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# PHYTOSOMES: A NOVEL HERBAL DRUG DELIVERY CARRIER FOR VARIOUS TREATMENTS

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# **ABSTRACT**

Novel drug delivery system called as phytosomes was developed to standardized incorporate plant extract or water-soluble phytoconstituents into phospholipids to produce lipid compatible molecular complexes. Phytosomes is made up of two words and expressed the meaning of "Phyto" means plant and "some" means celllike. There is and intermolecular bonding between the lipids used and the polyphenol. Formulations which are administered orally or topically, they have some limitations in bioavailability. For the enhancement of bioavailability of herbal extracts, this emerging technology is applied for various applications for orally as well as topically. These are the herbosomes that can travel from a hydrophilic

to the lipid environment of the cell membrane and lastly to the blood. Herbal extract in form of phytosomes is generally bioavailable than the original extract due to enhancement in absorption and crossing lipid-rich biomembrane. For phytosomes mainly flavonoids are used as bioactive constituents as they show poor bioavailability. There is a dual advantage of phytosomes for topical pharmaceutical agents and cosmetics with improved efficacy and it can be used effectively as functional cosmetics. Phytosome is a connection between conventional drug delivery and novel drug delivery systems. The present review represents the recent advances and applications of herbal extract phytosome as a tool for drug delivery for various treatments.

**KEYWORDS:** phytosome, phytoconstituent, phosphatidylcholine, bioavailability, antiinflammatory, flavonoid.

# INTRODUCTION

The major drawback of plant extracts is their inability to properly cross the lipid membrane and deliver the drug at specific site of action. [1] Herbal plant extracts have reported therapeutic benefit. There always exhibit limited clinical utility as the synergistic effect of various natural ingredients after individual extraction of compounds. [2] For improvement of bioavailability, herbal products must have proper homeostasis between hydrophilic (for absorption into gastrointestinal tract fluid) and lipophilic (to cross lipid bio membrane balance). [3] The biologically active constituents of plants are mostly polar or water-soluble molecules. Toxicity and absorption problem limit the use of these constituents. [4] Due to their large molecular size, which cannot be absorbed by passive diffusion or due to their poor lipid solubility, water-soluble phytoconstituents like flavonoids, tannins, glycosidal aglycones etc are poorly absorbed thus severely limiting their ability to transport across lipid-rich biological membranes, resulting in their poor bioavailability. [5] In this novel drug delivery technology control of the distribution of drug, is achieved by incorporating the drug (plant actives) in carrier system or in changing the structure of the drug at molecular level. This mechanism aids in increasing solubility, stability, protection from toxicity, pharmacological activity, improved tissue macrophage distribution and sustained delivery. [6] The term phyto means "plant" while "some" means cell like. This are advanced forms of herbal formulation that contains bioactive phytoconstituents of herbal extract surrounded by a lipid. [7] Phytosomes show better physical stability, due to the creation of an H- bond between phospholipids and the phytoconstituents enhancing absorption of hydrophilic polar phytoconstituents resulting in enhanced bioavailability and greater therapeutic benefits. [8] Phosphatidylcholine, phosphatidylserine, phosphatidylethanolamine, phosphatidylinositol are the phospholipids used, but phosphatidylcholine are widely used because of their certain therapeutic value in case of liver diseases, alcoholic steatosis, drug induced liver damage and hepatitis. [9] These are better absorbed, utilized, and have better results. It has higher bioavailability than conventional drug. There are increased pharmacokinetic and pharmacodynamic properties. [10] The current review highlights the future scope and emerging technologies in the field of NDDS for the benefit of herbal and traditional medicines prepared from plant origins.<sup>[11]</sup>

# PHYTOSOME STRUCTURE

These phyto-complex can be considered as a novel entity by reacting phospholipid (either of natural or synthetic origin) with selected botanical constituents with an appropriate solvent, and due to their physical and chemical efficiency.<sup>[12]</sup> The choline part attached with

hydrophilic chief active constituents as shown in Fig. 1.where the phosphatidyl part lipid soluble compound attached with choline bound complex. It results in the formation of lipid complex with better stability and bioavailability.<sup>[11]</sup> One class of phytomedicines currently receiving increased scrutiny is the polyphenols used as extract. These number in the thousands and include, but are not limited to, the various flavonoid subclasses. But many polyphenols are very poorly absorbed when taken orally, posing the greatest obstacle to routine clinical application.<sup>[5]</sup>

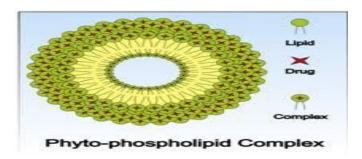


Fig. 1: Diagrammatic representation of a phytosome. It shows the complex formation between the phospholipid and the phytochemical extract. It also involves chemical bond (hydrogen bond).

# DIFFERENCE BETWEEN THE LIPOSOMES AND PHYTOSOMES

The active principle is dissolved in the medium contained in the cavity or in the layers of the membrane in liposome, whereas in the phytosome it is an integral part of the membrane, being the molecules anchored through chemical bonds to the polar head of the phospholipids. Several hundred phospholipid molecules are involved in liposomes for the entrapment and are usually now being used for cosmetic purposes. There is an interaction of 1- 4 phospholipid molecules with the phytoconstituents which are chemically anchored to each other in phytosome formation. This difference results in phytosomes being much better absorbed that liposomes as shown in Fig. 3.Phytosomes are superior to liposomes in skin care products. Comparison between phytosomes and liposomes is represented.

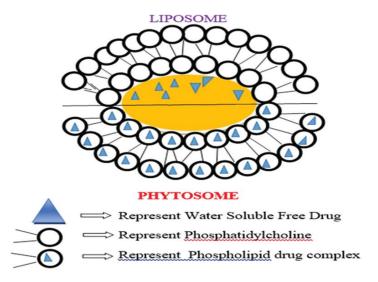


Fig. 2. Schematic representation between liposome (upper segment) and Phytosome (lower segment). In phytosomes the molecules are bonded together having chemical bond. While the liposome is an aggregate of many phospholipid molecules that can enclose other phytoactive molecules but without specifically bonding to them and having no chemical bond.

# DRUG-PHOSPHOLIPID COMPLEX

By reacting with the standardized plant extract and a synthetic or natural phospholipid in a ratio ranging from 0.5-2.0 but usually 1:1 ratio is preferable phytosomes are prepared. Phospholipid is selected from group in which acyl group may be same or different and mostly derived from palmitic, stearic, oleic and linoleic acid like soya lecithin, from bovine or swine brain or dermis, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine. [19-23] The phytosomes are prepared by dissolving the phospholipid such as phosphatidylcholine, phosphatidyl ethanolamine, or phosphatidylserine and the herbal extract in the aprotic solvent such as methylene chloride, dioxane, and ethyl acetate. [24] After solubilisation, this mixture is concentrated to ensure bonding of reactants. Complex thus formed is isolated by solvent removal under vacuum, by lyophilisation or by precipitation with non-solvents. [22,25]

#### Phytochemicals for Phytosomes

Crucial candidates of phytosome are usually terpenes, edusmocoides, ginsenoside, flavonoids, epigallocatechi-3-o-gallate, procyanidins, flavones polyphenols. Phytochemicals are defined as the substances found in edible fruits and vegetables that exhibit a potential for modulating human metabolism in a manner beneficial for the prevention of chronic and degenerative diseases.<sup>[26]</sup> Health promoting effects of flavonoids are reported by many

researchers. The scavenging of oxygen derived free radicals is an important effect of flavonoid. In vitro experimental systems also showed that flavonoids possess anti-inflammatory, antioxidant, anti-viral, and anti-carcinogenic properties. <sup>[27]</sup> This article includes some examples of medicinal plants in Table. 1 with different families and parts of plants containing flavonoid. <sup>[28]</sup>

Table 1: List of medicinal plants having the phytoconstituent as a flavonoid.

<b>Botanical Name</b>	Family	Common name	Plant type	Part used
Adhatoda vasica Nees	Acanthaceae	Adulsa	Shrub	Leaf
Catharanthus roseus Linn.	Apocynaceae	Sadafuli	Herb	Leaf and Root
Phyllanthus emblica Linn.	Euphorbiaceae	Awala	Tree	Fruit and Bark
Coriandrum salivum Linn.	Umbelliferae	Dhaniya	Herb	Leaf and Flower

# Solvents

Different solvents have been utilized by different researchers as the reaction medium for formulating phyto-phospholipid complexes. Aprotic solvents, such as aromatic hydrocarbons, halogen derivatives, methylene chloride, ethyl acetate, or cyclic ethers have been used to prepare phyto- phospholipid complexes but they have been largely replaced by protic solvents like ethanol and methanol as they have been successfully used for complex formation. [29,30]

# Phospholipid

A human biological membrane constitutes different classes of phospholipids, like phosphatidylethanolamine (PE), phosphatidylinositol (PI), phosphatidylcholine (PC), phosphatidic acid (PA), and phosphatidylserine (PS). [31] Phosphatidylcholine is an effective emulsifier because it consists of head or hydrophilic moiety choline (serine) and a tail or lipophilic moiety phosphatidyl. Hence phosphatidylcholine produce a lipid compatible phytoconstituents.<sup>[32]</sup> with In phytosome molecular complex preparation, the phosphatidylcholine having a great role in biological membrane and also act as hepatoprotective(18). The structural representation is shown in Fig. 3. PC molecules exhibit hepatoprotective activities, and have been reported to show clinical effects in the treatment of liver diseases, such as hepatitis, fatty liver, and hepatocirrhosis. [33,34]

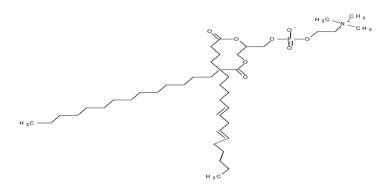


Fig. 3. Molecular structure of phosphatidylcholine. This phospholipid is composed of a choline head group and glycerol phosphoric acid, with a variety of fatty acids. Usually, one is a saturated fatty acid (in the given figure, this is palmitic acid (hexadecanoic acid,  $H_3C$ -( $CH_2$ )<sub>14</sub>-COOH; margaric acid (heptadecanoic acid,  $H_3C$ -( $CH_2$ )<sub>15</sub>-COOH).

# PREPARATION OF EXTRACT OF VARIOUS PARTS OF PLANTS

#### Extract of flowers

The flowers were washed, air dried, homogenized to fine powder and stored in airtight bottles for future use. 30g of dried powder was first defatted with petroleum ether and then extracted by soxhlation using 80% methanol. The extract was concentrated in a rotatory vaccum evaporator and further dried in desiccator till use.<sup>[35]</sup>

# Extract of leaves

The leaves of plant were air-dried until dryness at room temperature and under shade. The dried leaves were then powdered to a fine grade by using laboratory scale mill. Further it was sequentially extracted successively with ethanol using soxhlet apparatus. The solvent was removed and concentrated in a rotary evaporator and water bath. The dried extracts were stored in refrigerator for further studies. [36,37]

# • Extract of seeds

Seeds were finely powdered to yield flour, then for example 100 g of flour was accurately weighed and soaked in 1 L of methanol at room temperature (25°C) with continuous agitation for 24 hours using a magnetic stirrer. The obtained extract solution was filtered and methanol was evaporated using a rotary evaporator at 30°C and 120 rpm. After complete solvent evaporation, the yielded dry extract was stored at 4°C until further use. [38]

# PROPERTIES OF PHYTOSOMES

# Physicochemical properties

Phytosome, a product is obtained from the stoichiometric reaction between phospholipid and plant extracts. <sup>[39]</sup> Cell like structure is observed when in contact with hydrophilic environment in phytosomes, but in a liposome the chief constituent interacts within the internal pocket while in phytosome the chief active constituents are enveloped the polar head of phospholipid and becoming an integral part of the membrane. <sup>[2]</sup> Solubility of Phytosome varies as: soluble in aprotic solvents, moderately soluble in fats, insoluble in water and relatively unstable in alcohol. <sup>[40,41]</sup> It can be deduced that the fatty chain gives unchanged signals both in free phospholipid and in the complex from the 1HNMR and 13CNMR data, which indicates that long aliphatic chains are wrapped around the active principle, producing lipophilic envelope. <sup>[42]</sup> Phytosome size varies from 50 nm - 100 μm. <sup>[8]</sup>

# • Biological properties

Phytosomes shows better result than the conventional herbal extract or non-complexed extracts as they absorbed and utilized better, hence they produce more bioavailability, which has been demonstrated by pharmacokinetic studies or by pharmacodynamic tests in experimental animals and inhuman subjects. Phytosomes improve absorption of phytoconstituents through skin, to regulate the physiology of skin compositions. The improvement in the functioning of skin suggests the functional importance of the phytosomes. [43]

# MECHANISM OF PHYTOSOME TECHNOLOGY

Phytosomes technology is mainly result with complexation of polyphenols with phospholipid in 1:1 ratio or 1:2 results in the formation of phytosomal complex with lipid covering around the constituents. [44] Molecules are anchored through chemical bonds to the polar choline head of the phospholipid. Precise chemical analysis indicates the unit phytosome is usually a flavonoid or polyphenol molecule linked with at least one phosphatidylcholine molecule. The result is a little microsphere or cell is produced. [42] Phospholipid complexes may be absorbed from the GIT through enterocyte based transport, and drug transport to the systemic circulation as shown in Fig. 4 via intestinal lymphatic system which has widespread network throughout the body. The major advantage of lymphatic transport is to bypass the first-pass metabolism and applicable for targeted drug delivery. [45]

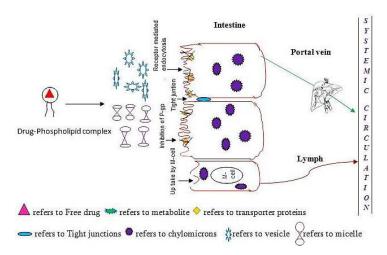


Fig. 4. Representation of mechanism of phospholipid complex. Phytosomes enhances the absorption of lipid insoluble polar phytoconstituents through oral as well as topical route showing better bioavailability, hence significantly greater therapeutic benefit. Absorption takes place towards systemic circulation via lymphatic system as it bypass the first pass metabolism.

# **ADVANTAGES**

- Due to their more skin penetration and high lipid profile phytosomes are widely used in cosmetic preparation.
- As the absorption of chief phytoconstituent is improved, its dose requirement is also reduced.<sup>[25]</sup>
- Herbal phytosome process produces a little cell whereby the valuable components of the herbal extracts are protected from destruction by digestive secretions and gut bacteria.
- in the phytosome process phosphatiylcholine is used which acting as a carrier and also nourishes the skin, as it is the essential part of cell membrane. [11,46]

# **DISADVANTAGES**

 Although Phytosome having so many advantages but instead of that this technology has some disadvantages like rapidly elimination of phytoconstituents from the phytosome.<sup>[47]</sup>

# METHODS OF PREPARATION

# Solvent evaporation method

The specific amount of herbal extract and soya lecithin were taken into a 100 ml round bottom flask and refluxed with 20 ml of acetone at a temperature 50 - 60°C for 2 h. The mixture is concentrated to 5-10 ml to obtain the precipitate which was filtered and collected.

The dried precipitate phytosome complex was placed in amber coloured glass bottle and stored at room temperature.<sup>[29]</sup>

# Rotatory evaporator method

The specific amount of extract e.g. curcumin was prepared by this method and soya lecithin were dissolved in dichloromethane in a rotary round bottom flask followed by stirring for 1 hour at a temperature not exceeding 40°C. Thin film of the sample was obtained to which n-hexane was added and continuously stirred until monolayer of phospholipid and then add phosphate buffer 6.8 and precipitate obtained was collected, placed in amber coloured glass bottle and stored at room temperature.<sup>[48]</sup>

# • Antisolvent precipitation technique

Known quantity of drug, phospholipids and polymer is taken in round bottom flask (RBF) and refluxed with specific solvent not exceeding 600C for 2 hours. The mixture is concentrated to 5-10ml and n –Hexane is carefully added to it with continuous stirring to get the precipitate, which is filtered and kept in vacuum desiccator overnight. The dried precipitate is crushed in mortar and sieved through 100- mesh size. As a result, phytosomes loaded with drug are obtained, which are placed in amber coloured glass bottles at room temperature. [42]

# • Mechanical dispersion method

In this method, the lipids dissolved in organic solvent are brought in contact with aqueous phase containing the drug.<sup>[48]</sup> Initially, pc is dissolved in diethyl ether which is later slowly injected to an aqueous solution of the phytoconstituents to be encapsulated. Phytophospholipid complex formation takes places by subsequent removal of the organic solvent under reduced pressure. Novel methods for the phospholipid complex preparation includes super critical fluids (SCF), which include gas anti-solvent technique (GAS) compressed anti solvent process (PCA), supercritical anti solvent method (SAS).<sup>[49,50]</sup>

# **EVALUATION OF PHYTOSOMES**

#### Visualization

For visualization, transmission electron microscopy (TEM) and by scanning electron microscopy (SEM) can be used. [51]

# • Determination of % yield

% yield of phytosome complex was determined by the following formula:

(%) Yield= (Practical yield) × 100 (Theoretical yield)

# • Determination of drug content

100mg complex is dissolved in 10ml methanol ti determine the drug content. After suitable dilution absorbance was determined by UV Spectrophotometer at 269 nm and drug content was determined.<sup>[52]</sup>

# • Entrapment efficiency

The entrapment efficiency of a phytosomal formulation can be determined by subjecting the formulation to ultracentrifugation technique.<sup>[53]</sup>

# • Transition temperature

Differential scanning calorimetry can be used for the determination of transition temperature of the vesicular lipid systems.<sup>[51]</sup>

# • Spectroscopic evaluations

The interaction between the phytoconstituent molecule and the phospholipid molecule is very important which can be determined with the help of following spectroscopic methods:

### • 1H-NMR

There are several examples whose examples whose NMR spectra can help to determine the interaction between the phospholipid and phytoconstituent. By NMR evaluation a marked change can be seen in 1H-NMR signal of those particular atoms which take part in the complex formation.<sup>[51,54]</sup>

### • 13C-NMR

The signals are broadened and some are shifted, while most of the resonances retain their original sharp line shape of fatty acid chains which are corresponding to the glycerol and choline portion of the lipid (between 60–80 ppm). After heating to 60°, all the signals belonging to the flavonoid moieties reappear, although they are still very broad and partially overlapping.<sup>[55]</sup>

# • Ultraviolet spectra

Samples that reflect different absorption in the UV wave- length range can be used to characterize own structural properties. Most studies have revealed no differences in the UV ab- sorption characteristics of constituents before and after complexation. Xu et al. prepared luteolin-phospholipid complexes and found that the characteristic peaks of luteolin remained present.<sup>[56]</sup>

# Biological evaluation

For evaluation of Phytosome using in-vitro and in-vivo models can be selected based upon the phytoconstituents the therapeutic activity which are present in the formulation. For example, for the assessment of ant-diabetic Phytosome formulation, the blood glucose or sugar is analysed.<sup>[8]</sup>

#### EXAMPLES OF PHYTOSOME FORMULATION

Plant extract or phytoconstituents that are incorporated in the Phytosome technology are reported in the literature but some examples are from them such as Ginkgo biloba, Grape seed, Centella, Green tea, and Ginseng.<sup>[8]</sup> In Table. 2 some commercial phytosomes formulations are listed.<sup>[22,54,57-61]</sup>

**Table 2: Some Examples of Phytosome Formulations.** 

Phytosomes	Phytoconstituents	Indications	
Green Tea	Epigallocatechin from Thea	Nutraceutical, systemic	
PhytosomeTM	sinensis	antioxidant. Anticancer	
Ginseng PhytosomeTM	37.5 % ginsenosides from immunomodulator Panax ginseng	Nutraceutical and immunomodulator	
Ginkgo biloba	24 % Ginkgo flavone	Protects brain and vascular	
Phytosome TM	glycosides from Ginkgo biloba	lining, anti-ageing agent.	
Centella	Tamanas	Used to treat Vein and	
	Terpenes	skin disorders.	
Grape seed	Procyanidins from vitis	Nutraceutical, systemic	
phytosomeTM	vinifera	anticancer, antioxidant	
Curcumin (Merinoselect) Phytosomes	Polyphenol from Curcuma Longa	Cancer chemo preventive agent improving the oral bioavailability and the plasma.	
Glycyrrhiza phytosome	18-beta glycyrrhetinic acid	Anti-inflammatory activity	
Hawthorn phytosome TM	Flavonoids from Crataegus sp.	Nutraceutical, cardio- protective and antihypertensive	

# APPLICATIONS OF PHYTOSOMES

- Ginkgo phytosome, prepared from Ginkgo biloba leaves showed better therapeutic effects in the treatment of Raynaud's disease and intermittent circulationin comparison to the conventional standardized plant extract.<sup>[62]</sup>
- Bacopa monnieri plant having antiamnesic activity has Bacopaside well-known chief
  constituents present in it. To prepare phytosome from bacopaside and its in vivo
  evaluation on rodents is an attempt that needed to be studied. There is remarkably great
  change in the therapeutic efficacy of the compound prepared by phospholipid as compare
  to simple B. monnnieri extract.<sup>[63]</sup>
- Then the single constituent the silymarin phytosomes showed much higher specific activity and a longer lasting action, with respect to percent reduction of edema, inhibition of myeloperoxidase activity, antioxidant and free radical scavenging activity. [64]
- Then the single constituent the silymarin phytosomes showed much higher specific activity and a longer lasting action, with respect to percent reduction of edema, inhibition of myeloperoxidase activity, antioxidant and free radical scavenging activity. [64]
- Merivaselect® phytosomes Curcumin polyphenols, [65] obtained from Curcuma longa family Zingiberaceae, are powerful scavengers of superoxide and hydroxyl free radicals. [66] They also lower the incidence of mutations and genetic disorders by the ability to prevent DNA oxidative damage. [67] Potential effect in cancer chemoprevention, [68] inflammation [69] and neuro-degenerative diseases. [70]
- The quercetin phospholipid phytosomal complex was developed by a simple and reproducible method showed that the formulation exerted better therapeutic efficacy as compared to the non-phytosomal conventional in rat liver injury induced by carbon tetrachloride.<sup>[21]</sup>
- The rutin in its free form it was observed that the Rutin phytosomes were better able to penetrate the impermeable stratum corneum. Skin uptake of Rutin phytosomes was  $33 \pm 1.33\%$  whereas that of Rutin was  $13 \pm 0.87\%$ . [71]
- Later formulated and characterized phytosome suspension of Urtica dioica (UD). These
  findings suggest that herbosome suspension of Urtica dioica(100 mg/kg) shows better
  antidiabetic activity in comparisons to the powdered marketed formulations of Urtica
  dioica.<sup>[72]</sup>

# **CONCLUSION**

Preparation of phytosomes is reproducible and simple. For plant extract a plant will be selected for flavonoid isolation and phytosome utilization for the treatment severe disease condition. The technology shows cost effective delivery of phytoconstituents and synergistic effects as functional cosmetics. Apart from that the phospholipids used have their own beneficial effect to the body. The information gathered herein will be useful for the researchers who wish to explore a vesicular drug delivery system which encompasses effective drug on target site without its metabolism. The formulation methodology for phytosome is simple and can easily be used on commercial scale. Phytosome is very useful in cosmetology also. It has a great future for use in formulation technology and applications of hydrophilic plant compounds as far as the potential of phytosome technology is concerned.

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#### REFERENCES

- 1. Puranik N, Kammar K, Sheeladevi. Anti-diabetic activity of Tinospora cordifolia (Willd.) in streptozotocin diabetic rats; does it act like sulfonylureas? Turkish Journal of Medical Sciences, 2010 Feb 10; 40: 265–70.
- Bhattacharya S, Ghosh AK. Phytosomes: the Emerging Technology for Enhancement of Bioavailability of Botanicals and Nutraceuticals. The Internet Journal of Aesthetic and Antiaging Medicine [Internet]. 2008 Dec 31 [cited 2020 Jun 9]; 2(1). Available from: http://ispub.com/IJAAM/2/1/7656
- 3. Semalty A, Semalty M, Rawat MSM, Franceschi F. Supramolecular phospholipids-polyphenolics interactions: the PHYTOSOME strategy to improve the bioavailability of phytochemicals. Fitoterapia, 2010 Jul; 81(5): 306–14.
- 4. Elliott M, Chithan K, Chithan K. The impact of plant flavonoids on mammalian biology: implications for immunity, inflammation and cancer [Internet]. The Flavonoids Advances in Research Since 1986. Routledge; 2017 [cited 2020 Jun 7]. p. 619–52. Available from: https://www.taylorfrancis.com/
- 5. Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: food sources and bioavailability. Am J Clin Nutr., 2004 May 1; 79(5): 727–47.
- 6. Dhiman A, Nanda A, Ahmad S. Novel Herbal Drug Delivery System (NHDDS): the need of Hour [Internet]. [cited 2020 Jun 13]. Available from: /paper/Novel-Herbal-Drug-

- Delivery-System-(-NHDDS-)-%3A-the-Dhiman-Nanda/683c33dc1eb996aadf65820d39fd32fbcb4150fd
- 7. Mali R, Hundiwale JC, Sonawane RS, Patil RN, Hatapakki BC. Evaluation of Capparis decidua for anthelmintic and antimicrobial activities. Indian J Nat Prod, 2004 Jan 1; 20: 10–3.
- 8. Amit P, Y.S. tanwar, S. Rakesh. Phytosome: Phytolipid Drug Dilivery System for Improving Bioavailability of Herbal Drug, 2013; 3(2): 7.
- 9. Shivanand P, Kinjal P. Phytosomes: Technical Revolution in Phytomedicine, 5.
- 10. Miladi S, Damak M. IN VITRO ANTIOXIDANT ACTIVITIES OF Aloe vera LEAF SKIN EXTRACTS, 2008; 9.
- 11. Kareparamban JA, Nikam PH, Jadhav AP, Kadam VJ, Mumbai N. PHYTOSOME: A NOVEL REVOLUTION IN HERBAL DRUGS [Internet]. 2012 [cited 2020 Jun 8]. Available from: /paper/PHYTOSOME%3A-A-NOVEL-REVOLUTION-IN-HERBAL-DRUGS-Kareparamban-Nikam/388804e6db89f4f7d608ae8aea576e74bb148f2b
- 12. Jain N, Gupta BP, Thakur N, Jain R, Banweer J, Jain DK, et al. PHYTOSOME: A NOVEL DRUG DELIVERY SYSTEM FOR HERBAL MEDICINE. International Journal of Pharmaceutical Sciences and Drug Research, 2010 Oct 1; 224–8.
- 13. BOMBARDELLI E, CRISTONI A, MORAZZONI P. Phytosome®s in functional cosmetics. Fitoterapia (Milano), 1994; 65(5): 387–401.
- 14. Darshan D, Satyaendra S, Shweta K, Gangwal A, Dubey PK. Phytosome: A novel dosage structure. Int J Chem Sci., 2007; 5(1): 132–34.
- 15. Gupta NK, Dixit VK. Development and evaluation of vesicular system for curcumin delivery. Arch Dermatol Res., 2011 Mar; 303(2): 89–101.
- 16. Kidd PM. Bioavailability and activity of phytosome complexes from botanical polyphenols: the silymarin, curcumin, green tea, and grape seed extracts. Altern Med Rev., 2009 Sep; 14(3): 226–46.
- 17. Bhattacharya S. Phytosomes: The New Technology for Enhancement of Bioavailability of Botanicals and Nutraceuticals. International Journal of Health Research, 2009; 2(3): 225–32.
- 18. Ghanbarzadeh B, Babazadeh A, Hamishehkar H. Nano-phytosome as a potential food-grade delivery system. Food Bioscience, 2016 Sep 1; 15: 126–35.
- 19. Marena C, Lampertico M. Preliminary clinical development of silipide: A new complex of silybin in toxic liver disorders. PLANTA MED [Internet]. 1991 [cited 2020 Jun 13]; 57(SUPPL. 2). Available from: https://moh-

- it.pure.elsevier.com/en/publications/preliminary-clinical-development-of-silipide-a-new-complex-of-sil
- 20. Maiti K, Mukherjee K, Gantait A, Saha BP, Mukherjee PK. Curcumin-phospholipid complex: Preparation, therapeutic evaluation and pharmacokinetic study in rats. Int J Pharm, 2007 Feb 7; 330(1–2): 155–63.
- 21. Maiti K, Mukherjee K, Gantait A, Nazeer Ahamed H, Saha BP, Kumar Mukherjee P. Enhanced Therapeutic Benefit of QuercetinPhospholipid Complex in Carbon Tetrachloride-Induced Acute Liver Injury in Rats: A Comparative Study. Iranian Journal of Pharmacology and Therapeutics, 2005 Nov 10; 4(2): 84–0.
- 22. Moscarella S, Giusti A, Marra F, Marena C, Lampertico M, Relli P, et al. Therapeutic and antilipoperoxidant effects of silybin-phosphatidylcholine complex in chronic liver disease: Preliminary results. Current Therapeutic Research, 1993 Jan 1; 53(1): 98–102.
- 23. Xiao Y, Yunmei S, Zhipeng C, Qineng P. The preparation of silybin-phospholipid complex and the study on its pharmacokinetics in rats. International journal of pharmaceutics, 2006 Feb 1; 307: 77–82.
- 24. Kumari P, Singh N, Cheriyan BP. PHYTOSOME: A NOVAL APPROACH FOR PHYTOMEDICINE. In 2011.
- 25. Raju TP, Reddy MS, Reddy VP. PHYTO SOMES: A NOVEL PHYTO PHOSPHOLIPID CARRIERS F OR HERBAL DRUG DELIVERY [Internet]. 2011 [cited 2020 Jun 10]. Available from: /paper/PHYTO-SOMES%3A-A-NOVEL-PHYTO-PHOSPHOLIPID-CARRIERS-F-Raju-Reddy/05751575bb3becc5aecb444ce0aa99501e1c40bd
- 26. Tripoli E, Guardia M, Giammanco S, Di Majo D, Giammanco M. Citrus flavonoids: Molecular structure, biological activity and nutritional properties: A review. Food Chemistry, 2007 Dec 31; 104: 466–79.
- 27. Nijveldt RJ, van Nood E, van Hoorn DE, Boelens PG, van Norren K, van Leeuwen PA. Flavonoids: a review of probable mechanisms of action and potential applications. Am J Clin Nutr., 2001 Oct; 74(4): 418–25.
- 28. Geissman TA, Crout DHG. Organic chemistry of secondary plant metabolism [Internet]. San Francisco: Freeman, Cooper & Co; 1969 [cited 2020 Jun 18]. Available from: https://trove.nla.gov.au/version/28594208
- 29. Khan J, Alexander A, Ajazuddin null, Saraf S, Saraf S. Recent advances and future prospects of phyto-phospholipid complexation technique for improving pharmacokinetic profile of plant actives. J Control Release, 2013 May 28; 168(1): 50–60.

- 30. Shakeri A, Sahebkar A. Phytosome: A Fatty Solution for Efficient Formulation of Phytopharmaceuticals. Recent patents on drug delivery & formulation, 2015 Aug 13; 10.
- 31. Szuhaj BF. Lecithins: sources, manufacture & uses. Champaign, ILL: American Oil Chemists' Society, 1989.
- 32. B AK, Habbu P, T T, L L, Hullatti P, S RK. Phytosomes as Novel Drug Delivery System for Herbal Medicine –A Review. SRP., 2016 Nov 19; 8(1): 5–7.
- 33. Kidd P, Head K. A review of the bioavailability and clinical efficacy of milk thistle phytosome: a silybin-phosphatidylcholine complex (Siliphos). Altern Med Rev., 2005 Sep; 10(3): 193–203.
- 34. Duric M, Sivanesan S, Bakovic M. Phosphatidylcholine functional foods and nutraceuticals: A potential approach to prevent non-alcoholic fatty liver disease. European Journal of Lipid Science and Technology, 2012; 114(4): 389–98.
- 35. Keerthi B, Pingali PS, Srinivas P. Formulation and Evaluation of Capsules of Ashwagandha Phytosomes [Internet]. 2014 [cited 2020 Jun 19]. Available from: /paper/Formulation-and-Evaluation-of-Capsules-of-Keerthi-Pingali/38b56180fe59f1a1cdc10085b03b70359d054e5d
- 36. Thani W, Vallisuta O, Siripong P, Ruangwises N. Anti-proliferative and antioxidative activities of Thai noni/Yor (Morinda citrifolia Linn.) leaf extract. Southeast Asian J Trop Med Public Health, 2010 Mar; 41(2): 482–9.
- 37. Singh RP, Narke R. Preparation and evaluation of phytosome of lawsone. International Journal of Pharmaceutical Sciences and Research, 2015; 6(12): 5217.
- 38. Zhang W, Popovich DG. Chemical and Biological Characterization of Oleanane Triterpenoids from Soy. Molecules, 2009 Aug 10; 14(8): 2959–75.
- 39. Pajardi G, Bortot P, Ponti V, Novelli C. Clinical usefulness of oral supplementation with alpha-lipoic Acid, curcumin phytosome, and B-group vitamins in patients with carpal tunnel syndrome undergoing surgical treatment. Evid Based Complement Alternat Med., 2014; 2014: 891310.
- 40. Maffei Facino R, Carini M, Aldini G, Bombardelli E, Morazzoni P, Morelli R. Free radicals scavenging action and anti-enzyme activities of procyanidines from Vitis vinifera. A mechanism for their capillary protective action. Arzneimittelforschung, 1994 May; 44(5): 592–601.
- 41. Jain N, Gupta BP, Thakur N, Jain R, Banweer J, Jain DK, et al. PHYTOSOME: A NOVEL DRUG DELIVERY SYSTEM FOR HERBAL MEDICINE. International Journal of Pharmaceutical Sciences and Drug Research, 2010 Oct 1; 224–8.

- 42. Bombardelli E, Patri GF. Complex compounds of bioflavonoids with phospholipids, their preparation and use, and pharmaceutical and cosmetic compositions containing them [Internet]. US5043323A, 1991 [cited 2020 Jun 9]. Available from: https://patents.google.com/patent/US5043323A/en
- 43. Ravi G, Chandur VK, Shabaraya AR, Sanjay K. Review Article CODEN: IJPRNK ISSN: 2277-8713 GS Ravi, IJPRBS, 2015; 4(3): 415-432 IJPRBS [Internet]. 2015 [cited 2020 Jun 9]. Available from: /paper/Review-Article-CODEN%3A-IJPRNK-ISSN%3A-2277-8713-GS-Ravi-Chandur/49fa2a7821fe96a86853ac77ec006a192f76035d
- 44. Agrawal VK, Gupta A, Chaturvedi S. IMPROVEMENT IN BIOAVAILABILITY OF CLASS-III DRUG: PHYTOLIPID DELIVERY SYSTEM. In 2012.
- 45. Kalepu S, Manthina M, Padavala V. Oral lipid-based drug delivery systems An overview. Acta Pharmaceutica Sinica B., 2013 Dec 1; 3.
- 46. Monica G, Naik VV. HERBOSOMES: A POTENTIAL CARRIERS FOR THE BIOAVAILABILITY ENHANCEMENT OF HERBAL EXTRACTS. World Journal of Pharmacy and Pharmaceutical Sciences, 2014; 4(01): 28.
- 47. Saha S, Sarma A, Saikia P, Chakrabarty T. Phytosome: A Brief Overview. 10.
- 48. Sikarwar MS, Sharma S, Jain AK, Parial SD. Preparation, characterization and evaluation of Marsupsin-phospholipid complex. AAPS Pharm Sci Tech., 2008; 9(1): 129–37.
- 49. Li Y, Yang D-J, Chen S-L, Chen S-B, Chan AS-C. Comparative physicochemical characterization of phospholipids complex of puerarin formulated by conventional and supercritical methods. Pharm Res., 2008 Mar; 25(3): 563–77.
- 50. Li Y, Yang D-J, Chen S-L, Chen S-B, Chan AS-C. Process parameters and morphology in puerarin, phospholipids and their complex microparticles generation by supercritical antisolvent precipitation. Int J Pharm., 2008 Jul 9; 359(1–2): 35–45.
- 51. Sowjanya J, Kumar Y, Saumya D, Pattanayak D. Phytosome a novel entity in herbal delivery system: a review. International J Pharm Res Dev., 2010 Jan 1; 2.
- 52. Zhang J, Tang Q, Xu X, Li N. Development and evaluation of a novel phytosome-loaded chitosan microsphere system for curcumin delivery. Int J Pharm, 2013 May 1; 448(1): 168–74.
- 53. El Maghraby GM, Williams AC, Barry BW. Oestradiol skin delivery from ultradeformable liposomes: refinement of surfactant concentration. Int J Pharm, 2000 Feb 25; 196(1): 63–74.
- 54. Patel J, Patel R, Khambholja K, Patel N. An overview of phytosomes as an advanced herbal drug delivery system. Asian J Pharm Sci., 2009; 4(6): 363–371.

- 55. Semalty A, Semalty M, Rawat M s. M, R. Singh. Phytosome in herbal drug delivery: a review. Indian Drugs, 2006 Dec 1; 43: 937–46.
- 56. Xu K, Liu B, Ma Y, Du J, Li G, Gao H, et al. Physicochemical properties and antioxidant activities of luteolin-phospholipid complex. Molecules, 2009 Sep 9; 14(9): 3486–93.
- 57. Amin T, Bhat SV. A Review on Phytosome Technology as a Novel Approach to Improve The Bioavailability of Nutraceuticals, 2012; 1: 15.
- 58. Naik SR, Pilgaonkar VW, Panda VS. Evaluation of antioxidant activity of Ginkgo biloba phytosomes in rat brain. Phytotherapy Research, 2006; 20(11): 1013–6.
- 59. Bombardelli E, Curri SB, Loggia RD, Delnegro P, Gariboldi P, Tubaro A. Anti-inflammatory activity of 18-ß-glycyrrhetinic acid in phytosome form. [Internet]. 1989 [cited 2020 Jun 22]. Available from: /paper/Anti-inflammatory-activity-of-18-%C3%9F-glycyrrhetinic-Bombardelli-Curri/689e7985e06651808a9b6991cb4964f3f7d64680
- 60. Karimi N, Ghanbarzadeh B, Hamishehkar H, Keivani F, Pezeshki A, Gholian MM. Phytosome and Liposome: The Beneficial Encapsulation Systems in Drug Delivery and Food Application. Applied Food Biotechnology, 2015 Jun 30; 2(3): 17–27.
- 61. Awasthi R, Kulkarni GT, Pawar VK. PHYTOSOMES: AN APPROACH TO INCREASE THE BIOAVAILABILITY OF PLANT EXTRACTS, 2011; 3(2): 3.
- 62. La Grange L, Wang M, Watkins R, Ortiz D, Sanchez ME, Konst J, et al. Protective effects of the flavonoid mixture, silymarin, on fetal rat brain and liver. Journal of Ethnopharmacology, 1999 Apr 1; 65(1): 53–61.
- 63. Zhang Z, Chen Y, Deng J, Jia X, Zhou J, Lv H. Solid dispersion of berberine-phospholipid complex/TPGS 1000/SiO<sub>2</sub>: preparation, characterization and in vivo studies. Int J Pharm, 2014 Apr 25; 465(1–2): 306–16.
- 64. Bombardelli E, Loggia R della, Sosa S, Spelta M, Tubaro A. Aging skin: protective effect of Silymarin-Phytosome(R) [Internet]. 1991 [cited 2020 Jun 13]. Available from: /paper/Aging-skin%3A-protective-effect-of-Bombardelli-Loggia/af041c3d41d5ae2fc6af8919f4b582669368a319
- 65. Goel A, Kunnumakkara AB, Aggarwal BB. Curcumin as "Curecumin": From kitchen to clinic. Biochemical Pharmacology, 2008 Feb 15; 75(4): 787–809.
- 66. Soni KB, Kuttan R. Effect of oral curcumin administration on serum peroxides and cholesterol levels in human volunteers. Indian J Physiol Pharmacol, 1992 Oct; 36(4): 273–5.
- 67. Garcea G, Berry DP, Jones DJL, Singh R, Dennison AR, Farmer PB, et al. Consumption of the putative chemopreventive agent curcumin by cancer patients: assessment of

- curcumin levels in the colorectum and their pharmacodynamic consequences. Cancer Epidemiol Biomarkers Prev., 2005 Jan; 14(1): 120–5.
- 68. Villegas I, Sánchez-Fidalgo S, Alarcón de la Lastra C. New mechanisms and therapeutic potential of curcumin for colorectal cancer. Mol Nutr Food Res., 2008 Sep; 52(9): 1040–61.
- 69. Strimpakos AS, Sharma RA. Curcumin: preventive and therapeutic properties in laboratory studies and clinical trials. Antioxid Redox Signal, 2008 Mar; 10(3): 511–45.
- 70. Begum AN, Jones MR, Lim GP, Morihara T, Kim P, Heath DD, et al. Curcumin structure-function, bioavailability, and efficacy in models of neuroinflammation and Alzheimer's disease. J Pharmacol Exp Ther., 2008 Jul; 326(1): 196–208.
- 71. Das MK, Kalita B. Design and Evaluation of Phyto-Phospholipid Complexes (Phytosomes) of Rutin for Transdermal Application. J app pharm sci., 2014 Oct 30; 4(10): 51–7.
- 72. Shakya R, Roy AK, Bhattacharya D, Rajesh T, Joshi B, Chettri N. DEVELPOPMENT AND EVALUATION OF HERBOSOME SUSPENSION. 10.