. In addition, grape seed procyanidins also stimulated glucose transported-4 translocation to the plasma membrane.^[53] The authors^[53] suggested that procyanidins possesses insulin-like effects in insulin sensitive cells. Therefore, all these findings suggest that procyanidins in grape seeds inhibited mRNA levels and decreased the risk of diseases associated with obesity and high fat diets.

3.6. Anti-Aging

In a study investigating the effect of grape seed extracts on the aged-associated accumulation of oxidative DNA damage in male albino rats of Wister strain, Balu et al.^[54] reported that grape seed extracts inhibited the accumulation of age-related oxidative DNA damage products such as 8-hydroxy-20 -deoxyguanosine (8-OHdG) and DNA protein cross-links in spinal cord and in various brain regions including the striatum, cerebral cortex, and hippocampus. Another study by Balu et al.^[55] reported that lipid peroxidation and antioxidant defenses in male albino rats supplemented with grape seed extract were normalized, suggesting that grape seed extracts could be used to improve the antioxidant status and

decreased the incidence of free radical-induced lipid peroxidation in the central nervous system of aged rats.

3.7. Anti-Microbial

Since grape seeds are rich sources of polyphenols, grape seeds have also shown promise as novel microbial agents. Jayaprakasha et al. [37] reported that the defatted grape seed extracts were shown to exhibit antibacterial effect against Bacillus cereus (B. cereus), Bacillus subtilis (B. subtilis), Staphylococcus aureus (S. aureus), Bacillus coagulans (B. coagulans), Escherichia coli (E. coli), and Pseudomonas aeruginosa (P. aeruginosa). In addition, the authors demonstrated that these extracts completely inhibited Gram-positive bacteria and Gram-negative bacteria at 850–1000 ppm and 1250–1500 ppm, respectively. A study by Ghouila et al. [26] demonstrated that grape seed extracts exhibited antimicrobial activity to Micrococcus luteus ATCC®9341, S. aureus ATCC®29213, E. coli ATCC®25992, P. aeruginosa ATCC®27853, Aspergillus niger, Fusarium oxysporum, with a diameter of the inhibition growth zone ranged from 15-20 mm. Baydar et al. [56] reported that grape seed extracts acted against S. aureus after 48 h and Aeromonas hydrophila after 1 h. A study by Anastasiadi et al. [21] reported that the minimum inhibition Antioxidants 2017, 6, 71 6 of 11 concentration of grape seed extract against Listeria monocytogenes (L. monocytogenes) was 0.26, suggesting that grape seeds could be used as an inexpensive source of natural antilisterial mixtures. Future research should be conducted on screening the grape seed extracts for natural antimicrobial compounds. Another study by Brown et al. [57] reported that muscadine grape seed extracts were effective in inhibiting H. pylori in vitro. The authors^[57] demonstrated that although muscadine grape seed extracts had the highest total phenolic content (646 mg GAE/g dry weight (dw)) than skin extracts (135 mg GAE/g dw), the skin extracts had a better efficacy than seed extracts against H. pylori. [57] Therefore, it is suggested that the anti-H. pylori activity might be influenced by the type and content of phenolics. [57] H. pylori infection is associated with several gastroduodenal diseases such as gastric cancer. [58]

3.8. Anti-Tumour

Grape seed extracts have also exhibited anti-tumor properties. Promising results have been shown by several studies performed in human colorectal carcinoma^[59], head and neck squamous cell carcinoma^[60], and prostate cancer cells.^[61] Therefore, it is suggested that the supplementation of grape seed extracts might be an effective anti-tumor agent in clinical settings.

3.9. Other Pharmacological Properties

A study by Feng et al.^[62] reported that grape seed extracts reduced brain weight loss from 20% in vehicle pups to 3% in treated pups (p < 0.01). The loss of brain weight was used to measure the brain damage in pups with neonatal hypoxic ischemic brain injury. In addition, grape seed extract also improved the histopathologic brain score in cortex, hippocampus, and thalamus. The authors^[62] suggested that the treatment with grape seed extracts could be used to inhibit lipid peroxidation and reduce hypoxic ischemic brain injury.

3.10. Toxicity

Grape seed extract has been approved as generally recognized as safe (GRAS) by the Food and Drug Administration (FDA) and is sold commercially as a dietary supplement listed on the Everything Added to Food in the United States (EAFUS) database. Bentivegna and Whitney reported that no-observed-adverse effect level (NOAEL) of grape seed extracts determined in the rats was 1.78 g/kg body weight/day and the normal amount of grape seed extract used in food applications was 0.01–1%. Sano et al. also reported that there were no abnormal changes in the physiological and clinical laboratory tests in participants after taking tablets containing 200 mg and 400 mg grape seed extracts. In addition, no problematic results were reported in the urinary sedimentation of these participants, suggesting that the tablets containing 200 and 400 mg grape seed extracts are safe to consume.

Similarly, a subchronic oral toxicity study by Yamakoshi et al.^[7] documented that there was no noticeable sign of toxicity in rat fed with grape seed extract as a dietary admixture at 0.02% (low dose group), 0.2% (middle dose group), and 2% (high dose group) (w/w) for 90 days. The authors^[7] reported that the NOAEL of grape seed extract used in the subchronic oral toxicity study was 2% in the diet, which was equivalent to 1410 mg/kg body weight in males and 1501 mg/kg body weight/day in females. Therefore, these studies indicate a lack of toxicity and more systematic toxicological studies on grape seed should be conducted.

4. Industrial Applications

Lipid peroxidation is one of the major reasons for the deterioration of food products during processing and storage. Therefore, antioxidants have been added to food products to extend their shelf life. Synthetic antioxidants such as tertiary butylhydroquinone (TBHQ), butylated hydroxyanisole (BHA), and butylated hydroxytoluene (BHT) have restricted use in food products because of their carcinogenic and toxic effects. [65] Therefore, the search for natural antioxidants, particularly of plant origin, has greatly increased in recent years. Antioxidants

2017, 6, 71 7 of 11 Grape seeds can also be exploitable for the preservation of food products. For example, grape seed extracts have been used as a food additive in Japan. A study by Jayaprakasha et al. reported that the grape seed extracts showed good reducing power at a 500 μg/L concentration when using potassium ferricyanide reduction methods. In addition, grape seed extracts also showed 65–90% antioxidant activity at 100 ppm. Grape seed extracts have been evaluated for their antioxidative effect on meat products. [66,67]

Ahn et al.^[66] reported that the addition of grape seed extracts had an improved oxidative stability and reduced the hexanal concentration by 97% in cooked ground beef after three days of refrigerated storage when compared to the control without added antioxidant. Similarly, Lau and King^[67] also reported similar improve oxidative stability in turkey patties. In addition, the authors^[67] also reported that when the grape seed extracts were added at 1.0% and 2.0% to turkey patties, the thiobarbituric acid reactive substances (TBARS) values were decreased nearly 10-fold when compared to the control. The grape seed extracts added to turkey patties were also reported to exhibit wine odor and slightly bitter aftertaste.^[67] In a study that investigated the effect of four different concentrations of grape seed extracts (i.e., 0.0, 0.4, 0.8, and 1.6 g/kg) on cooked turkey breast meat, Mielnik et al.^[68] reported that the addition of grape seed extracts before cooking significantly improved the lipid stability of turkey breast meat during the cooking process and chill storage.

The authors^[68] reported that the efficiency of grape seed extract to prevent oxidation increased when the concentration of antioxidant was from 0.4 g/kg to 1.6 g/kg. Therefore, it is suggested that grape seed extracts can be a very effective antioxidant in inhibiting lipid oxidation of cooked turkey meat during chill-storage. Similarly, Brannan^[69] also demonstrated a reduction in the biomarkers of lipid oxidation and sensory scores for key attributes in ground chicken when the grape seed extracts were added during refrigerated storage. Brannan and Mah^[70] documented that 0.1% grape seed extract was effective in inhibiting the formation of primary (e.g., hexanal and lipid hydroperoxides) and secondary oxidation products (e.g., TBARS) in both raw and cooked meat systems during refrigerated and frozen storage. It is suggested that in vivo mechanism of antioxidant activity for grape seed extracts includes the inhibition of nitrositive stress, stimulation of enzymatic production of nitric oxide^[71], and oxygen radical scavenging.^[72] The antioxidant activity of grape seed extracts can be influenced by heating conditions such as temperature and time. In a study that evaluated the effect of heating conditions on the antioxidant activity of grape seed extracts,

Kim et al.^[73] reported that heat treatment significantly increased the concentrations of caffeine and gallocatechin gallate in grape seed extracts, suggesting that heating could be used as a method to improve the antioxidant activity of grape seed extracts.

Since polyphenols in grape seeds possess antibacterial activity, grape seeds could be incorporated into the foods in order to prevent the growth of foodborne pathogens such as L. monocytogenes.^[21] Grape seed extracts are also included in the formulations of cosmetic products for anti-aging reasons. This is because grape seed extracts are rich in proanthocyanidins which have strong free radical scavenging properties.^[74]

5. CONCLUSIONS AND FUTURE RESEARCH

Grape seeds have high antioxidant potential. Their potential health benefits include protection against oxidative damage, and anti-diabetic, anti-cholesterol, and anti-platelet properties. The health benefits of grape seed consumption are thought to arise mainly from bioactivities of their polyphenols. Potential health benefits of dietary polyphenols on major chronic non-communicable diseases have been shown in several meta-analyses.^[75–78]

Therefore, the screening of individual polyphenol constituents that exhibit health-promoting properties in grape seed requires further investigation. This is because a cause-effect relationship between the intake of grape seed and its health effects can only be established when the composition of grape seeds is properly characterized and standardized. Furthermore, extensive investigation is needed to examine the effects of adding these beneficial Antioxidants 2017, 6, 71 8 of 11 polyphenol constituents from grape seeds using advanced technologies into food systems. Further research is needed to evaluate the effectiveness of grape seed extract in the food ecosystem and to establish their role as an antimicrobial agent in food safety.

REFERENCES

- 1. Matthäus B. Virgin grape seed oil: Is it really a nutritional highlight? Eur. J. Lipid Sci. Technol, 2008; 110: 645–650. doi: 10.1002/ejlt.200700276. [CrossRef] [Google Scholar]
- Girard B., Mazza G. Functional grape and citrus products. In: Mazza G., editor. Functional Foods—Biochemical and Processing Aspects. Volume 1. Technomic Publishing Co., Inc.; Lancaster, PA, USA, 1998; 139–191. [Google Scholar]

- 3. Estreicher S.K. Wine. In: Bagnall R.S., Brodersen K., Champion C.B., Erskine A., Huebner S.R., editors. The Encyclopedia of Ancient History. John Wiley & Sons, Inc.; Oxford, UK, 2013; 1–5. [Google Scholar]
- 4. Duba K.S., Fiori L. Supercritical CO2 extraction of grape seed oil: Effect of process parameters on the extraction kinetics. J. Supercrit. Fluids, 2015; 98: 33–43. doi: 10.1016/j.supflu.2014.12.021. [CrossRef] [Google Scholar]
- Cádiz-Gurrea M., Borrás-Linares I., Lozano-Sánchez J., Joven J., Fernández-Arroyo S., Segura-Carretero A. Cocoa and grape seed byproducts as a source of antioxidant and antiinflammatory proanthocyanidins. Int. J. Mol. Sci, 2017; 18: 376. doi: 10.3390/ijms18020376. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 6. Maier T., Schieber A., Kammerer D.R., Carle R. Residues of grape (Vitis vinifera L.) seed oil production as a valuable source of phenolic antioxidants. Food Chem, 2009; 112: 551–559. doi: 10.1016/j.foodchem.2008.06.005. [CrossRef] [Google Scholar]
- 7. Yamakoshi J., Saito M., Kataoka S., Kikuchi M. Safety evaluation of proanthocyanidin-rich extract from grape seeds. Food Chem. Toxicol, 2002; 40: 599–607. doi: 10.1016/S0278-6915(02)00006-6. [PubMed] [CrossRef] [Google Scholar]
- 8. Rodríguez Montealegre R., Romero Peces R., Chacón Vozmediano J.L., Martínez Gascueña J., García Romero E. Phenolic compounds in skins and seeds of ten grape Vitis vinifera varieties grown in a warm climate. J. Food Comp. Anal, 2006; 19: 687–693. doi: 10.1016/j.jfca.2005.05.003. [CrossRef] [Google Scholar]
- Kim T.H., Jeon E.J., Cheung D.Y., Kim C.W., Kim S.S., Park S.H., Han S.W., Kim M.J., Lee Y.S., Cho M.L. Gastroprotective effects of grape seed proanthocyanidin extracts against nonsteroid anti-inflammatory drug-induced gastric injury in rats. Gut Liver, 2013;
 282–289. doi: 10.5009/gnl.2013.7.3.282. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 10. El-Beshbishy H.A., Mohamadin A.M., Abdel-Naim A.B. In vitro evaluation of the antioxidant activities of grape seed (Vitis vinifera) extract, blackseed (Nigella sativa) extract and curcumin. J. Taibah Univ. Med. Sci, 2009; 4: 23–35. [Google Scholar]
- 11. Weseler A.R., Bast A. Masquelier's grape seed extract: From basic flavonoid research to a well-characterized food supplement with health benefits. Nutr. J, 2017; 16: 5. doi: 10.1186/s12937-016-0218-1. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 12. Sun B., Spranger M.I. Review: Quantitative extraction and analysis of grape and wine proanthocyanidins and stilbenes. Ciência e Técnica Vitivinícola, 2005; 20: 59–89. [Google Scholar].

- 13. He F., Liang N., Mu L., Pan Q., Wang J., Reeves M.J., Duan C. Anthocyanins and their variation in red wines I. Monomeric anthocyanins and their color expression. Molecules, 2012; 17: 1571–1601. doi: 10.3390/molecules17021571. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 14. Yilmaz Y., Toledo R.T. Health aspects of functional grape seed constituents. Trends Food Sci. Technol, 2004; 15: 422–433. doi: 10.1016/j.tifs.2004.04.006. [CrossRef] [Google Scholar].
- Shinagawa F.B., Santana F.C.D., Torres L.R.O., Mancini-Filho J. Grape seed oil: A potential functional food? Food Sci. Technol. 2015; 35: 399–406. doi: 10.1590/1678-457X.6826. [CrossRef] [Google Scholar].
- 16. Parry J., Hao Z., Luther M., Su L., Zhou K., Yu L. Characterization of cold-pressed onion, parsley, cardamom, mullein, roasted pumpkin, and milk thistle seed oils. J. Am. Oil Chem. Soc, 2006; 83: 847–854. doi: 10.1007/s11746-006-5036-8. [CrossRef] [Google Scholar]
- 17. Escribano-Bailon T., Gutierrez-Fernandez Y., Rivas-Gonzalo J.C., Santos-Buelga C. Characterization of procyanidins of Vitis vinifera variety Tinta del Pais grape seeds. J. Agric. Food Chem, 1992; 40: 1794–1799. doi: 10.1021/jf00022a013. [CrossRef] [Google Scholar]
- 18. Fuleki T., Ricardo da Silva J.M. Catechin and procyanidin composition of seeds from grape cultivars grown in Ontario. J. Agric. Food Chem, 1997; 45: 1156–1160. doi: 10.1021/jf960493k. [CrossRef] [Google Scholar]
- 19. Gabetta B., Fuzzati N., Griffini A., Lolla E., Pace R., Ruffilli T., Peterlongo F. Characterization of proanthocyanidins from grape seeds. Fitoterapia, 2000; 71: 162–175. doi: 10.1016/S0367-326X(99)00161-6. [PubMed] [CrossRef] [Google Scholar]
- 20. Pastrana-Bonilla E., Akoh C.C., Sellappan S., Krewer G. Phenolic content and antioxidant capacity of muscadine grapes. J. Agric. Food Chem, 2003; 51: 5497–5503. doi: 10.1021/jf030113c. [PubMed] [CrossRef] [Google Scholar]
- 21. Anastasiadi M., Chorianopoulos N.G., Nychas G.J.E., Haroutounian S.A. Antilisterial activities of polyphenol-rich extracts of grapes and vinification byproducts. J. Agric. Food Chem, 2009; 57: 457–463. doi: 10.1021/jf8024979. [PubMed] [CrossRef] [Google Scholar]
- 22. Waterhouse A.L., Ignelzi S., Shirley J.R. A comparison of methods for quantifying oligomeric proanthocyanidins from grape seed extracts. Am. J. Enol. Vitic, 2000; 51: 383–389. [Google Scholar]

- 23. Hernández-Jiménez A., Gómez-Plaza E., Martínez-Cutillas A., Kennedy J.A. Grape skin and seed proanthocyanidins from Monastrell × Syrah grapes. J. Agric. Food Chem, 2009; 57: 10798–10803. doi: 10.1021/jf903465p. [PubMed] [CrossRef] [Google Scholar]
- 24. Monteleone E., Condelli N., Dinnella C., Bertuccioli M. Prediction of perceived astringency induced by phenolic compounds. Food Qual. Prefer, 2004; 15: 761–769. doi: 10.1016/j.foodqual.2004.06.002. [CrossRef] [Google Scholar]
- 25. Negro C., Tommasi L., Miceli A. Phenolic compounds and antioxidant activity from red grape marc extracts. Bioresour. Technol, 2003; 87: 41–44. doi: 10.1016/S0960-8524(02)00202-X. [PubMed] [CrossRef] [Google Scholar]
- 26. Ghouila Z., Laurent S., Boutry S., Vander Elst L., Nateche F., Muller R., Baaliouamer A. Antioxidant, antibacterial and cell toxicity effects of polyphenols from Ahmeur Bouamer grape seed extracts. J. Fund. Appl. Sci, 2017; 9: 392–410. doi: 10.4314/jfas.v9i1.24. [CrossRef] [Google Scholar]
- 27. Guendez R., Kallithraka S., Makris D.P., Kefalas P. An analytical survey of the polyphenols of seeds of varieties of grape (Vitis vinifera) cultivated in Greece: Implications for exploitation as a source of value-added phytochemicals. Phytochem. Anal, 2005; 16: 17–23. doi: 10.1002/pca.804. [PubMed] [CrossRef] [Google Scholar]
- 28. Guendez R., Kallithraka S., Makris D.P., Kefalas P. Determination of low molecular weight polyphenolic constituents in grape (Vitis vinifera sp.) seed extracts: Correlation with antiradical activity. Food Chem, 2005; 89: 1–9. doi: 10.1016/j.foodchem.2004.02.010. [CrossRef] [Google Scholar]
- 29. Prieur C., Rigaud J., Cheynier V., Moutounet M. Oligomeric and polymeric procyanidins from grape seeds. Phytochemistry, 1994; 36: 781–784. doi: 10.1016/S0031-9422(00)89817-9. [CrossRef] [Google Scholar]
- 30. De Freitas V.A.P., Glories Y., Bourgeois G., Vitry C. Characterisation of oligomeric and polymeric procyanidins from grape seeds by liquid secondary ion mass spectrometry. Phytochemistry, 1998; 49: 1435–1441. doi: 10.1016/S0031-9422(98)00107-1. [CrossRef] [Google Scholar]
- 31. Varzakas T., Zakynthinos G., Verpoort F. Plant food residues as a source of nutraceuticals and functional foods. Foods, 2016; 5: 88. doi: 10.3390/foods5040088. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 32. Quesada H., del Bas J.M., Pajuelo D., Diaz S., Fernandez-Larrea J., Pinent M., Arola L., Salvado M.J., Blade C. Grape seed proanthocyanidins correct dyslipidemia associated with a high-fat diet in rats and repress genes controlling lipogenesis and VLDL

- assembling in liver. Int. J. Obes, 2009; 33: 1007–1012. doi: 10.1038/ijo.2009.136. [PubMed] [CrossRef] [Google Scholar]
- 33. Montagut G., Bladé C., Blay M., Fernández-Larrea J., Pujadas G., Salvadó M.J., Arola L., Pinent M., Ardévol A. Effects of a grapeseed procyanidin extract (GSPE) on insulin resistance. J. Nutr. Biochem, 2010; 21: 961–967. doi: 10.1016/j.jnutbio.2009.08.001. [PubMed] [CrossRef] [Google Scholar]
- 34. Montagut G., Onnockx S., Vaqué M., Bladé C., Blay M., Fernández-Larrea J., Pujadas G., Salvadó M.J., Arola L., Pirson I., et al. Oligomers of grape-seed procyanidin extract activate the insulin receptor and key targets of the insulin signaling pathway differently from insulin. J. Nutr. Biochem, 2010; 21: 476–481. doi: 10.1016/j.jnutbio.2009.02.003. [PubMed] [CrossRef] [Google Scholar]
- 35. Poudel P.R., Tamura H., Kataoka I., Mochioka R. Phenolic compounds and antioxidant activities of skins and seeds of five wild grapes and two hybrids native to Japan. J. Food Comp. Anal, 2008; 21: 622–625. doi: 10.1016/j.jfca.2008.07.003. [CrossRef] [Google Scholar].
- 36. Kwatra B. Effects of Mineral Separation by Time and Enteric Coating Mechanism for Calcium and Iron Absorption in Mammalia. Int J Sci Res [Internet], 2019; 8(12): 1265–70. Available from: https://www.ijsr.net/get_abstract.php?paper_id=13121901
- 37. Kwatra B. HYDROQUINONE: A novel growth inhibitor and apoptosis inducer in U-251 MG CELLS. Int J Med Biomed Stud [Internet], 2019 Jun 6; 3(6): 15–6. Available from: https:
 - //www.researchgate.net/publication/333726255_HYDROQUINONE_A_novel_growth_i nhibitor_and_apoptosis_inducer_in_U-251_MG_CELLS.
- 38. Kwatra B, Arora C. THE UNEXPLAINED SIMILARITY BETWEEN THE ATOMIC AND GRAVITATIONAL MODELS. Int J Adv Res [Internet], 2020 Mar 31; 8(3): 1099–107. Available from: http://www.journalijar.com/article/31993/the-unexplained-similarity-between-the-atomic-and-gravitational-models/.
- 39. Kwatra B. A REVIEW ON POTENTIAL PROPERTIES AND THERAPEUTIC APPLICATIONS OF LYCOPENE. Int J Med Biomed Stud [Internet], 2020; 4(4): 33–44. Available from: https://doi.org/10.32553/ijmbs.v4i4.1081.
- 40. Kwatra B, Mudgil M. LIGHT ASSISTED TIO2-BASED NANOCOMPOSITE SYSTEMS: A NOVEL TREATMENT FOR CANCER. Int J Med Biomed Stud [Internet]. 2020; 4(4): 28–32. Available from: https://doi.org/10.32553/ijmbs.v4i4.1080.
- 41. Kwatra B. A Review on Potential Properties and Therapeutic Applications of Vitamin D.

- Int J Sci Res [Internet], 2020; 9(4): 682–91. Available from: https://www.researchgate.net/publication/340595290_A_Review_on_Potential_Properties_and _Therapeutic_Applications_of_Vitamin_D
- 42. Kwatra B. THE SIMVASTATIN AND DMXAA ON THE CO-CULTURE OF B16.F10 MELANOMA CELLS AND MACROPHAGES SHOWS ANTITUMOR ACTIVITY. World J Pharm Res [Internet], 2019; 8(9): 1318–9. Available from: https://wipr.net/dashboard/abstract_id/12821
- 43. Kwatra B, Mudgil M. PROTONATED CRAB SHELL WASTE AS FUNGAL INHIBITOR. Int J Med Biomed Stud [Internet], 2019 Apr 17; 3(4): 111–6. Available from:

 https://www.researchgate.net/publication/332537052_PROTONATED_CRAB_SHELL_WAS TE_AS_FUNGAL_INHIBITOR.
- 44. Bharat Kwatra. Procuring Natural Dye for Solar Cell Using Leaf Waste. Int J Sci Eng Res [Internet], 2019; 7(5): 46–7. Available from: https://www.ijser.in/archives/v7i5/21051905.pdf.
- 45. Mudgill M, Kwatra2 B. Mosquito Menace Aim: Observing the life cycle of Aedes aegypti mosquito and understanding its behavior towards different natural oils for encouraging natural methods of repellence. Int J Sci Res [Internet], 2019; 8(1): 1314–5. Available from: https://www.ijsr.net/get_abstract.php?paper_id=ART20194593.
- 46. Kwatra B, Mudgil M. Untangling the Mathematical Relation Between Natural Selection and Adaptive Radiation Using Macromolecules and Microevolutionary Forces. Int J Sci Res Sci Technol IJSRST | [Internet], 2020; 7(2): 313–39. Available from: http://ijsrst.com/IJSRST207260
- 47. Kwatra B. MECHANISMS OF PATTERN FORMATION OF FBP17 IN MAST CELLS. Int J Adv Res [Internet], 2019 Apr 30; 7(4): 413–4. Available from: http://www.journalijar.com/article/27536/mechanisms-of-pattern-formation-of-fbp17-in-mast-cells/
- 48. Chaitanya Arora BK. Mathematical and Statistical Approach to Define Past Present Future Events. Int J Sci Res [Internet], 2019; 8(4): 261–3. Available from: https://www.researchgate.net/publication/332594418_Mathematical_and_Statistical_Approach_to_Define_Past_Present_Future_Events
- 49. Kwatra B. Maprovit 3, 6, 9: Perfect Companion of your Immune System to Fight Corona Virus Hit. Int J Sci Res [Internet], 2020; 9(4): 241–241. Available from: https://www.ijsr.net/get abstract.php?paper id=SR20406111953.

- 50. Kwatra B. Holothuroidea (Sea Cucumber): Key to Anti-Aging. Int J Sci Res [Internet], 2019; 8(6): 884–884. Available from: https://www.ijsr.net/get_abstract.php?paper_id=6061901
- 51. Kwatra B. HACKING THE BLOOD-BRAIN BARRIER. Eur J Biol Med Sci Res [Internet], 2017; 5(4): 10–3. Available from: https://www.eajournals.org/journals/european-journal-of-biology-and-medical-science-research-ejbmsr/vol-5-issue-4-june-2017/hacking-blood-brain-barrier/
- 52. Kwatra B. EXPRESSION AND CHARACTERIZATION IN PICHIA PASTORIS BY CLONING OF DELTA 4 DESATURASE FROM ISOCHRYSIS GALBANA. Indian J Appl Res [Internet], 2019; 9(5): 1–2. Available from: https://www.researchgate.net/publication/332798347_EXPRESSION_AND_CHARACTERIZ ATION_IN_PICHIA_PASTORIS_BY_CLONING_OF_DELTA_4_DESATURASE_FR OM_ISOCHRYSIS_GALBANA
- 53. Kwatra B. A Review on Potential Properties and Therapeutic Applications of DHA and EPA. ijppr.humanjournals [Internet], 2019; 16(4): 140–76. Available from: https://www.researchgate.net/publication/337745268_A_Review_on_Potential_Properties_and _Therapeutic_Applications_of_DHA_and_EPA
- 54. Kwatra B. CALCIUM AND IRON ABSORPTION: INVITRO STUDIES. Int J Med Biomed Stud [Internet], 2019 Dec 14; 3(12): 59–61. Available from: https://www.researchgate.net/publication/337943738_CALCIUM_AND_IRON_ABSORPTION_INVITRO_STUDIES
- 55. Kwatra B. A REVIEW ON POTENTIAL PROPERTIES AND THERAPEUTIC APPLICATIONS OF BROMELAIN. www.wjpps.com [Internet], 2019; 8(11): 488–500. Available from: https://www.researchgate.net/publication/337745347_A_REVIEW_ON_POTENTIAL_PROPERTIES_AND_THERAPEUTIC_APPLICATIONS_OF_BROMELAIN
- 56. Kwatra B. Bioactive-Compounds: alternative to control Candida spp. Int J Sci Res Rev [Internet], 2019 [cited 2015 Mar 8]; 8(3): 221–3. Available from: https://www.ijsrr.org/down_83045.php
- 57. Kwatra B. Allicin-An After Digestion Antimicrobial Agent. ACTA Sci Microbiol [Internet], 2019; 2(5): 48–51. Available from: https://actascientific.com/ASMI/pdf/ASMI-02-0213.pdf
- 58. Kwatra B. Tinospora Crispa As A Future Cure For Obesity/Cholesterol. Int J Sci Technol Res [Internet], 2017; 6(7): 340–1. Available from: https://www.ijstr.org/final-

- print/july2017/Tinospora-Crispa-As-A-Future-Cure-For-Obesitycholesterol.pdf
- 59. Kwatra B. LOCATOR THEORY FOR ELE-MENTS IN PERIODIC TABLE "LEPT." Glob J Pure Appl Chem Res [Internet], 2017; 5(1): 9–10. Available from: https://www.eajournals.org/journals/global-journal-of-pure-and-applied-chemistry-research-gjpacr/vol-5-issue-1-march-2017/locator-theory-ele-ments-periodic-table-lept/(1)(2)(1,2,11–20,3,21–24,4–10)
- 60. Jayaprakasha G.K., Singh R.P., Sakariah K.K. Antioxidant activity of grape seed (Vitis vinifera) extracts on peroxidation models in vitro. Food Chem, 2001; 73: 285–290. doi: 10.1016/S0308-8146(00)00298-3. [CrossRef] [Google Scholar]
- 61. Jayaprakasha G.K., Selvi T., Sakariah K.K. Antibacterial and antioxidant activities of grape (Vitis vinifera) seed extracts. Food Res. Int, 2003; 36: 117–122. doi: 10.1016/S0963-9969(02)00116-3. [CrossRef] [Google Scholar]
- 62. Puiggròs F., Llópiz N., Ardévol A., Bladé C., Arola L., Salvadó M.J. Grape seed procyanidins prevent oxidative injury by modulating the expression of antioxidant enzyme systems. J. Agric. Food Chem, 2005; 53: 6080–6086. doi: 10.1021/jf050343m. [PubMed] [CrossRef] [Google Scholar]
- 63. Guo L., Wang L.H., Sun B., Yang J.Y., Zhao Y.Q., Dong Y.X., Spranger M.I., Wu C.F. Direct in vivo evidence of protective effects of grape seed procyanidin fractions and other antioxidants against ethanol-induced oxidative DNA damage in mouse brain cells. J. Agric. Food Chem, 2007; 55: 5881–5891. doi: 10.1021/jf070440a. [PubMed] [CrossRef] [Google Scholar]
- 64. Vinson J.A., Mandarano M.A., Shuta D.L., Bagchi M., Bagchi D. Beneficial effects of a novel IH636 grape seed proanthocyanidin extract and a niacin-bound chromium in a hamster atherosclerosis model. Mol. Cell. Biochem, 2002; 240: 99–103. doi: 10.1023/A: 1020611925819. [PubMed] [CrossRef] [Google Scholar]
- 65. Preuss H.G., Wallerstedt D., Talpur N., Tutuncuoglu S.O., Echard B., Myers A., Bui M., Bagchi D. Effects of niacin-bound chromium and grape seed proanthocyanidin extract on the lipid profile of hypercholesterolemic subjects: A pilot study. J. Med, 2000; 31: 227–246. [PubMed] [Google Scholar]
- 66. Olas B., Wachowicz B., Tomczak A., Erler J., Stochmal A., Oleszek W. Comparative anti-platelet and antioxidant properties of polyphenol-rich extracts from: Berries of Aronia melanocarpa, seeds of grape and bark of Yucca schidigera in vitro. Platelets, 2008; 19: 70–77. doi: 10.1080/09537100701708506. [PubMed] [CrossRef] [Google Scholar]

- 67. Cetin A., Kaynar L., Kocyigit I., Hacioglu S.K., Saraymen R., Ozturk A., Sari I., Sagdic O. Role of grape seed extract on methotrexate induced oxidative stress in rat liver. Am. J. Chin. Med, 2008; 36: 861–872. doi: 10.1142/S0192415X08006302. [PubMed] [CrossRef] [Google Scholar]
- 68. Seigers R., Loos M., Van Tellingen O., Boogerd W., Smit A.B., Schagen S.B. Neurobiological changes by cytotoxic agents in mice. Behav. Brain Res, 2016; 299: 19–26. doi: 10.1016/j.bbr.2015.10.057. [PubMed] [CrossRef] [Google Scholar]
- 69. Sano A., Uchida R., Saito M., Shioya N., Komori Y., Tho Y., Hashizume N. Beneficial effects of grape seed extract on malondialdehyde-modified LDL. J. Nutr. Sci. Vitaminol, 2007; 53: 174–182. doi: 10.3177/jnsv.53.174. [PubMed] [CrossRef] [Google Scholar]
- 70. Amaki T., Suzuki T., Nakamura F., Hayashi D., Imai Y., Morita H., Fukino K., Nojiri T., Kitano S., Hibi N., et al. Circulating malondialdehyde modified LDL is a biochemical risk marker for coronary artery disease. Heart, 2004; 90: 1211–1213. doi: 10.1136/hrt.2003.018226. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 71. Natella F., Belelli F., Gentili V., Ursini F., Scaccini C. Grape seed proanthocyanidins prevent plasma postprandial oxidative stress in humans. J. Agric. Food Chem, 2002; 50: 7720–7725. doi: 10.1021/jf0203460. [PubMed] [CrossRef] [Google Scholar]
- 72. Aviram M., Rosenblat M., Bisgaier C.L., Newton R.S., Primo-Parmo S.L., La Du B.N. Paraoxonase inhibits high-density lipoprotein oxidation and preserves its functions. A possible peroxidative role for paraoxonase. J. Clin. Investig, 1998; 101: 1581–1590. doi: 10.1172/JCI1649. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 73. Terra X., Montagut G., Bustos M., Llopiz N., Ardevol A., Blade C., Fernandez-Larrea J., Pujadas G., Salvado J., Arola L., et al. Grape-seed procyanidins prevent low-grade inflammation by modulating cytokine expression in rats fed a high-fat diet. J. Nutr. Biochem, 2009; 20: 210–218. doi: 10.1016/j.jnutbio.2008.02.005. [PubMed] [CrossRef] [Google Scholar]
- 74. Dandona P., Aljada A., Bandyopadhyay A. Inflammation: The link between insulin resistance, obesity and diabetes. Trends Immunol, 2004; 25: 4–7. doi: 10.1016/j.it.2003.10.013. [PubMed] [CrossRef] [Google Scholar]
- 75. Chacon M.R., Ceperuelo-Mallafre V., Maymo-Masip E., Mateo-Sanz J.M., Arola L., Guitierrez C., Fernandez-Real J.M., Ardevol A., Simon I., Vendrell J. Grape-seed procyanidins modulate inflammation on human differentiated adipocytes in vitro. Cytokine, 2009; 47: 137–142. doi: 10.1016/j.cyto.2009.06.001. [PubMed] [CrossRef] [Google Scholar]

- 76. Terra X., Pallarés V., Ardèvol A., Bladé C., Fernández-Larrea J., Pujadas G., Salvadó J., Arola L., Blay M. Modulatory effect of grape-seed procyanidins on local and systemic inflammation in diet-induced obesity rats. J. Nutr. Biochem, 2011; 22: 380–387. doi: 10.1016/j.jnutbio.2010.03.006. [PubMed] [CrossRef] [Google Scholar]
- 77. Pinent M., Blay M., Bladé M.C., Salvadó M.J., Arola L., Ardévol A. Grape seed-derived procyanidins have an antihyperglycemic effect in streptozotocin-induced diabetic rats and insulinomimetic activity in insulin-sensitive cell lines. Endocrinology, 2004; 145: 4985–4990. doi: 10.1210/en.2004-0764. [PubMed] [CrossRef] [Google Scholar]
- 78. Balu M., Sangeetha P., Murali G., Panneerselvam C. Modulatory role of grape seed extract on age-related oxidative DNA damage in central nervous system of rats. Brain Res. Bull, 2006; 68: 469–473. doi: 10.1016/j.brainresbull.2005.10.007. [PubMed] [CrossRef] [Google Scholar]
- 79. Balu M., Sangeetha P., Haripriya D., Panneerselvam C. Rejuvenation of antioxidant system in central nervous system of aged rats by grape seed extract. Neurosci. Lett, 2005; 383: 295–300. doi: 10.1016/j.neulet.2005.04.042. [PubMed] [CrossRef] [Google Scholar]
- 80. Baydar N.G., Sagdic O., Ozkan G., Cetin S. Determination of antibacterial effects and total phenolic contents of grape (Vitis vinifera L.) seed extracts. Int. J. Food Sci. Technol, 2006; 41: 799–804. doi: 10.1111/j.1365-2621.2005.01095.x. [CrossRef] [Google Scholar]
- 81. Brown J.C., Huang G., Haley-Zitlin V., Jiang X. Antibacterial effects of grape extracts on Helicobacter pylori. Appl. Environ. Microbiol, 2009; 75: 848–852. doi: 10.1128/AEM.01595-08. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 82. Ma Z.F., Majid N.A., Yamaoka Y., Lee Y.Y. Food allergy and Helicobacter pylori infection: A systematic review. Front. Microbial, 2016; 7: 368. doi: 10.3389/fmicb.2016.00368. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 83. Kaur M., Singh R.P., Gu M., Agarwal R., Agarwal C. Grape seed extract inhibits in vitro and in vivo growth of human colorectal carcinoma cells. Clin. Cancer Res, 2006; 20: 6194–6202. doi: 10.1158/1078-0432.CCR-06-1465. [PubMed] [CrossRef] [Google Scholar]
- 84. Sun Q., Prasad R., Rosenthal E., Kativar S.K. Grape seed proanthocyanidins inhibit the invasive potential of head and neck cutaneous squamous cell carcinoma cells by targeting EGFR expression and epithelial-to-mesenchymal transition. BMC Complement. Altern. Med, 2011; 11: 134. doi: 10.1186/1472-6882-11-134. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

- 85. Park S.Y., Lee Y.H., Choi K.C., Seong A.R., Choi H.K., Lee O.H., Hwang H.J., Yoon H.G. Grape seed extract regulates androgen receptor-mediated transcription in prostate cancer cells through potent anti-histone acetyltransferase activity. J. Med. Food, 2011; 14: 9–16. doi: 10.1089/jmf.2010.1264. [PubMed] [CrossRef] [Google Scholar]
- 86. Feng Y., Liu Y.M., Fratkins J.D., LeBlanc M.H. Grape seed extract suppresses lipid peroxidation and reduces hypoxic ischemic brain injury in neonatal rats. Brain Res. Bull, 2005; 66: 120–127. doi: 10.1016/j.brainresbull.2005.04.006. [PubMed] [CrossRef] [Google Scholar]
- 87. Perumalla A.V.S., Hettiarachchy N.S. Green tea and grape seed extracts—Potential applications in food safety and quality. Food Res. Int, 2011; 44: 827–839. doi: 10.1016/j.foodres.2011.01.022. [CrossRef] [Google Scholar]
- 88. Bentivegna S.S., Whitney K.M. Subchronic 3-month oral toxicity study of grape seed and grape skin extracts. Food Chem. Toxicol, 2002; 40: 1731–1743. doi: 10.1016/S0278-6915(02)00155-2. [PubMed] [CrossRef] [Google Scholar]
- 89. Taghvaei M., Jafari S.M. Application and stability of natural antioxidants in edible oils in order to substitute synthetic additives. J. Food Sci. Technol, 2015; 52: 1272–1282. doi: 10.1007/s13197-013-1080-1. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 90. Ahn J., Grün I.U., Fernando L.N. Antioxidant properties of natural plant extracts containing polyphenolic compounds in cooked ground beef. J. Food Sci, 2002; 67: 1364–1369. doi: 10.1111/j.1365-2621.2002.tb10290.x. [CrossRef] [Google Scholar]
- 91. Lau D.W., King A.J. Pre- and post-mortem use of grape seed extract in dark poultry meat to inhibit development of thiobarbituric acid reactive substances. J. Agric. Food Chem. 2003; 51: 1602–1607. doi: 10.1021/jf020740m. [PubMed] [CrossRef] [Google Scholar]
- 92. Mielnik M.B., Olsen E., Vogt G., Adeline D., Skrede G. Grape seed extract as antioxidant in cooked, cold stored turkey meat. Food Sci. Technol, 2006; 39: 191–198. doi: 10.1016/j.lwt.2005.02.003. [CrossRef] [Google Scholar]
- 93. Brannan R.G. Effect of grape seed extract on descriptive sensory analysis of ground chicken during refrigerated storage. Meat Sci, 2009; 81: 589–595. doi: 10.1016/j.meatsci.2008.10.014. [PubMed] [CrossRef] [Google Scholar]
- 94. Brannan R.G., Mah E. Grape seed extract inhibits lipid oxidation in muscle from different species during refrigerated and frozen storage and oxidation catalyzed by peroxynitrite and iron/ascorbate in a pyrogallol red model system. Meat Sci, 2007; 77: 540–546. doi: 10.1016/j.meatsci.2007.05.001. [PubMed] [CrossRef] [Google Scholar]

- 95. Roychowdhury S., Wolf G., Keilhoff G., Bagchi D., Horn T. Protection of primary glial cells by grape seed proanthocyanidin extract against nitrosative/oxidative stress. Nitric Oxide, 2001; 5: 137–149. doi: 10.1006/niox.2001.0335. [PubMed] [CrossRef] [Google Scholar]
- 96. Bagchi D., Bagchi M., Stohs S.J., Das D.K., Ray S.D., Kuszynski C.A., Joshi S.S., Pruess H.G. Free radicals and grape seed proanthocyanidin extract: Importance in human health and disease prevention. Toxicology, 2000; 148: 187–197. doi: 10.1016/S0300-483X(00)00210-9. [PubMed] [CrossRef] [Google Scholar]
- 97. Kim S.Y., Jeong S.M., Park W.P., Nam K.C., Ahn D.U., Lee S.C. Effect of heating conditions of grape seeds on the antioxidant activity of grape seed extracts. Food Chem, 2006; 97: 472–479. doi: 10.1016/j.foodchem.2005.05.027. [CrossRef] [Google Scholar]
- 98. Allemann I.B., Baumann L. Antioxidants used in skin care formulations. Skin Therapy Lett, 2008; 13: 5–9. [PubMed] [Google Scholar]
- 99. Grosso G., Micek A., Godos J., Pajak A., Sciacca S., Galvano F., Giovannucci E.L. Dietary flavonoid and lignan intake and mortality in prospective cohort studies: Systematic review and dose-response meta-analysis. Am. J. Epidemiol, 2017 doi: 10.1093/aje/kww207. [PubMed] [CrossRef] [Google Scholar]
- 100. Menezes R., Rodriguez-Mateos A., Kaltsatou A., González-Sarrías A., Greyling A., Giannaki C., Andres-Lacueva C., Milenkovic D., Gibney E.R., Dumont J., et al. Impact of flavonols on cardiometabolic biomarkers: A meta-analysis of randomized controlled human trials to explore the role of inter-individual variability. Nutrients, 2017; 9: 117. doi: 10.3390/nu9020117. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
- 101. Wang X., Ouyang Y.Y., Liu J., Zhao G. Flavonoid intake and risk of CVD: A systematic review and meta-analysis of prospective cohort studies. Br. J. Nutr, 2014; 111: 1–11. doi: 10.1017/S000711451300278X. [PubMed] [CrossRef] [Google Scholar]
- 102. Liu Y.J., Zhan J., Liu X.L., Wang Y., Ji J., He Q.Q. Dietary flavonoids intake and risk of type 2 diabetes: A meta-analysis of prospective cohort studies. Clin. Nutr, 2014; 33: 59–63. doi: 10.1016/j.clnu.2013.03.011. [PubMed] [CrossRef] [Google Scholar]