

**SYNTHETIC NUCLEIC ACIDS FROM THYMIDINE AND  
ORGANOTIN DIHALIDES AND THEIR ABILITY TO INHIBIT  
HUMAN CANCER CELL LINES INCLUDING PANCREATIC ACID  
AND GLIOBLASTOMAS CELL LINES**

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### **ABSTRACT**

Using the interfacial polymerization technique, synthetic nucleic acid polymers derived from thymidine and organotin dihalides were formed in moderate yield and chain length. Products were synthesized using the interfacial polycondensation process and commercially available reagents for easy reproducibility. Structural characterization of polymers was carried out using Infrared Spectroscopy (IR), and Matrix-assisted Laser Desorption/Ionization Mass Spectrometry (MALDI-MS). Light scattering Photometry was used to determine chain length. The polymers showed good inhibition of a battery of human cancer cell lines including two breast, two pancreatic, and two glioblastoma brain cancer lines. All polymers tested were able to differentiate between healthy and malignant cells, showing excellent

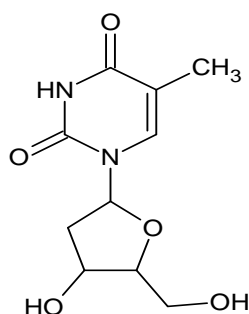
selectivity toward malignant cells.

**KEYWORDS:** thymidine, organotin dihalides, organotin polymers, synthetic nucleic acids, pancreatic cancer, breast cancer, glioblastomas brain cancer, MALDI MS, interfacial polymerization.

## INTRODUCTION

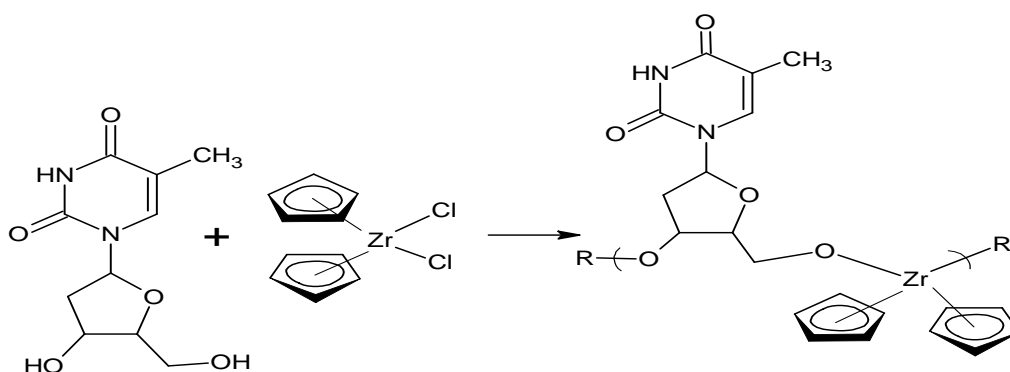
Carraher and Millich were issued the first patent for the chemical synthesis of nucleic acids in 1971.<sup>[1]</sup> The nucleic acid was derived from thymidine and 2-deoxy-D-ribose. This was followed by the synthesis of additional related nucleic acid like polymers,<sup>[2-7]</sup> More recently, we focused on the synthesis metal-containing polymers due to their ability to inhibit a variety of viruses, bacteria, fungi, and cancers. These included a variety of metal-containing polymers including tin,<sup>[8-22]</sup> Group 4 and 5 metallocenes,<sup>[23-30]</sup> platinum and palladium<sup>[31-33]</sup> and Group 15 metals.<sup>[31-40]</sup>

Thymidine, (Figure 1), is one of the four bases present in DNA. Since thymidine has two reactive functional groups, as a difunctional compound with two hydroxyls on the sugar moiety, the di-functionality of this compound prevents crosslinking during polymerization. The other natural nucleic acids contain additional functional groups so complicated the synthesis of linear products.



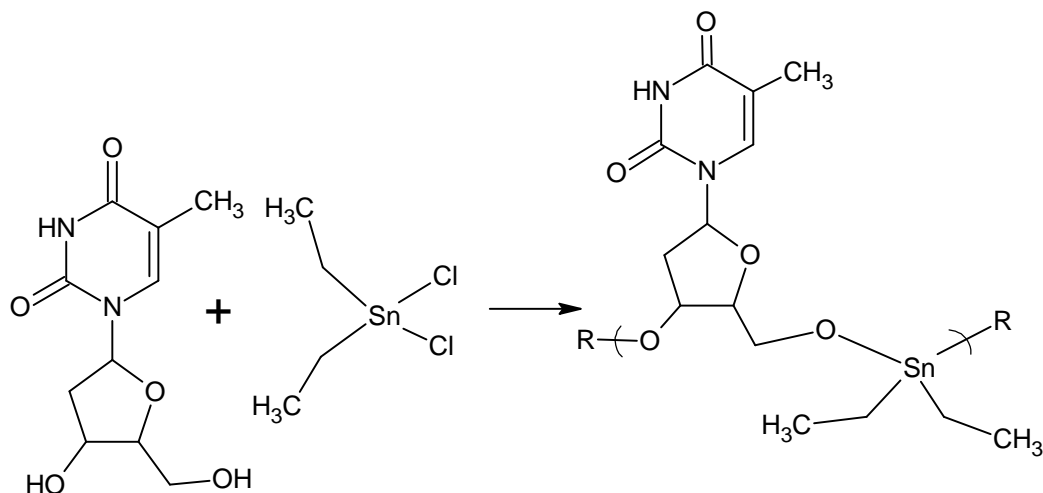
**Figure 01: Thymidine structure.**

Recently, we described in this journal the synthesis of Group 4 metallocene nucleic acid like polymers (Figure 2) and their ability to inhibit a wide range of human cancer cells.<sup>[41]</sup>



**Figure 02: General repeat unit for the product of zirconocene dichloride and thymidine where R represents simple chain extension.**

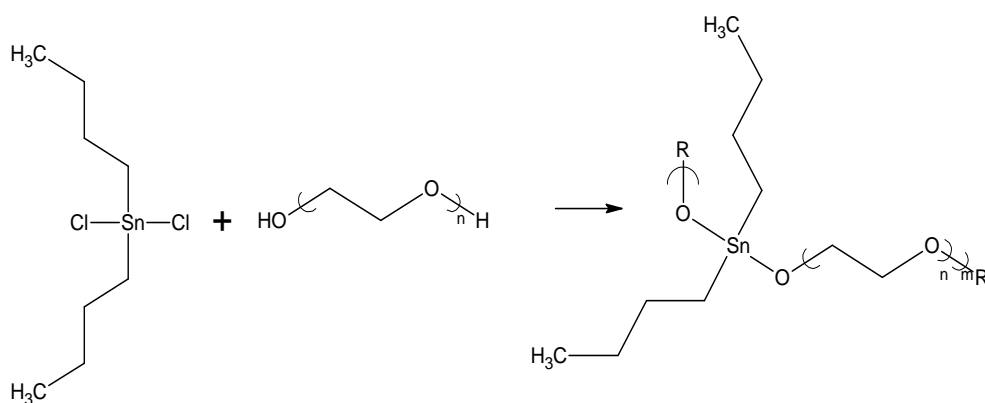
Here we describe the synthesis of the analogous organotin polymers (Figure 3).



**Figure 03: Reaction scheme for the synthesis of organotin nucleic acids derived from diethyltin dichloride.**

While it has characteristics of a nucleic acid with the presence of thymidine in a polyether repeat unit, there is no guarantee that the chain will have the precise 3'-5' unit throughout the chain. Even so, as will be shown these products exhibit good inhibition of a wide group of human cancer cell lines including breast, pancreatic, and glioblastomas brain cancer cells.

Structurally, the polymers can be named as polyethers. We have synthesized a wide variety of organotin polyethers. [10,15, 42-50] These include the first water-soluble metal-containing polymers from reaction of poly(ethylene glycol) (also called poly(ethylene oxide)) with various organotin dihalides. [10,15,42-44]



**Figure 04: Synthesis of organotin poly(ethylene glycol) polymers.**

## EXPERIMENTAL

**Reactions and Reactants:** Reactions were carried out using the interfacial polycondensation technique. Briefly, an aqueous solution (30 ml) containing the thymidine (0.00300 mol) and sodium hydroxide (0.00600 mol) was transferred to a one-quart Kimax emulsifying jar fitted on top of a Waring Blender (model 1120; no load speed of about 18,000 rpm; reactions were carried out at room temperature, at about 25°C). Stirring was begun and a hexane solution (30 ml) containing the organotin dichloride (0.00300 mol) was rapidly added (about 3-4 seconds) through a hole in the jar lid using a powder funnel. The resulting solution was blended for 15 seconds. The precipitate was recovered using vacuum filtration and washed several times with deionized water and hexane to remove unreacted materials and unwanted by-products. The solid was washed onto a glass petri dish and allowed to dry at room temperature.

Diphenyltin dichloride (1135-99-5), dimethyltin dichloride (753-73-1), thymidine (50-89-5) and dibutyltin dichloride (683-18-1) were purchased from Aldrich Chemical Co., Milwaukee, WI; diethyltin dichloride (866-55-7) was obtained from Peninsular Chemical Res., Gainesville, FL; and dioctyltin dichloride (3542-36-7), was obtained from Ventron Alfa Inorganics, Beverly, Mass.

**Physical Characterization:** Light scattering photometry was carried out employing a Brice-Phoenix Universal Light Scattering Photometer Model 4000. Infrared spectra were obtained employing attenuated total reflectance infrared spectroscopy utilizing a JASCO FT/IR-4100 fitted with an ATR Pro 450-s.

High resolution electron impact positive ion matrix assisted laser desorption ionization time of flight, HR MALDI-TOF, mass spectrometry was carried out employing a Voyager-DE STR BioSpectrometer, Applied Biosystems, Foster City, CA. The standard settings were used with a linear mode of operation and an accelerating voltage of 25,000 volts; grid voltage 90% and an acquisition mass range of 2000 to 100,000. Fifty to two hundred shots were typically taken for each spectrum. Results employing trans-2-[3-(4-tert-butylphenyl)-2-propenylidene], malonitrile (DCTB) as the matrix are included in the present paper.

**Cell Testing:** The toxicity of each test compound was evaluated using a variety of cancer cell lines and with human normal embryonic lung fibroblast (WI-38) and mouse embryo-fibroblast (NIH/3T3) cell line as standards. Following a 24 hr incubation period, the test compounds were added at concentrations ranging from 0.0032 to 32 microg/mL and allowed

to incubate at 37°C with 5% CO<sub>2</sub> for 72 hrs. Following incubation, Cell Titer-Blue reagent (Promega Corporation) was added (20 uL/well) and incubated for 2 hrs. Fluorescence was determined at 530/590 nm and converted to % cell viability versus control cells.

All cytotoxicity values were calculated against a base-line value for each line that was generated from “mock-treatment” of the normal and tumor cell lines with media supplemented with all diluents used to prepare the chemotherapeutic compounds. For example, if the compounds were dissolved in DMSO and serial dilutions prepared in Eagle’s minimal essential medium, MEM, to treat the cells, then the mock-treated cells were “treated” with the same serial dilutions of DMSO without added chemotherapeutic compound. This was done to ensure that any observed cytotoxicity was due to the activity of the compound and not the diluents. For the studies reported here, the mock-treatment never resulted in a loss of cell viability of more than one percent, demonstrating that the activity observed was not due to cytotoxicity of any of the diluents used, but was due to activity of the tested compounds. When inhibition begins, the slope of the concentration/inhibition curve is steep until total inhibition occurs.

## RESULTS AND DISCUSSION

Polymerization occurs employing the interfacial polymerization process developed by Morgan and coworkers at DuPont and enlarged by Carraher.<sup>[51-53]</sup> The interfacial synthetic technique is employed industrially in the production of aramid fibers and polycarbonates.<sup>[53,54]</sup> The rapidity of the reaction is a consequence of at least two factors.<sup>[53,54]</sup> First, the activation energy between acid chlorides and alcohols is about 20 kcal/mol compared with the typical reaction between acids and diols of about 40 kcal/mol. Second, the rapid stirring enlarges the interfacial surface on the order of 10,000 times. The reaction is rapid being completed within 10 seconds.

Table 1 contains the yield and chain length, DP, for the products as a function of the organotin moiety. Yield is in the minimum range. Chain length decreases as the size of the organic moiety on the tin increases consistent with size being an important factor in polymer chain growth.

**Table 01: Yield and chain length as a function of the organotin dihalide for the thymidine polymers.**

Organotin Moiety	Yield, %	Molecular Weight	Chain Length, DP
Me <sub>2</sub> Sn	51	4.9 x 10 <sup>6</sup>	12,000
Et <sub>2</sub> Sn	48	1.4 x 10 <sup>6</sup>	3,300
Bu <sub>2</sub> Sn	58	1.5 x 10 <sup>6</sup>	3,200
OC <sub>2</sub> Sn	67	1.0 x 10 <sup>6</sup>	1,700
Ph <sub>2</sub> Sn	85	5.6 x 10 <sup>6</sup>	11,000

**Infrared Spectral Results:** Table 2 contains band assignments for the diethyltin dichloride, thymidine, <sup>[55]</sup> and the resulting polymer. The polymer IR shows the absence of bands derived from the presence of the OH group as expected since it is now part of the connecting group Sn-O-C. The presence of bands characteristic of the thymidine are present including those derived from the C=O, N-H, and numerous C-H stretching, scissoring, wagging, rocking, and twisting associated bands. Diethyltin dichloride has seven characteristic bands, all present in the IR of the polymer. The Sn-C is found in the monomer at 521 and 492 and in the polymer at 533 and 490. The Sn-O band is generally found about 1000. The presence of a new band found at 1009 assigned to the formation of the Sn-O linkage. The presence of the O-C linkage in the Sn-O-C grouping is generally found about 770. A new band is found at 777 and is assigned to this linkage. Thus, IR spectral results are consistent with the proposed organotin polyether linkage.

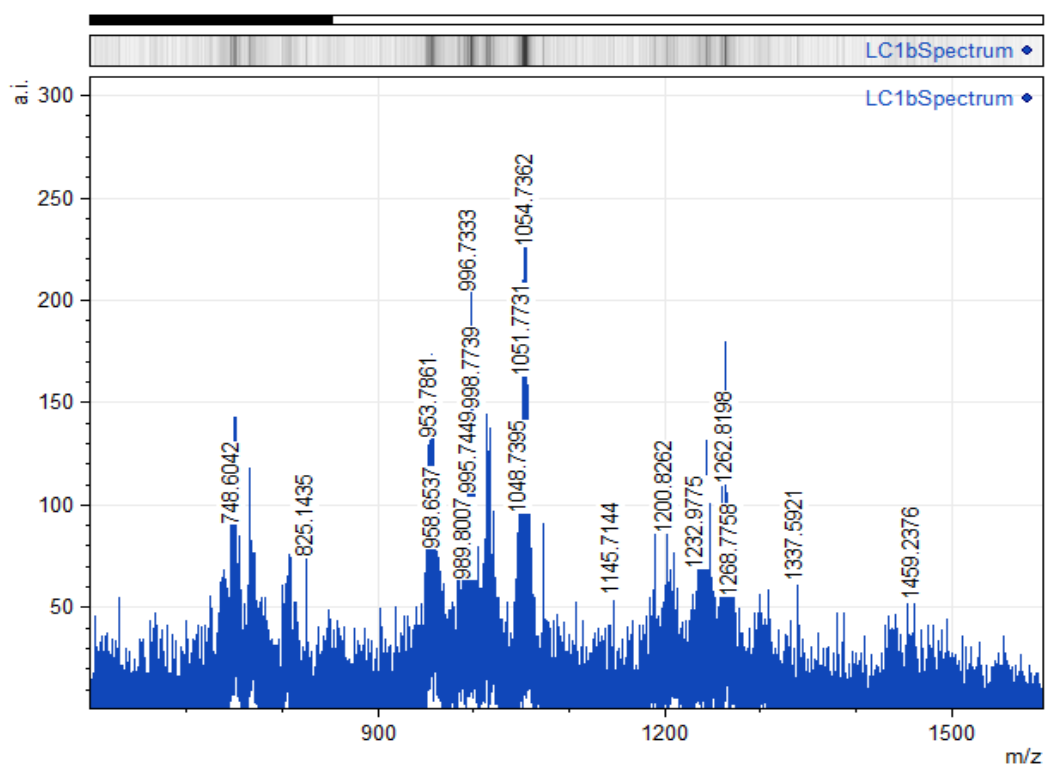
**Table 02: Infrared band assignments for the monomers and polymer from diethyltin dichloride and thymidine.**

Assignment	Et <sub>2</sub> SnCl <sub>2</sub>	Thymidine	Polymer
O-H		3641,3620,3482	
N-H		3428	3446
CH <sub>2</sub> st		2994	2990
CH <sub>3</sub> asym st	2979	2977	2975
CH <sub>2</sub> asym st	2962	2964, 2934	2947
CH <sub>3</sub> sym st	2929		2916
CH <sub>2</sub> sym st	2869		2868
C=O		1690,1665	1698,1657
CH <sub>2</sub> scissor bend	1448		1454
CH <sub>3</sub> op scissor		1435	1435
CH <sub>2</sub> wag		1172	1172
Sn-O-C st			1009
(Sn-)O-C			777
N-C op deformation		565	565
Sn-C asym st	521		533
Sn-C Sym st	492		490

**MALDI MS Results:** We have been employing MALDI MS to assist in the structural identification of our products. While MALDI MS was advertised as a method to obtain chain lengths, and molecular weights of polymers, it will do so only in certain cases. To be effective, the matrix and polymer must be soluble in volatile liquids that allow a “cocoon” to be made around the polymer. This casing both holds the polymer chain together and promotes the “volatilization” of the polymer allowing it to be ionized and analyzed. For most polymers this is not the case, so we utilize an approach that allows the MALDI MS to be used for samples that do not conform to this requirement. Polymers that possess heterochains tend to fracture under MALDI MS giving polymer chain fragments and it is the fragments rather, than entire chains, that are studied. The technique as employed by us has been recently reviewed. [55-59]

The following abbreviations are employed: T = thymidine fragment, Na = sodium (the sodium is present as a contaminant in the samples), U = one unit, 2U = 2 units, 3U = three units, etc.

A portion of the MALDI MS spectra for the polymer derived from dibutyltin dichloride and thymidine is given in Figure 5.



**Figure 05:** MALDI MS of the product of dibutyltin dichloride and thymidine over the approximate mass range of 600 to 1600 daltons.

The major ion fragments are given in Table 3. Ion fragments for up to nine units are found.

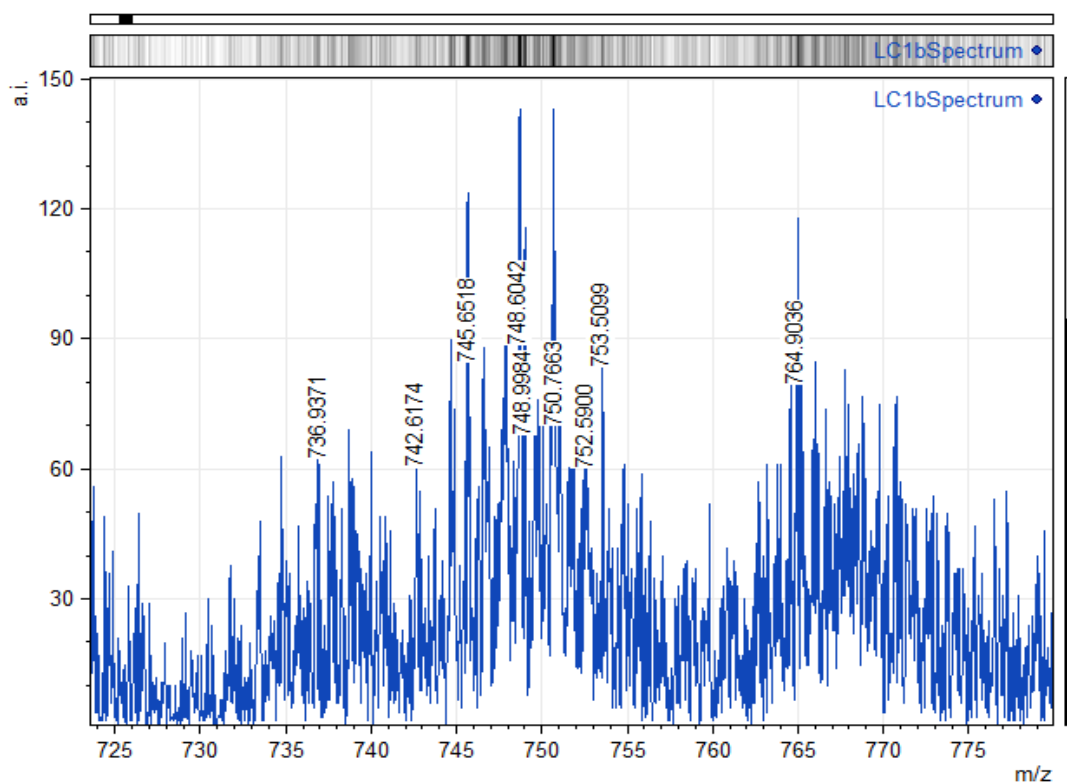
**Table 03: Major ion fragments for the product from dibutyltin dichloride and thymidine over the mass range of 600 to 4500 daltons.**

<b>Ion fragment</b>	<b>(Tentative) Structural Assignment</b>
749	U+Bu <sub>2</sub> Sn, O, Na
954	2U+Na-0
959	2U-0,Na
990	2U+0,Na
1145	2U+BuSn,Na
1201	2U+Bu <sub>2</sub> Sn,0
1459	3U+0,Na
1896	4U
2132	4U+Bu <sub>2</sub> Sn
2370	5U
2604	5U+Bu <sub>2</sub> Sn
2844	6U
3080	6U+Bu <sub>2</sub> Sn
3318	7U
3554	7U+Bu <sub>2</sub> Sn
3792	8U
4032	8U+T
4268	9U

Each of the spectral bands is actually a combination of many bands associated with different masses from isotopes associated with the isotopes of tin as shown in Figure 6 for the band centered about 749 daltons. The length of each band is associated with the abundance of the particular isotope and its natural abundance shown in the left column. The relative abundance is given in Table 4 for this particular peak. The relative abundance is similar to that predicted shown in the next to the furthest left-hand column consistent with the presence of two tin atoms contained within the overall band at 749 daltons.

A portion of the MALDI MS spectra for the product derived from the reaction of dioctyltin dichloride and thymidine is given in Figure 6. The major ion fragments and their tentative identify are given in Table 3.





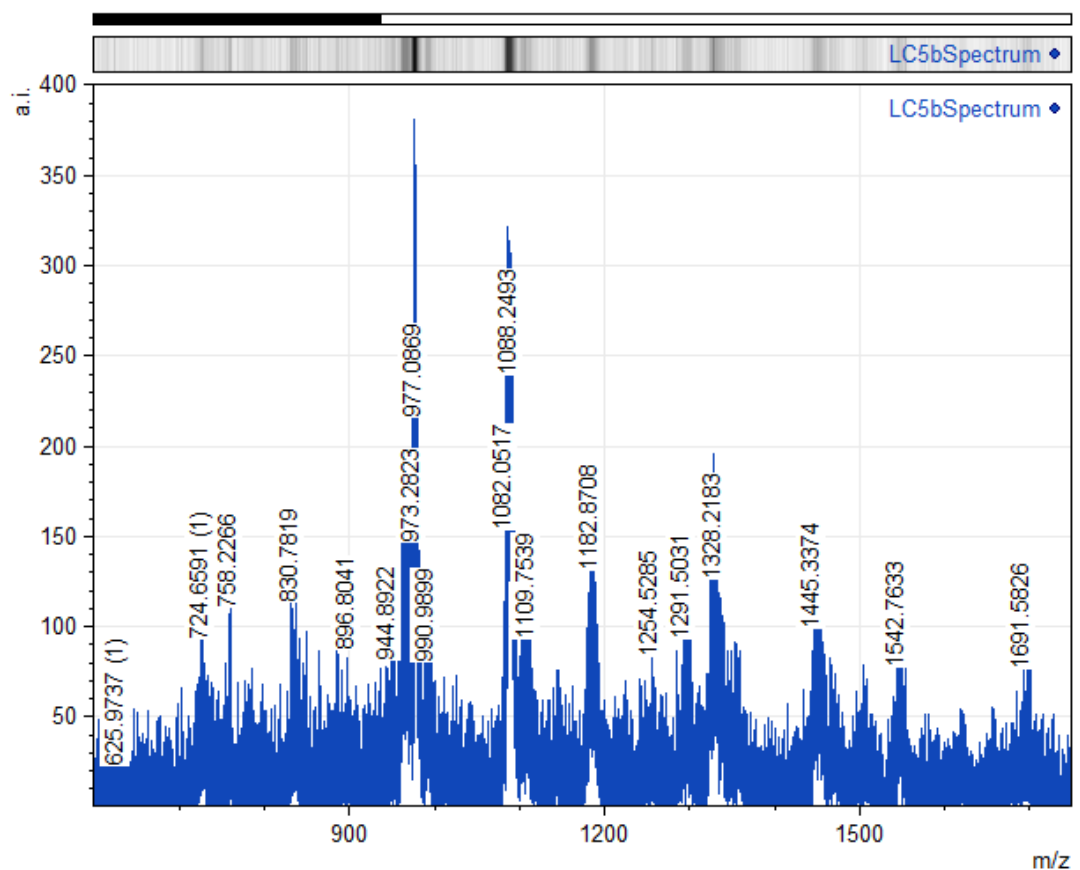
**Figure 06:** Portion of MALDI MS showing ion fragment bands for the polymer derived from the reaction of dibutyltin dichloride and thymidine.

Table 4 contains the ion fragment cluster analysis of two ions derived from the dibutyltin/thymidine product. The match is reasonable to the ion fragment cluster containing two tin atoms.

**Table 04:** Percentage relative abundances for two ion fragment clusters containing two tin atoms. Only ions with a (standard) relative abundance > 10% are given.

Standard		U+Bu <sub>2</sub> Sn <sub>2</sub> O <sub>2</sub> ,Na		2U+Na-0	
Da.	%-Rel. Abund.	Da.	%-Rel. Abund. Found	Da.	%-Rel. Abund. Found
232	12	743	12	948	12
233	13	744	13	949	14
234	43	745	42	950	42
235	35	746	34	951	34
236	94	747	92	952	91
237	51	748	52	953	52
238	100	749	100	954	100
239	35	750	34	955	36
240	81	751	82	956	81
242	32	753	31	958	30
244	22	755	21	960	23

Figure 7 contains a portion of the MALDI MS derived from the product of dioctyltin dichloride and thymidine.



**Figure 07:** MALDI MS of the product from dioctyltin dichloride and thymidine over the approximate mass range of 700 to 1700 daltons.

**Table 05:** contains major ions present in the MALDI MS of the product of dioctyltin dichloride and thymidine.

m/e	(Tentative) Structural Assignment
626	U+O,Na
831	U+T
945	U+Oc <sub>2</sub> Sn,0
973	U+Oc <sub>2</sub> Sn,O,Na
1183	2U+O
1445	2U+O,Na
1542	2U+Oc <sub>2</sub> Sn,O
1758	3U
1993	3U+T

2328	4U-O
2584	4U+T
2930	5U
3170	5U+T
3276	5U+Oc <sub>2</sub> Sn
3539	6U+Oc <sub>2</sub> Sn
3589	6U+Na
3862	6U+Oc <sub>2</sub> Sn
4118	7U+O
4358	7U+A

Ion fragments are present containing up to seven units. Table 5 contains a relative abundance match for an ion fragment cluster containing a single tin. The relative isotope abundances are consistent with the presence of a single tin atom in the cluster.

**Table 06: Percentage relative abundances for the ion fragment cluster assigned to U+T containing one tin atom. Only ions with a (standard) relative abundance > 10% are given.**

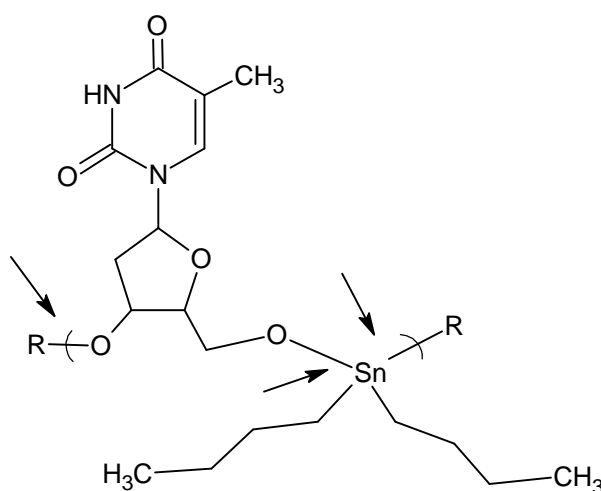
Standard		Found	
Da.	%-Relative Abundance	Da.	%-Relative Abundance
116	45	45	827
117	24	23	828
118	75	77	829
119	26	28	830
120	100	100	831
122	14	15	833
124	17	18	835

Table 7 contains isotope abundance matches for two ion fragment clusters containing two tin atoms.

**Table 07: Percentage relative abundances for ion fragment clusters containing two tin atoms.**

Standard		2U+O		2U+O,Na	
Da.	%-Rel. Abund.	Da.	%-Rel. Abund. Found	Da.	%-Rel. Abund. Found
232	12	1177	13	1439	13
233	13	1178	15	1440	15
234	43	1179	44	1441	44
235	35	1180	34	1442	36
236	94	1181	94	1443	95
237	51	1182	53	1444	51
238	100	1183	100	1445	100
239	35	1184	35	1446	34
240	81	1185	84	1447	83
242	32	1187	34	1449	29
244	22	1189	24	1451	22

As in essentially all cases with metal-containing products, the major sites for chain breakage is at the hetero atoms as shown in Figure 8.

**Figure 08: Sites where major chain scission occurs.**

**Cell Inhibition Results:** Much of our recent effort is the synthesis of potential drugs for the purpose of evaluation of structure/cancer activity building a catalogue of results that will assist in further syntheses. The cancer cells employed in the current study are given in Table 9.

**Table 09: Cell lines employed in the current study.**

Strain Number	NCI Designation	Species	Tumor Origin	Histological Type
3465	PC-3	Human	Prostate	Carcinoma
7233	MDA MB-231	Human	Pleural effusion breast	Adenocarcinoma
1507	HT-29	Human	Recto-sigmoid colon	Adenocarcinoma
7259	MCF-7	Human	Pleural effusion-breast	Adenocarcinoma
ATCC CCL-75	WI-38	Human	Normal embryonic lung	Fibroblast
CRL-1658	NIH/3T3	Mouse	Embryo-continuous cell line of highly contact-inhibited cells	Fibroblast
	U251	Human	Glioblastoma multiforme	Astrocytoma
	G55	Human	Glioblastoma	astrocytoma's
	AsPC-1	Human	Pancreatic cells	Adenocarcinoma
	PANC-1	Human	Epithelioid pancreatic cells	Carcinoma

Two values are typically taken when evaluating cell inhibition results. The first one is the amount of material that inhibits the particular cell line. It is generally given the name effective concentration, EC, and recorded as the amount necessary to inhibit 50% of the growth, EC<sub>50</sub>. Physically it is described at the amount that induces a response halfway between the baseline and maximum. As inhibition begins, the slope of the concentration vs inhibition is steep and continues to total inhibition. Table 10 contains EC<sub>50</sub> for monomers and polymers and the standard, cisplatin. Cisplatin is widely used in the treatment of a variety of cancers including brain tumors, ovarian cancer, testicular cancer, neuroblastoma, lung cancer, mesothelioma, bladder cancer, breast cancer, head and neck cancer and cervical cancer. It is quite toxic offering many unwanted side effects. Its high toxicity is shown from its low EC<sub>50</sub> values for the standard WI-38 and NIH-3T3 cells.

The EC<sub>50</sub> values for the polymers are generally less than those for the monomers. Thus, it is the combination and/or polymeric nature of the drug that results in the low EC<sub>50</sub> values. Polymeric drugs present several potential advantages over traditional small, monomeric drugs, for the inhibition of cancer. To follow is a brief list that highlights some of these advantages.<sup>[43-49]</sup> To begin, the much larger size of polymers compared to monomeric drugs can aid in the retention of the polymer drug in the body. Polymer drugs inability to easily pass through biological membranes due to their larger size increases the retention time which allows more time for drug to take effect. Also, a result, of their larger size, polymers are filtered by the kidneys more slowly compared to small molecules reducing possible kidney

damage. Next, again due to their larger size, polymers may allow for greater binding possibilities to the target than a monomeric drug. Polymers offer the ability to bind more locations increasing their activity and effectiveness. Another possible advantage is that polymers exhibit the ability to incorporate multiple anti-cancer drugs into a single polymer chain. This ability to contain more than one anti-cancer agent is unique to polymers and may prove beneficial as it allows for a single drug to have varying modes of action to combat cancer. Additionally, polymers can have the ability to behave as a prodrug allowing them to enter the cell in an inactive form and slowly degrade into an active form inside the cell. The slow degradation of the polymer into the active form can be thought of a sort of time release drug. Large molecules, like polymers, may have the ability to collect to a greater extent in solid tumors compared to healthy tissues due to the fact that tumors have limited lymphatic drainage therefore, the polymer drug is able to be built up in the tumor. This effect of the polymers collecting in the tumors is known as the enhanced permeability and retention (EPR) effect. Polymeric drugs also have the advantage of greater design possibilities compared to a monomeric drug. Polymers may be synthesized with specific characterizations, chain length, polarity, monomeric components and cross-linking allowing polymers the possibility to maximize their anticancer activity. Finally, polymers may be coupled to molecules that have the ability to target a specific transport mechanism. The coupling of these molecules can be used as an escort to transport the polymer drug to a specific site without influencing the drugs activity.<sup>[35]</sup>

Of interest here are the two cancer cell lines where there are currently no decent chemo agents, pancreatic and glioblastomas brain cancers. In the United States about 32,000 individuals are diagnosed with pancreatic cancer yearly, with the average life span after diagnosis being less than one year. Worldwide, it is the fourth leading cause of cancer related death. In the current study the two most widely studied human pancreatic cancer cell lines are selected. AsPC-1 is an adenocarcinoma pancreatic cell line, which accounts for around 85% of the diagnosed human pancreatic cancers, and PANC-1 is an epithelioid carcinoma pancreatic cell line, accounting for about 10% of the human pancreatic cancer cases. For the current study, the organotin polymers displayed the ability to inhibit both cell lines, the best inhibition for PANC-1 being the diethyltin (0.13 µg/mL) and the best inhibition for AsPC-1 also being the diethyltin (0.12 µg/mL). This similarity in the inhibition of both cell lines is consistent with the idea that the polymers offer broad-spectra inhibition of other pancreatic cancer cell lines.

Similar to pancreatic cancer, the expected life expectancy after diagnosis for glioblastoma is less than 1 year. The polymers were tested against two glioblastoma cell lines. G55 is a new human glioblastoma cell line. It was developed by C. David James (Department of Neurological, University of California San Francisco). G55 is a human glioblastoma cell line that had been passed through nude mice and re-established as a stable xenograft cell line. The U251 cell line is one of the most used cancer cell lines. It is derived from a human malignant glioblastoma multiforme. Astrocytoma's are brain cancer that originates in astrocytes. High-grade astrocytoma's, called glioblastoma multiforme, are the most malignant of all brain tumors. The thymidine polymers inhibit both cell lines.

The polymers also inhibit all of the other tested human cancer cell lines.

**Table 10: EC<sub>50</sub> Concentrations (micrograms/mL) for the tested compounds. Values Given in ( ) are standard deviations for each set of measurements.**

Sample	WI-38	PANC-1	AsPC-1	U251	G55
Me <sub>2</sub> SnCl <sub>2</sub>	0.22(.1)	0.80(.1)	0.71(.1)	0.91(.5)	1.2(.6)
Me <sub>2</sub> Sn/TH	0.45(.05)	0.25(.02)	0.23(.02)	0.51(.04)	0.55(.04)
Et <sub>2</sub> SnCl <sub>2</sub>	0.20(.1)	0.48(.1)	0.90(.1)	1.1(.6)	1.3(.6)
Et <sub>2</sub> Sn/TH	0.45(.05)	0.13(.02)	0.12(.02)	0.23(.04)	0.28(.04)
Bu <sub>2</sub> SnCl <sub>2</sub>	0.20(.05)	0.0032(.001)	0.012(.01)	1.0(.2)	1.0(.4)
Bu <sub>2</sub> Sn/TH	0.46(.05)	0.24(.02)	0.22(.02)	0.42(.04)	0.44(.04)
OC <sub>2</sub> SnCl <sub>2</sub>	0.30(.1)	0.85(.1)	0.85(.1)	1.3(.7)	0.95(.6)
OC <sub>2</sub> Sn/TH	0.44(.05)	0.23(.02)	0.22(.02)	0.44(.04)	0.49(.04)
Ph <sub>2</sub> SnCl <sub>2</sub>	0.25(.1)	0.71(.1)	0.83(.1)	0.68(.6)	0.89(.6)
Ph <sub>2</sub> Sn/TH	0.43(.05)	0.20(.02)	0.22(.02)	0.35(.04)	0.37(.04)
Thymidine	1.7(.5)	1.7(.5)	1.8(.5)	1.8(.5)	1.8(.5)
Cisplatin	0.012(.01)	0.0023(.005)	0.0035(.005)	0.015(.01)	0.020(.01)

Sample	PC-3	MDA-MB-231	HT-29	MCF-7	3T3
Me <sub>2</sub> SnCl <sub>2</sub>	0.51(.1)	0.44(.1)	0.56(.1)	0.66(.1)	0.43 (.1)
Me <sub>2</sub> Sn/TH	0.27(.02)	0.24(.02)	0.22(.02)	0.22(.02)	0.43(.05)
Et <sub>2</sub> SnCl <sub>2</sub>	0.61(.1)	0.64(.1)	0.71(.1)	0.77(.1)	0.46(.1)
Et <sub>2</sub> Sn/TH	0.13(.02)	0.11(.02)	0.12(.02)	0.11(.02)	0.42(.05)
Bu <sub>2</sub> SnCl <sub>2</sub>	1.4(1.1)	1.4(1.3)	1.2(.1)	0.70(.06)	0.20 (.05)
Bu <sub>2</sub> Sn/TH	0.23(.02)	0.25(.02)	0.21(.02)	0.23(.02)	0.45(.05)
OC <sub>2</sub> SnCl <sub>2</sub>	0.55(.1)	0.65(.1)	0.65(.1)	0.70(.1)	0.56(.1)
OC <sub>2</sub> Sn/TH	0.21(.02)	0.22(.02)	0.21(.02)	0.22(.02)	0.45(.05)
Ph <sub>2</sub> SnCl <sub>2</sub>	0.82(.1)	0.76(.1)	0.56(.1)	0.68(.1)	0.66(.1)
Ph <sub>2</sub> Sn/TH	0.24(.02)	0.24(.02)	0.21(.02)	0.21(.02)	0.42(.05)
Thymidine	1.8(.05)	1.8(.05)	1.8(.05)	1.7(.5)	1.8(.5)
Cisplatin	0.0044(.004)	0.0029(.002)	0.0041(.003)	0.0057(.003)	0.015(.01)

The second value used to evaluate the ability of materials to inhibit cell growth is the ratio of the  $EC_{50}$  for the polymers divided into the  $EC_{50}$  for a standard cell line, here the WI-38 human cell or NIH 3T3 cell line. The term chemotherapeutic index,  $CI_{50}$ , is often used to describe this ratio. It is generally accepted that WI-38 results are better at indicating ability of the tested compounds to prevent cancer growth in animals, and finally humans.

$CI_{50}$  values based on only WI-38 cells are presented in Table 11. Large values are desired since it indicates that there is a preference for inhibiting the cancer cell lines in comparison to the standard cells.

There are a number of instances with the  $CI_{50}$  values are near and greater than two with  $CI_{50}$  values in the same vicinity as cisplatin.

**Table 11:  $CI_{50}$  values for polymers and monomers based on data given in Table 10.**

Sample	$EC_{50}$ WI-38/ $EC_{50}$ PNC-1	$EC_{50}$ WI-38/ $EC_{50}$ AsPC-1	$EC_{50}$ WI-38/ $EC_{50}$ PC-3	$EC_{50}$ WI-38/ $EC_{50}$ MDA
$Me_2SnCl_2$	0.28	0.31	0.43	0.50
$Me_2Sn/TH$	1.8	2.0	1.7	1.9
$Et_2SnCl_2$	0.83	0.81	0.91	0.91
$Et_2Sn/TH$	3.5	3.8	3.5	2.8
$Bu_2SnCl_2$	60	17	0.14	0.14
$Bu_2Sn/TH$	1.9	2.1	2.3	1.8
$Oc_2SnCl_2$	0.35	0.35	0.55	0.46
$Oc_2Sn/TH$	1.9	1.9	2.1	2.0
$Ph_2SnCl_2$	0.35	0.31	0.30	0.33
$Ph_2Sn/TH$	2.2	2.0	1.8	1.8
Cisplatin	5.2	3.4	2.7	4.1

Sample	$EC_{50}$ WI-38/ $EC_{50}$ U251	$EC_{50}$ WI-38/ $EC_{50}$ G55	$EC_{50}$ WI-38/ $EC_{50}$ HT-29	$EC_{50}$ WI-38/ $EC_{50}$ MCF-7
$Me_2SnCl_2$	0.45	0.31	0.39	0.39
$Me_2Sn/TH$	0.88	0.82	2.1	2.1
$Et_2SnCl_2$	0.34	0.29	0.67	0.71
$Et_2Sn/TH$	0.20	0.16	3.8	4.1
$Bu_2SnCl_2$	0.33	0.33	0.17	0.29
$Bu_2Sn/TH$	1.0	1.2	2.2	2.3
$Oc_2SnCl_2$	0.34	0.46	0.46	0.43
$Oc_2Sn/TH$	0.12	0.12	2.1	2.0
$Ph_2SnCl_2$	0.29	0.30	0.45	0.37
$Ph_2Sn/TH$	1.0	0.90	2.0	2.0
Cisplatin	0.80	0.57	2.9	2.1



The lowest  $CI_{50}$  values are for the brain cancer cell lines and even here the dibutyltin polymers shows values of 1 and 1.2. A number of the polymers have  $CI_{50}$  values of two and greater for the pancreatic cancer cell lines. This is consistent with the drugs showing a good ability to differentiate between the healthy and cancer cell.

In summary, all of the polymers show decent  $EC_{50}$  values in the tenths of a microgram/mL range and many exhibit  $CI_{50}$  values of two and greater. Thus, they show good inhibition of the tested human cancer cell lines.

## SUMMARY

Synthetic nucleic acid polymers were synthesized in moderate yield from the interfacial polymerization of thymidine and organotin dihalides. Molecular weight decreases as the bulk on the organotin increases consistent with size having an influence on polymer formation. Infrared spectroscopy shows the formation of Sn-O bonds and absence of the R-OH group consistent with what is expected for the polyether formation. The polymers show good inhibition of a group of human cancer cell lines to the nanogram/mL range with good differentiation between the cancer and standard cell lines. They are a group of polymers that represent an additional structure for further study in the war against cancer.

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