

WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

Coden USA: WJPRAP

Impact Factor 8.453

Volume 14, Issue 22, 399-431.

Review Article

ISSN 2277-7105

MAJOR GENERA OF PAPAVERACEAE (POPPY) PLANT FAMILY IN ISRAEL AND PALESTINE. PART II: PAPAVER (POPPY)

Abdullatif Azab*

*Eastern Plants Company, Box 868, Arara, Israel, 30025.

Article Received on 18 Oct. 2025, Article Revised on 07 Nov. 2025, Article Published on 16 Nov. 2025,

https://doi.org/10.5281/zenodo.17616004

*Corresponding Author Abdullatif Azab

*Eastern Plants Company, Box 868, Arara, Israel, 30025.



How to cite this Article: Abdullatif Azab*. (2025). MAJOR GENERA OF PAPAVERACEAE (POPPY) PLANT FAMILY IN ISRAEL AND PALESTINE. PART II: PAPAVER (POPPY). World Journal of Pharmaceutical Research, 14(22), 399–431. This work is licensed under Creative Commons Attribution 4.0 International license.

ABSTRACT

Papaver is one of the genera included in the Papaveraceae plant family. All the species of this family have notable alkaloid content, and Papaver alkaloids that were isolated from the species of the reviewed region will be presented and discussed. These plants were used by humans since the dawn of humanity, and traditional uses are well documented. The greater part of this review article will be dedicated to Papaver somniferum because it is the most studied and most published among all Papaver species, including those that are not native to the reviewed region (RR). The chemical compositions of the plants will be extensively presented and discussed, including the factors that affect these compositions. Most of the information will be presented in tables, which enable readers to find it easily. Special attention will be drawn to dangers and toxicity of these plants. Finally, carefully selected, previously published

review articles about the *Papaver* species will also be presented.

KEYWORDS: Papaveraceae, *Papaver*, Anticancer, Alkaloids, Ethnomedicine, Chemical composition, *Papaver somniferum*, Papaverine, Thebaine, Toxicity.

Abbreviations: ABTS 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid), ahc and her/his colleagues, AChE acetylcholine esterase, BuChE butyrylcholine esterase, CUPRAC cupric reducing antioxidant capacity, DEE diethyl ether, DMSO dimethyl sulfoxide, DPPH 2,2-Diphenyl-1-picrylhydrazyl, EO essential oil, FRAP ferric reducing activity power, GCC general chemical composition, GC-MS gas chromatography mass spectrometry, GPx

www.wjpr.net Vol 14, Issue 22, 2025. ISO 9001: 2015 Certified Journal 399

glutathione peroxidase, HPLC high performance liquid chromatography, LC-MS liquid chromatography mass spectrometry, MDA malondialdehyde, NI not indicated, NPs nanoparticles, PE petroleum ether, RR reviewed region, SOD superoxide dismutase, TAC total antioxidant capacity, TFC total flavonoid content, TOS total oxidative status, TPC total phenolic content

1. Taxonomy, Archeology and Presence of Wild *Papaver* Plants in the RR

The *Papaver* genus as part of the Papaveraceae (Poppy, פֿרגיים, בּשׁבּוֹשׁנֵּא, English, Arabic, Hebrew, respectively) plant family, has a common name of Poppy, בֹשׁבּוֹשׁ in these languages, and the common name of the family is derived from the name of the genus and not vice versa.

In the first article of this series, we briefly presented the taxonomy of the Papaveraceae plant family. As for the taxonomy of the *Papaver* genus, it includes 100 to 110 species.^[1,2] In the RR, ten *Papaver* species grow wild, according to the "Flora of Israel and adjacent areas" website.^[3] These are: *Papaver argemone*, *P. carmeli*, *P. decaisnei*, *P. humile*, *P. hybridum*, *P. libanoticum*, *P. polytrichum*, *P. somniferum*, *P. syriacum* and *P. umbonatum*.

Archaeological studies of *Papaver* uses by ancient humans are mostly focused on *Papaver somniferum* due to its high activity compared to other *Papaver* species, and consequently, its economic value. A. Salavert ahc used radiocarbon dating and found that this species has been used in Western Europe since 5900 years before present time.^[4] A. D'Agostino ahc used the same dating method and found in a grave in Italy from the Neolithic period (around 6340 years before present) that this plant species was in use.^[5] Another study of Italian group by G. Giordano ahc revealed use of this plant in seventeenth century hospital in Milan.^[6] However, there are evidences that *Papaver somniferum* was utilized by human for medicinal purposes earlier than these findings, and some of these studies will be presented in the **Discussion** section.

2. Remarkable Published Review Articles about Papaver Genus and Species

A large number of review articles was previously published about this plant genus, where the vast majority of them focused on *Papaver somniferum* and its active constituents. For this reason, a very careful selection was applied. Most of these articles are presented in **Table 1**, and a few will be presented in the **Discussion** section.

Table 1: Selected published review articles about Papaver until 30/10/2025.

Author(s)	Pages	Ref's	Major topic(s), Ref.
J. Novák & V. Preininger	13	43	Chemotaxonomic review of the <i>Papaver</i> genus ^[7]
M. Butnariu ahc	23	181	Papaver genus. Very detailed and comprehensive: ethnomedicine, phytochemistry, medicinal activities, economics, safety ^[8]
F.S. Mohammed ahc	9	82	<i>Papaver</i> genus. Detailed, comprehensive, EOs and nutrition. Lacks ethnobotany. [9]
Z. Aghaali ahc	14	146	Papaver genus. Use of hairy roots to enhance alkaloids production. Outstanding figures. [10]
Y. Özgen & D. Burucu	9	37	Papaver genus. Focus on botany and farming in Turkey. Lacks important information. [11]
F. Labanca ahc	19	59	<i>P. somniferum.</i> Detailed taxonomy, uses, alkaloids in plant parts, biosynthesis ^[12]
M. Masihuddin ahc	5	43	<i>P. somniferum.</i> Traditional uses, phytochemistry, morphology ^[13]
A. Muhammad ahc	26	116	<i>P. somniferum.</i> Focus on nutritional and composition properties of seeds. [14]
M. Hedayati- Moghadama ahc	14	107	<i>P. somniferum.</i> Health benefits and adverse effects mainly on cardiovascular system ^[15]
Y. Jan & L.A. Peer	5	40	<i>P. somniferum.</i> Phytochemistry (no structures), detailed biological activities, toxicity ^[16]
A. Vadhel ahc	14	118	<i>P. somniferum.</i> Alkaloids (detailed), biosynthesis, health effects, link to cancer, mechanisms of action (detailed figures) ^[17]
H. Hamiksha	17	132	P. somniferum. Comprehensive. History, composition, legal aspects, health benefits and adverse effects, synthesis of active compounds ^[18]

3. Ethnobotanical Uses of *Papaver* Plants of Israel and Palestine

As mentioned earlier, *Papaver somniferum* is used by humanity for several millennia. These uses are nutritional, medicinal and rarely spiritual. However, some more uses will be presented in the **Discussion** section. So, unless special, the ethnobotanical uses of this species will not be presented in this section, unlike another wild *Papaver* plants of the RR. Summary of this data is presented in **Table 2**.

Table 2: Ethnobotanical Uses of *Papaver* Plants of Israel and Palestine.

Papaver Species	Country ; Plant part(s); method(s); use(s); reference		
P. argemone	Iran; flowers and capsules; antidiabetic, hepatoprotective, analgesic, neuroprotective, anti-irritant, anti-ulcerogenic ^[19] Poland; NI; NI; NI ^[20] Turkey; flowers; added to drink (Sherbet) ^[21] Seeds; infusion; cardiovascular problems ^[22] Leaves; cooked; food ^[23]		

<u>www.wjpr.net</u> | Vol 14, Issue 22, 2025. | ISO 9001: 2015 Certified Journal | 401

	Flowers, aerial parts; cooked; food ^[24]		
P. carmeli	Jordan; whole plant; eaten; CNS depressant, hypnotic ^[25]		
P. decaisnei	Pakistan; whole plant; NI; NI ^[26]		
P. humile	RR; flowers; ointment; wounds ^[27]		
P. numue	Aerial parts; animal food, eaten (grazing) ^[28]		
	Pakistan; flowers, fruits, stems; decoction; diaphoretic, sedative,		
	demulcent, flu, cough, diarrhea ^[29]		
	Flowers, fruits, decoction; ear and eye pain, cough, constipation ^[30]		
D. landari dana	Flowers; NI; flu, cough ^[31]		
P. hybridum	Fruits; decoction; insomnia ^[32]		
	Seeds; decoction; weak eyesight, pain killer, narcotic, body strength		
	enhancer ^[33]		
	Spain; leaves, stems; human food; eaten ^[34]		
	<u>Lebanon</u> : flowers; infusion; catarrhs ^[35]		
	Flowers; infusion; emollient, expectorant, sudorific sedative, cough,		
P. libanoticum	sleep enhancer ^[37]		
	Turkey; flowers, leaves; tea; sedative, somniferous, analgesic,		
	antitussive ^[36]		
	<u>China</u> ; fruits; NI; gastroenteritis, anti-inflammatory ^[38]		
	India; seeds; NI; demulcent, spasmolytic ^[39]		
	<u>Iran;</u> fruits, seeds; NI; laxative, tonic, hypnotic ^[40]		
	Jordan; whole plant; eaten; CNS depressant, hypnotic ^[25]		
	Pakistan; seeds, fruits; NI; cough ^[31]		
	Fruits (unripe); decoction; insomnia ^[32]		
	Seeds; decoction, tea; body strength enhancer, sedative, dental		
P. somniferum	problems, cough, memory enhancer, cough ^[33]		
	Fruits, seeds; tea; narcotic, cough, fever ^[41]		
	Poland; flowers, fruits, seeds; pressed oil, decoction, infusion; food,		
	varnish, feet pain, sleep enhancer, stomach disorders, cough ^[20]		
	<u>Turkey</u> ; fresh leaves; eaten; vasodilator ^[22]		
	Flowers, leaves (fresh or dry), seeds, fruits (fresh or dry), bark; tea;		
	analgesic, somniferous, vasodilator, children cry, sleep enhancer,		
	massage (oil) ^[36]		
	Jordan; NI; NI; earache, toothache, neuralgia, cough, insomnia, poor		
P. umbonatum	digestion ^[42]		
	<u>Lebanon</u> : fruits; decoction; measles in children ^[37]		

4. Published Medicinal Properties-Activities of Papaver Plants of Israel and Palestine

Compared with other plant genera, the number of medicinal properties-activities is relatively limited. And again, like in previous sections, *Papaver somniferum* is with clear lead. A major part of these properties will focus on chemical compositions and selected reported natural ingredients will be presented in the figures after **Table 3**.

Table 3: Medicinal Properties-Activities of *Papaver* Plants of Israel and Palestine.

Danguar anasias	Dronauty Activity Method Degulta Defende
Papaver species	Property-Activity, Method, Results, Reference
	Aerial parts were separately extracted with PE, DEE, chloroform,
	acetone and ethanol. All five extracts were tested for antimicrobial
	activity (six bacterial and one fungal species), showing moderate
	activities. Three standard drugs were used as references. [43]
	Whole plant methanol-chloroform (25-75%)* extract was tested for
	antibacterial activity against six bacteria species, showing significant
	effect. The extract was analyzed by HPLC for alkaloids resulting the
P. argemone	detection of four compounds: Morphine (major), Codeine, Papaverine
	and Thebaine (Figure 1). [44]
	Seeds were extracted (66-34% chloroform-methanol) and the extract
	was chromatographed for fatty acids composition. Results showed equal
	presence of saturated and unsaturated acids. [45]
	Fruits from various locations were harvested in several timings and
	were extracted (ethanol) for total alkaloid content. The extracts were
	analyzed with HPLC for major components: Morphine, Codeine,
	Papaverine Thebaine and Noscapine (Figure 1). [46]
P. carmeli	No published articles about medicinal properties-activities and/or
1. carmen	chemical composition
	Flowers, leaves and roots were extracted with methanol, and each
	extract was analyzed for GCC. They were tested for antioxidant
	(ABTS, CUPRAC, DPPH and FRAP methods) and anticancer (against
	Caco-2, MCF-7 and HeLa cancer cell lines) activities. The three
	extracts were chromatographed (HPLC) yielding previously known
	compounds, with very high content of Roemerine and lower content of
	Decarbomethoxytabersonine (Figure 2). [47]a
	Aerial parts were extracted with methanol and this extract was extracted
	again for total tertiary alkaloids content. Both extracts were tested for
P. decaisnei	antibacterial activity against E. faecalis and P. aeruginosa. TLC
	analysis of the methanolic extract afforded: Mecambrine (Figure 2) and
	Roemerine. ^[49]
	Aerial parts methanolic extract had antiulcer activity in ethanol-
	induced rats. Effect was measured with oxidative-antioxidative and
	inflammatory-anti-inflammatory biomarkers. It was tested for acute
	toxicity in theses animals and found nontoxic. [50]
	a- Interestingly, the same results were published by the same research
	group in the same year (different title and very slight changes in the
	text) in another journal. See ^[48]
n	No published articles about medicinal properties-activities and/or
P. humile	chemical composition
P. hybridum	Seeds were extracted (66-34% chloroform-methanol) and the extract
	was chromatographed for fatty acids composition. Results showed equal
	presence of saturated and unsaturated acids. [45]
	Aerial parts aqueous (alkaloid) extract was tested for antimicrobial
	activity against several bacterial and fungal stains, showing moderate
	effect. Several standard drugs were used as reference. [51]
	Aerial parts methanolic and methanolic-alkaloid extracts were evaluated
	for antibacterial activity (against 13 strains), with several standard
	101 distributed for the (against 15 strains), with several standard

www.wjpr.net | Vol 14, Issue 22, 2025. | ISO 9001: 2015 Certified Journal | 403

	drugs as references. Effect ranged from weak to moderate. [52]
	Flowers and leaves were combinedly extracted with 80% aqueous
	ethanol, and the extract was tested for antimicrobial activity against
	several bacteria and fungi strains. Results showed no effect. [53]
	Aerial parts (several locations and seasons) 80% aqueous methanolic
	extract was tested for antioxidant activity (DPPH method) and
	analyzed for GCC. The contents of Codeine and Papaverine were
	determined. [54]
	Follow up of previous cited study: same methods and same results. [55]
	Aerial parts 86% aqueous ethanolic extract was analyzed for TPC and
	total alkaloid content. It was tested for acute toxicity in mice (nontoxic)
	and analgesic activity, using tail flick, hot plate and acetic acid-induced
	writhing tests. Significant effect was detected with proposed mechanism
P. libanoticum	of action that includes involvement of opioids receptors. ^[56]
	Aerial parts 96% aqueous ethanolic (alkaloid) extract was analyzed
	affording Dehydroremerine, Berberine, Alborine, Remrefidine (Figure
	3), Roemerine and Mecambrine. These compounds were tested for
	anticancer activity against MCF7 and HCT116 cancer cells showing
	significant effects. Doxorubicin was reference drug. ^[57]
	Aerial parts aqueous (alkaloid) extract was tested for antimicrobial
P. polytrichum	activity against several bacterial and fungal stains, showing moderate
	effect. Several standard drugs were used as reference. ^[51]
	Seeds were extracted (66-34% chloroform-methanol) and the extract
	was chromatographed for fatty acids composition. Results showed equal
	presence of saturated and unsaturated acids. [45]
	Fruits from various locations were harvested in several timings and
	were extracted (ethanol) for total alkaloid content. The extracts were
	analyzed with HPLC for major components: Morphine, Codeine,
	Papaverine Thebaine and Noscapine (Figure 1). [46]
	Oripavine (Figure 4) was isolated from fruits and straws by HPLC and
	tested for analgesic activity in mice using hot plate method. It was also
	tested for toxicity and found nontoxic. Commercial Morphine and
	Thebaine were reference compounds. ^[58]
	Fruits, leaves, roots and stems were separately extracted with <i>n</i> -hexane,
	ethyl acetate and methanol. All twelve extracts were tested for
	anticancer activity (against four cancer cell lines), with 5-fluorouracil
P. somniferum	as reference drug. [59]
	Flowers, fruits (without seeds), leaves, roots, seeds and stems were
	separately and combinedly extracted with methanol. All seven extracts
	were tested for anticancer (against five cancer cell lines) and
	antioxidant (ABTS, DPPH, FRAP methods). Extracts were analyzed
	for TFC, TPC and total alkaloid content. They were also partially
	analyzed for chemical composition of 3-Hydroxy-β-damascone,
	Meconine (Figure 4), Codeine, Morphine and Papaverine. [60]
	From ethanolic (alkaloid) fruits extract Noscapine was isolated. [61]
	Leaves methanolic (alkaloid) extract was analyzed with GC-MS
	affording 25 alkaloids. Molecular docking for antiepileptic activity was
	performed for: Salutaridinol, R and S-Scoulerine, Coclaurine (-)-
	Codeinone and 7- <i>O</i> -Acetylsalutaridinol, (Figure 4). [62]
	Theoretical work (molecular docking) of active components for
	Theoretical work (molecular docking) of active components for

404

antiepileptic activity resulted highest potency of: γ -Isomorphine (**Figure 4**) Morphine, Codeine, Codeinone and *S*-Scoulerine. [63]

Seeds methanolic (alkaloid) extract had **antimicrobial** activity against *P. aeruginosa*, *L. monocytogenes*, *S. aureus*, *K. pneumoniae*, and *C. albicans*.^[64]

Leaves were separately extracted with ethyl acetate, ethanol, methanol, DMSO and water. The extracts were analyzed for TFC, TPC, total alkaloids content, glycosides, saponins and steroids. The extracts were tested for **antibacterial** (against six bacteria species) and **antioxidant** (DPPH method) activities. Effect ranged between non-existent to weak with clear dependence on solvent type.^[65]

Flowers EO was obtained by hydrodistillation and was tested for **antibacterial** activity against eight bacteria species. The EO was also analyzed for fatty acids (12 identified) composition using GC-MS.^[66]

Rats were orally administered with opium for a month until addiction was achieved. Th control group received Naloxone, a Morphine antagonist. Brain-related properties were tested such as **memory changes** (water maze method), **passive avoidance learning** (shuttle box method) and **oxidant-antioxidant** parameters (TAC, TOS, MDA, GPx and SOD).^[67]

Seeds were separately extracted with PE, chloroform and methanol, and the extracts were analyzed for GCC. They were also tested for **antioxidant** activity using DPPH method. [68]

Seeds (several cultivars) were defatted with *n*-hexane and extracted with methanol. The extract was analyzed for TPC and tested for **antioxidant** activity using DPPH and FRAP methods. [69]

Seeds (several genotypes) 50% aqueous ethanolic extract was analyzed for TFC and TPC, and tested for **antioxidant** (FRAP, TAC and reduction power methods) and **proteinase inhibition** activities.^[70]

Stalks were separately extracted with ethyl acetate, methanol and water. The extracts were tested for **antioxidant** (CUPRAC, FRAP and Fe⁺² chelating methods) and **enzyme inhibition** (tyrosinase, AChE, BuChE, α -amylase and α -glucosidase) activities. The extracts were partially analyzed for chemical composition yielding 4-Hydroxybenzoic acid as major component in each extract. [71]

Morphine content of cultivated plants is determined. [72]

Seeds were extracted with PE and the resulting oil was analyzed for triglycerides, monoglycerides and free fatty acids. [73]

Plants were cultivated in cold areas in Finland and their alkaloid (Morphine, Codeine, Thebaine, Noscapine and Papaverine) was determined by HPLC. [74]

Fruits GCC and enzymes (Lipase, Phospholipase, Peroxidase and Protease) activities were determined. [75]

Oil from different germplasms/varieties was extracted with PE and the yield and quality were determined. [76,78]

Amine oxidase was isolated from seedlings.^[77]

Fruits from several varieties were extracted with 75% aqueous methanol and the extracts were analyzed with HPLC to determine total alkaloid content and Morphine, Thebaine, Noscapine and Papaverine. [79]

Literature summary of metal (Cd, Pb, Fe) contents in seeds from

	Clavelrie and Czech Denublie in the 1000s [80]
	Slovakia and Czech Republic in the 1990s. [80]
	Seeds EO was obtained by n -hexane extraction, and it was tested for
	antibacterial activity against E. coli (inactive) S. aureus (active). The
	residue left after extraction was active against both bacteria. The EO
	was analyzed for GCC. ^[81]
	Seeds of twelve varieties harvested in different several seasons and
	several locations were analyzed for protein contents and fatty acids
	compositions. GCCs were also determined and physical properties
	(mainly seeds and flowers colour) were compared. [82]
	Seeds of several varieties grown in Estonia were extracted with 50%
	aqueous ethanol and the extracts were analyzed (HPLC) for Morphine,
	Codeine, Papaverine, and Apomorphine (Figure 4) contents. ^[83]
	Seeds were analyzed with LC-MS before and after thermal treatment to
	compare the contents of Morphine, Codeine, Thebaine, Noscapine and
	Papaverine. In this study, <i>P. setigerum</i> was also analyzed. ^[84]
	Seeds were sequentially extracted with <i>n</i> -pentane, ethyl acetate and
	70% aqueous methanol and the extract was analyzed for bitter-tasting
	components and their derivatives. Five compounds were found:
	Palmitic, Oleic, Linoleic (and ethyl ester) and α-Linolenic acids. [85]
	Seeds EOs were prepared using n -hexane, supercritical CO_2 and CO_2
	with 10% of acetone, ethanol, or ethyl acetate as co-solvents. The yields
	and compositions of the EOs were clearly different: fatty acids,
	phytosterols, squalene, tocopherols and phenolic acids. [86]
	Seeds with low Morphine contents from several varieties were cold
	pressed to obtain their EOs, which were analyzed for lipids (fatty acids,
	tocopherols and some volatile components). Higher concentrations were
	recorded compared with higher Morphine containing seeds. [87]
	Total alkaloid, Morphine, Codeine, Thebaine contents were determined
	in several Slovakian varieties. ^[88]
	Feeding pigs with seeds had positive effects growth and health. [89]
	Commercial seeds oil had insecticidal against Cowpea weevil
	(Callosobruchus maculatus) in stored Cowpea (Vigna unguiculata). [90]
	Seeds were harvested from several locations (Hungary) and their flour
	sensory and oil composition (volatiles and fatty acids) are reported. [91]
	Four out of 28 workers in morphine production in Spain were diagnosed
	with occupational asthma. Authors link this toxicity to their work. [92]
	Aerial parts aqueous (alkaloid) extract was tested for antimicrobial
	activity against several bacterial and fungal stains, showing moderate
	effect. Several standard drugs were used as reference. [51]
P. syriacum	Roemerine was isolated from aerial parts methanolic (alkaloid) extract
	and used as reference compound for the antimicrobial activity of
	alkaloids with very close structure, isolated from another plants. [93]
	Flowers were separately extracted with acetone, ethanol, methanol and
	water. The four extracts were tested for antibacterial activity (against
D umb on atum	three bacterial strains). Activities ranged from none to moderate, where
P. umbonatum	ethanolic extract was most active and acetone extract was weakest. [42]
	Aerial parts aqueous (alkaloid) extract was tested for antimicrobial
	activity against several bacterial and fungal stains, showing moderate
	effect. Several standard drugs were used as reference. [51]

^{*} Unless stated otherwise, percentages of solvent mixtures are V/V.

www.wjpr.net Vol 14, Issue 22, 2025. ISO 9001: 2015 Certified Journal 406

Figure 1: Natural products isolated from *Papaver argemone*.

Figure 2: Natural products isolated from Papaver decaisnei.

Figure 3: Natural products isolated from *Papaver libanoticum*.

Figure 4: Natural products isolated from Papaver somniferum.

5. DISCUSSION

Reviewing the publications about *Papaver* plants of Israel and Palestine is a task of very high interest, yet it is also challenging since a clear majority of these publications focus on a single species: Papaver somniferum. But some published works had comprehensiveness like the work of O. Bayazeid and F.N. Yalçin. [94] They carried out in silico predictions of possible activities of all 92 known alkaloids of Papaver genus. In addition, they classified and arranged these alkaloids according to their structural groups and linked these groups according to their biosynthesis. These groups are (with an example of each, Figure 5): Benzylisoquinoline (Armepavine), Promorphinane (8,14-Dihydropromorphinane), Morphinane (O-Methilflavinantine), Aporphine (N-Acetylanonaine), Proaporphine (Amuronine), Protoberberine (Canadine), Secoberbine (Macrantaline), Phthalideisoquinoline (Corlumine), Protopine (Hunnemanine), Rhoeadine (Oreogenine), and Isoquinoline (Cotarnoline) alkaloids.

Figure 5: Alkaloid Groups of *Papaver* Genus and Examples. [94]

Because of its high economic and medical importance, this alkaloid "paradise" was thoroughly studied, especially in *P. somniferum*. As mentioned above, the biosynthesis of these alkaloids was briefly presented in the previous cited article, and it is notably presented by J. Ziegler ahc.^[95] In a detailed scheme, the biosynthesis is presented, step by step, from L-Tyrosine to (*S*)-Norcoclaurine (**Figure 6**) to (*S*)-Reticuline, and then the biosynthesis branches to several pathways. The major interest of this publication is the biosynthesis of Morphine, and all alkaloids in this path were presented in **Figures 1-4** except Neopionine (**Figure 6**). It is important to mention that this publication presents the enzymes that catalyse the reactions.

Figure 6: Two of the Alkaloids Involved in the Biosynthesis of Morphine. [95]

For this review article, we summarized (**Table 4**) selected publication that presented the influence of various factors on the chemical composition and components isolation and identification of *Papaver somniferum*.

Table 4: Influencing Factors on the Chemical Composition of *Papaver somniferum.*

Author(s), Reference	Influencing Factor	Affected Factor
J.C. Laughlin ^[96]	Time of harvest	Yield of Morphine
J.C. Laughlin, D. Munro ^[97]	Fungal colonization	Yield of Morphine
P.J. Hofman, R.C. Menary ^[98]	Kiln drying	Alkaloid concentration (loss)
R. Luthra, N. Singh ^[99]	Triacylglycerols deposition	Fatty acid composition
K. Subrahmanyam ahc ^[100]	Sulfur fertilizers	Seed oil, alkaloid content
J.C. Laughlin, B. Chung ^[101]	N-fertilizers, irrigation	Yield and composition of alkaloids
R. Chizzola ^[102]	Low Cadmium stress	Micronutrient composition
T. Grothe ahc ^[103]	Salutaridinol 7- <i>O</i> -Acetyltransferase	Morphine biosynthesis
É. Németh ahc ^[104]	Successful breedings	Low alkaloid content
A. Balažová ahc ^[105]	Fungal elicitor	Sanguinarine content, Polyphenoloxidase activity
U. Fisinger ahc ^[106]	Thebaine synthase	Morphine biosynthesis
R. Khan ahc ^[107]	Gibberellic acid and Triacontanol	Morphine production
A. Mahdavi-Damghani ahc ^[108]	Water shortage	Alkaloids yield
A. Mahdavi-Damghani ahc ^[108] I. Holkova ahc ^[109,111]	Purification method	Isolated Lipoxygenase
N. Kara ^[110]	Sowing season	Oil and Morphine yields
R.K. Verma ahc ^[112]	Forensic methods	Compounds identification (review article)
M. Satranský ahc ^[113]	Cultivar type	Oil content, fatty acid profile
L. Yazici ^[114]	Sowing season, genotype	Oil and alkaloid contents
R.W. Neugschwandtner ahc ^[115]	Sowing season and rate	Yield and composition of components
Z. Aghaali ahc ^[116]	Various methods	Yield and composition of components (review article)
S.C. Carr ahc ^[117]	Aldo-Keto reductases	Morphine production

I. Varga ahc ^[118]	Selenium compounds	Seed and oil yields, fatty acid composition
D. Apaydin ^[119]	γ-Irradiation	Yields and compositions of phytosterols and fatty acids

One of the articles that demonstrate the extensive interest in *Papaver somniferum* was published by A. Diaz-B'arcena and P. Giraldo. [120] It is a review article that presents analysis and statistics of published articles about this plant species (and *Cannabis sativa*). Along with very readable text, this article includes very useful figures and graphs, which makes it very informative. In a similar fashion, yet less extensive but highly illustrative and informative, T. Beycioğlu recently published a bibliometric analysis of *Papaver* alkaloids and Morphine biosynthesis. [121] The earliest recorded use of *Papaver somniferum* by humans started around 8000 years ago, stated R.M. Kunwar and R.W. Bussmann in their review article of the ethnobotany in the Nepal Himalaya. [122] Another very early and very interesting article about this plant species was published by F.R. Stermitz and H. Rapoport in 1961. [123] By supplying 14CO_{2 (g)} to the growing plantlets, they investigated the biosynthesis and the interconversions of alkaloids. Interestingly, they reported that *Papaver somniferum* does not contain Oripavine (**Figure 4**). This report is inaccurate according to later publication (1998) by M.P. Gómez-Serranillos that isolated this compound from this species and even reported its analgesic activity. [58]

V. Lopez asks, "are traditional medicinal plants and ethnobotany still valuable approaches in pharmaceutical research?". He positively answers his question and gives *Papaver somniferum* as an example with its components and medicinal activities, and that it was described by the Greek physician-researcher Dioscorides (70 AD). The relevance and importance of *Papaver somniferum* can be found in other modern fields of research and medicine like preparation of nanoparticles (NPs). A summary of published *Papaver somniferum* extract assisted preparations of NPs and their activities, is presented in **Table 5**.

Table 5: NPs prepared with Papaver somniferum plants and their medicinal activities.

Nanoparticle(s)	Property-Activity, Reference
Ag	None ^[125]
Ag-Polyvinyl alcohol	Skin repair ^[126]
Au	None ^[127]
Co ₃ O ₄ -NiO	NaBH ₄ hydrolysis ^[128]
Fe ₂ O ₃ , PbO	Antimicrobial, antioxidant, antidiabetic, anticancer ^[129]
ZnO	Antidiabetic, haemolytic activity, antibacterial ^[130]
ZnO	Anticancer ^[131]

But *Papaver somniferum* is TOXIC and its toxicity reached fatality in some cases and was published many times. Here, four of these toxicity cases will be cited. H. Küçüker and F. Aydıner reported the death of 65-year-old woman in Turkey after boiling 10-15 capsules and drinking the solution.^[132] G. Bozan ahc reported another fatality of 4-years-old female, also from Turkey, that ingested raw plants.^[133] Y.P. Tan ahc reported saving a 21-years-old male that had acute cardiotoxicity after drinking self prepared seeds tea, and in the hospital (United Kingdom) was intensively treated with Naloxone.^[134] The fourth case, H. Hoummani ahc reported saving the life of a two-month-old infant (Morocco) who developed respiratory distress and hypotonia after his mother administered an oral preparation of opium poppy to calm him and help him sleep at a family party.^[135] This plant must be used legally and with extreme caution.

To conclude this discussion, we will elaborate on three out of six major alkaloids, Morphine, Papaverine and Thebaine (**Figure 1**), where the other three are Codeine, Oripavine and Noscapine.^[136]

In an outstanding review article, K. Brook ahc present the chemical history of Morphine. [137] The presentation advances step by step along 8000 years of use, historically and chemically, with very useful timeline at the end of the article. Morphine was tested and found potent in many medicinal activities, and consequently, many review articles were published about it. For example, and one of the most recent, the review article of A.J. Teunissen ahc which focuses on its use in non-abdominal surgery. [138] On the top of the properties of Morphine and the first that humans used is the analgesic activity. Modern science approved this ancient use and pain killing is the major use of this compound. Here are two examples from two different centuries: the work of A.A. Megens ahc and the work of X. Song ahc. [139,140] But morphine is addictive. [141,142] Morphine has adverse side effects: dry mouth, sedation, constipation, nausea, myoclonus, dysphoria and other minor health disorders. [143] And Morphine is toxic with some fatal cases. [144-148] This compound must be used legally and with extreme caution.

Papaverine was first isolated from *Papaver somniferum* by George Merck Fraz in 1848.^[149] One of its earliest and most common use was as spasmolytic drug^[150], where this study compared between Papaverine and two of its synthetic analogues, Octaverine and Perparine (**Figure 7**). Both synthetic compounds were more active than Papaverine.

Figure 7: Papaverine, Perparine and Octaverine - Spasmolytic Drugs. [150]

Modern pharmacological research discovered many more medicinal activities of this natural product, and here are some selected examples. Antiviral^[151,152], cardioprotective^[152,153] and anti-Parkinson.^[154] As a result, several review articles were published about this compound and the article of S. Ashrafi ahc is a very good example.^[155] It is comprehensive and detailed, and even though they name this compound "miraculous alkaloid", they also indicate some of its negative side effects and complications. But Papaverine has adverse side effects.^[156,157] And Papaverine can be toxic.^[158-160 (review article)] This compound must be used legally and with high caution.

Thebaine is an important alkaloid that has no direct use as a drug, but it can be easily transformed to other useful compounds like Morphine and Codeine (see below). For this reason, many attempts were carried out to find cheaper methods than chemical synthesis to produce Thebaine. Among these, the works of P.G. Vincent ahc and D. Palevitch and A. Levy, where both research groups increased the concentration of this natural product in *Papaver bracteatum*. [161,162]

Since Thebaine is an intermediate in the biosynthesis of Morphine^[163], many studies were published about Thebaine transformations that can occur or be induced because of the effects of many active factors. H. Kodaira and S. Spector transformed Thebaine to Oripavine, Codeine, and Morphine using rat liver, kidney, and brain microsomes.^[164] X. Li ahc used engineered *Escherichia coli* strains that increased the major enzymes (especially neopinone isomerase), that catalyse production codeine from thebaine.^[165] And an interesting finding was made by M.G. Carlin ahc that interconversion between the epimers of Thebaine in the presence of 80% water or more (**Figure 8**).^[166]

$$H_3CO$$
 H_3CO
 H_3CO
 H_3CO
 H_3CO
 H_3CO
 H_3CO
 H_3CO
 H_3CO
 H_3CO

Figure 8: Interconversion Between Thebaine Epimers. [166]

Several methods were developed to detect Thebaine. G. Cassella ahc published a new method for detection of Thebaine in urine using GC-MS.^[167] This method aimed to detect this compound as an indication of Poppy seed consumption. S. Lee ahc also used GC-MS to detect Thebaine in hair of rats that were administered with Poppy seeds.^[168] This method aimed to detect this compound as an indication of illegal opium substances chronic use.

But Thebaine has adverse side effects. Seeds of some cultivars of *Papaver somniferum* are used for food. The German Federal Institute for Risk Assessment (BfR) has published a clear warning that "the content of opium alkaloid thebaine should be reduced as much as possible". [169] J.W. Sloan ahe reported that Thebaine caused convulsions in rats after administration of a single dose of 20 mg/kg. [170] In a short review article, A. Eisenreich ahe stated that "thebaine exhibits a higher acute toxic potential than morphine. Therefore, exposure to thebaine by consumption of poppy seed-containing food could pose a health risk". [171] G. Navarro and H.W. Elliot reported that high doses of Thebaine caused adverse effects in rabbits and cats, tested by electroencephalogram (EEG). [172] Finally, R. Penafiel ahe reported after examining the cases of toxicity, stated that Thebaine can be toxic to "those consuming non-food grade poppy seeds as a tea". [173] This compound must be used legally and with high caution.

6. CONCLUSIONS

- 1. Papaver species of Israel and Palestine have extraordinary medical importance.
- 2. Some of these species were extensively studied and published while others were hardly published or not at all. So, research efforts are needed.
- 3. It is also important to test these plants and their materials for additional medicinal properties-activities.

- 4. Natural products isolated from these plants can be toxic. They must be legally used with high caution.
- 5. Because of their potential toxicity, these plants must be used legally and with high caution.

7. REFERENCES

- Stranska, I., Skalicky, M., Novak, J., Matyasova, E., Hejnak, V., Analysis of selected poppy (*Papaver somniferum* L.) cultivars: Pharmaceutically important alkaloids. *Indu. Crops Prod.*, 2013; 41: 120-126. DOI: 10.1016/j.indcrop.2012.04.018
- 2. Liu, L., Du, Y., Shen, C., Li, R., Lee, J., Li, P., The complete chloroplast genome of *Papaver setigerum* and comparative analyses in Papaveraceae. *Genet. Mol. Biol.*, 2020; 43: e20190272. DOI: 10.1590/1678-4685-GMB-2019-0272
- 3. Flora of Israel and Adjacent Areas: <u>Papaver</u>
- Salavert, A., Zazzo, A., Martin, L., Antolín, F., Gauthier, C., Thil, F., Tombret, O., Bouby, L., Manen, C., Mineo, M., Mueller-Bieniek, A., Piqué, R., Rottoli, M., Rovira, N., Toulemonde, F., Vostrovská, I., Direct dating reveals the early history of opium poppy in western Europe. *Sci. Rep.*, 2020; *10*: 20263. DOI: 10.1038/s41598-020-76924-3
- 5. D'Agostino, A., Di Marco, G., Rolfo, M.F., Canini, A., Gismondi, A., Exploring prehistoric plant use by molecular analyses of Neolithic grave goods. *Veg. His. Archaeobot*, 2023; *32*: 339-348. DOI: 10.1007/s00334-023-00910-8
- 6. Giordano, G., Mattia, M., Biehler-Gomez, L., Boracchi, M., Tritella, S., Maderna, E., Porro, A., Corsi Romanelli, M.M., Franchini, A.F., Galimberti, P.M., Slavazzi, F., Sardanelli, F., Di Candia, D., Cattaneo, C., *Papaver somniferum* in seventeenth century (Italy): archaeotoxicological study on brain and bone samples in patients from a hospital in Milan. *Sci. Rep.*, 2023; *13*: 3390. DOI: 10.1038/s41598-023-27953-1
- 7. Novák J., Preininger, V., Chemotaxonomic review of the genus *Papaver. Preslia.*, 1987; 59: 1-13. [Journal Website]
- 8. Butnariu, M., Quispe, C., Herrera-Bravo, J., Pentea, M., Sarac, I., Küşümler, A.S., Özçelik, B., Painuli, S., Semwal, P., Imran, M., Gondal, T.A., Emamzadeh-Yazdi, S., Lapava, N., Yousaf, Z., Kumar, M., Eid, A.H., Al-Dhaheri, Y., Suleria, H.A., Contreras, M., Sharifi-Rad, J., Cho, W.C., *Papaver* plants: current insights on phytochemical and nutritional composition along with biotechnological applications. *Oxid. Med. Cell. Longev*, 2022; 2041769. DOI: 10.1155/2022/2041769

- 9. Mohammed, F.S., Uysal, I., Yaz, H.H., Sevindik, M., *Papaver* species: usage areas, essential oil, nutrient and elements contents, biological activities. *Prosp. Pharm. Sci.*, 2023; *21*: 1-9. DOI: 10.56782/pps.142
- 10. Aghaali, Z., Naghavi, M.R., Zargar, M., Collaboration of hairy root culture and scale-up strategies for enhancing the biosynthesis of medicinal and defensive alkaloids in *Papaver* sp. *Curr. Plant Biol.*, 2024; *40*: 100381. DOI: 10.1016/j.cpb.2024.100381
- Özgen, Y., Burucu, D., History, Cultivation, and Adaptation of *Papaver* Species Globally. *Turk. J. Agric*, 2025; *13*: 1083-1091. DOI: 10.24925/turjaf.v13i4.1083-1091.7425
- 12. Labanca, F., Ovesnà, J., Milella, L., *Papaver somniferum* L. taxonomy, uses and new insight in poppy alkaloid pathways. *Phytochem. Rev.*, 2018; *17:* 853-871. DOI: 10.1007/s11101-018-9563-3
- 13. Masihuddin, M., Jafri, M.A., Siddiqui, A., Chaudhary, S., Traditional uses, phytochemistry and pharmacological activities of *Papaver somniferum* with special reference of Unani medicine: an updated review, J. *Drug Deliv. Therap.*, 2018; 8: 110-114. DOI: 10.22270/jddt.v8i5-s.2069
- 14. Muhammad, A., Akhtar, A., Aslam, S., Khan, R.S., Ahmed, Z., Khalid, N., Review on physicochemical, medicinal and nutraceutical properties of poppy seeds: a potential functional food ingredient. *Funct. Foods Health Dis.*, 2021; 11: 522-547. DOI: 10.31989/ffhd.v11i10.836
- 15. Hedayati-Moghadam, M., Moezi, S.A., Kazemi, T., Sami, A., Akram, M., Rida, Z., Khazdair, M.R., The effects of *Papaver somniferum* (Opium poppy) on health, its controversies and consensus evidence. *Toxin Rev.*, 2021; 41. DOI: 10.1080/15569543.2021.1958232
- 16. Jan, Y., Peer, L.A., *Papaver somniferum*: Phytochemistry, Biological Activity and Toxicology; a review. *Int. J. Bot. Stud.*, 2021; 6: 753-757. [ResearchGate]
- 17. Vadhel, A., Bashir, S., Mir, A.H., Girdhar, M., Kumar, D., Kumar, A., Mohan, A., Malik, T., Mohan, A., Opium alkaloids, biosynthesis, pharmacology and association with cancer occurrence. *Open Biol.*, 2023; 13: 220355. DOI: 10.1098/rsob.22.0355
- 18. Hamiksha. H., The Mind Based and Most Controversial Plant: *Papaver somniferum* the Opium Poppy: A Plant with Many Faces and Roles in Human History and Culture. *Neuropsychiatry*, 2024; *14*: 1-17. DOI: 10.37532/1758-2008.2024.14(4).727

- 19. Mossiavand, A.M., Tarnian, F., Rastegar, A., Ethnobotanical survey of medicinal plants in Delfan County, Lorestan Province, Iran. *J. Rangeland Sci.*, 2025; *15*: 152538. DOI: 10.57647/j.jrs.2025.1504.38
- 20. Zemanek, A., Zemanek, B., Klepacki, P., Madeja, J., The poppy (*Papaver*) in old Polish botanical literature and culture. *Edipuglia*, 2009; 217-226. [ResearchGate]
- 21. Kadioglu, Z., Cukadar, K., Kalkan, N.N., Vurgun, H., Kaya, O., Wild edible plant species used in the Ağrı province, eastern Turkey. *An. Jard. Bot. Madr.*, 2020; 77: e098. DOI: 10.3989/ajbm.2554
- 22. Güleç, M., Erarslan, Z.B., Kültür, Ş., The Medicinal Plants Traditionally Used Against Cardiovascular Diseases in Türkiye. *Int. J. Trad. Complement. Med. Res.*, 2023; *4:* 81-96. DOI: 10.53811/ijtcmr.1232190
- 23. Akbulut, S., Zengin, Z., Ethnobotanical survey of wild plants used in Gümüşhane province (Turkey). *Bol. Latinoam. Caribe Plantas Med. Aromát*, 2023; 22: 237-254. DOI: 10.37360/blacpma.23.22.2.18
- 24. Emre, G., Senkarde, I., Iscan, K., Evcimen, O., Yilmaz, I., Osman Tugay, O., An Ethnobotanical Study in Kirsehir (Turkiye). *Plants*, 2024; *13*: 2895. DOI: 10.3390/plants13202895
- 25. Al-Qura'n, S., Ethnobotanical survey of folk toxic plants in southern part of Jordan. *Toxicon*, 2005; 46: 119-129. DOI: 10.1016/j.toxicon.2005.04.010
- 26. Ulhaq, A., Saeed, S., Ahmed, A., Species diversity and ethnobotanical inventory of wild flora used by the folk community of Shinghar Balochistan, Pakistan. *Nusantara Biosci.*, 2021; *13*: 148-157. DOI: 10.13057/nusbiosci/n130203
- 27. Bailey, C., Danin, A., Bedouin plant utilization in Sinai and the Negev. *Econ. Bot.*, 1981; *35:* 145-162. DOI: 10.1007/BF02858682
- 28. Abou Auda, M.M., Contribution to the Plant Ecology and the Most Palatable Species for Grazing in the Gaza Strip Mediterranean Coast, Palestine. *Asian J. Plant Sci.*, 2010; 9: 88-93. DOI: 10.3923/ajps.2010.88.93
- 29. Iqbal, H., Sher, Z., Medicinal plants from salt range Pind Dadan Khan, district Jhelum, Punjab, Pakistan. *J. Med. Plants Res.*, 2011; 5: 2157-2168. [ResearchGate]
- 30. Afzal, T., Munawar, T., Narejo, G.A., Proćkow, J., Bibi, Y., Jabeen, A., Ethnomedicinal plant utilization in rural Northern Punjab, Pakistan: A quantitative and qualitative analysis. *Intl. J. Agric. Biol.*, 2025; *34*: 340312. DOI: 10.17957/IJAB/15.2372
- 31. Alamgeer, A., Younis, W., Asif, H., Sharif, A., Riaz, H., Bukhari, I.A., Assiri, A.M., Traditional medicinal plants used for respiratory disorders in Pakistan: a review of the

- ethno-medicinal and pharmacological evidence. *Chin. Med.*, 2018; *13*: 48. DOI: 10.1186/s13020-018-0204-y
- 32. Khan, A.W., Khan, A.U., Shah, S.M., Ullah, A., Faheem, M., Saleem, M., An Updated List of Neuromedicinal Plants of Pakistan, Their Uses, and Phytochemistry. *Evid. Based Complement. Alternat. Med.*, 2019; 6191505. DOI: 10.1155/2019/6191505
- 33. Ishtiaq, M., Maqbool, M., Ajaib, M., Ahmed, M., Hussain, I., Khanam, H., Mushtaq, W., Hussain, T., Azam, S., Hayat Bhatti, K., Ghani, A., Ethnomedicinal and folklore inventory of wild plants used by rural communities of valley Samahni, District Bhimber Azad Jammu and Kashmir, Pakistan. *PLoS One.*, 2021; *16*: e0243151. DOI: 10.1371/journal.pone.0243151
- 34. Blanco-Salas, J., Gutierrez-Garcia, L., Labrador-Moreno, J., Ruiz-Tellez, T., Wild plants potentially used in human food in the Protected Area" Sierra Grande de Hornachos" of Extremadura (Spain). *Sustainability*, 2019; *11*: 456. DOI: 10.3390/su11020456
- 35. Baydoun, S., Chalak, L., Dalleh, H., Arnold, N., Ethnopharmacological survey of medicinal plants used in traditional medicine by the communities of Mount Hermon, Lebanon. *J. Ethnopharmacol*, 2015; *173*: 139-156. DOI: 10.1016/j.jep.2015.06.052
- 36. Memnune Erucar, F., Tan, N., Miski, M., Ethnobotanical Records of Medicinal Plants of Turkey Effective on Stress Management Complied with the Literature Survey in Their Chemical Content and Activities. *Rec. Nat. Prod.*, 2023; 17: 1-92. DOI: 10.25135/rnp.324.2203.2372
- 37. Arnold, N., Baydoun, S., Chalak, L., Raus, T., A contribution to the flora and ethnobotanical knowledge of Mount Hermon, Lebanon. *Fl. Medit.*, 2015; 25: 13-55. DOI: 10.7320/FlMedit25.013
- 38. Gao, L., Wei, N., Yang, G., Zhang, Z., Liu, G., Cai, C., Ethnomedicine study on traditional medicinal plants in the Wuliang Mountains of Jingdong, Yunnan, China. *J. Ethnobiol. Ethnomed*, 2019; *15*: 41. DOI: 10.1186/s13002-019-0316-1
- 39. Goyal, M., Use of ethnomedicinal plants for prophylaxis and management of postpartum complications among the Marwari community of Jodhpur District of Rajasthan. *Food Qual. Saf.*, 2017; *1:* 203-209. DOI: 10.1093/fqsafe/fyx013
- 40. Buso, P., Manfredini, S., Ahmadi-Ashtiani, H.R., Sciabica, S., Buzzi, R., Vertuani, S., Baldisserotto, A., Iranian Medicinal Plants: From Ethnomedicine to Actual Studies. *Medicina*, 2020; *56*: 97. DOI: 10.3390/medicina56030097

- 41. Akhtar, N., Rashid, A., Murad, W., Bergmeier, E., Diversity and use of ethno-medicinal plants in the region of Swat, North Pakistan. *J. Ethnobiol. Ethnomed.*, 2013; 9: 25. DOI: 10.1186/1746-4269-9-25
- 42. Obeidat, M., Antimicrobial activities from extracts of seven medicinal plant species against multidrug-resistant bacteria and fungi. *J. Pharmacog. Phytother*, 2018; *10*: 45-55. DOI: 10.5897/JPP2017.0482
- 43. Ünsal, Ç., Özbek, B., Sariyar, G., Mat, A., Antimicrobial activity of four annual *Papaver* species growing in Turkey, *Pharm. Biol.*, 2009; 47: 4-6. DOI: 10.1080/13880200802392468
- 44. Mohamadi, S., Bahramnejad, B., Majdi, M., Soltani, J., Alkaloid profiling and antimicrobial activities of some Papaver species from Kurdistan province, Iran. *Iran. J. Med. Arom. Plants Res.*, 2024; 40: 352-366. DOI: 10.22092/ijmapr.2023.363324.3362 [Article in Persian]
- 45. Marin, P., Sajdl, V., Kapor, S., Tatić, B., Fatty acid composition of seeds of the Papaveraceae and Fumariaceae. *Phytochemistry*, 1989; 28: 133-137. DOI: 10.1016/0031-9422(89)85024-1
- 46. Almousawi, U.M., Alwan, A.A., The significance of opium alkaloids in the classification of Papaveraceae in Iraq. *J. Pharmacog. Phytochem*, 2017; 6: 430-437. [ResearchGate]
- 47. Jabbar, A.A., Abdullah, F.O., Abdulrahman, K.K., Galali, Y., Sardar, A.S., GC-MS Analysis of Bioactive Compounds in Methanolic Extracts of *Papaver decaisnei* and Determination of Its Antioxidants and Anticancer Activities. *J. Food Qual*, 2022; 1405157. DOI: 10.1155/2022/1405157
- 48. Jabbar, A.A., Abdullah, F.O., Abdulrahman, K.K., Galali, Y., Sardar, A.S., *Papaver decaisnei*: GC-MS alkaloids profiling, in vitro antioxidant, and anticancer activity. *Res. Square*, 2022; 1-18. DOI: 10.21203/rs.3.rs-1207324/v1
- 49. Jafaar, H.J., Isbilen, O., Volkan, E., Sariyar, G., Alkaloid profiling and antimicrobial activities of *Papaver glaucum* and *P. decaisnei*. *BMC Res. Notes.*, 2021; *14*: 348. DOI: 10.1186/s13104-021-05762-x
- 50. Jabbar, A.A., Abdullah, F.O., Abdoulrahman, K., Galali, Y., Ibrahim, I.A., Alzahrani, A.R., Hassan, R.R., Gastroprotective, biochemical and acute toxicity effects of *Papaver decaisnei* against ethanol-induced gastric ulcers in rats. *Processes*, 2022; 10: 1985. DOI: 10.3390/pr10101985
- 51. Shbibe, L.M., Babojian, G.A., Study of the Effectiveness of *Papaver Sp.* Alkaloids as Future Therapeutic Alternatives against *Enterococcus Sp.* Causing Hospital-Acquired

- Septicemic Infections. *J. Biosci. Med.*, 2024; *12*: 107-127. DOI: 10.4236/jbm.2024.128008
- 52. Nanaha, H., Bello, A., Tahan, Z.S., Comparative Evaluation of the Antibacterial Efficacy of *Papaver rhoeas* and *Papaver hybridum* Extracts against Respiratory Tract Pathogens. *Sci. J. Aleppo Univ.*, 2024; *174*: 1-19. [ResearchGate]
- 53. Dulger, B., Gonuz, A., Antimicrobial activity of some Turkish medicinal plants. *Pak. J. Biol. Sci.*, 2004; 7: 1559-1562. [ResearchGate]
- 54. Nazari, N., Sharifnia, F., Salimpour, F., Mehrnia, M., Gran, A., Evaluation of phytochemical traits and codeine and papaverine alkaloids of *Papaver* species in west and central Iran: application of principal component analysis and cluster analysis. *Yafte.*, 2024; 26: 56-72. [Journal Website, article in Persian]
- 55. Nazari, N., Sharifnia, F., Salimpour, F., Mehrnia, M., Gran, A., Zarafshar, M., Phytochemical richness, antioxidant activity, and molecular diversity in *Papaver* species from Iran's western and central regions. *Front. Plant Sci.*, 2025; *16*: 1635867. DOI: 10.3389/fpls.2025.1635867
- 56. Hijazi, M.A., El-Mallah, A., Aboul-Ela, M., Ellakany, A., Evaluation of Analgesic Activity of *Papaver libanoticum* Extract in Mice: Involvement of Opioids Receptors. *Evid. Based Complement. Alternat. Med.*, 2017; 8935085. DOI: 10.1155/2017/8935085
- 57. Hijazi, M.A., Aboul-Ela, M., Bouhadir, K., Fatfat, M., Gali-Muhtasib, H., Ellakany, A., Alkaloids of *Papaver libanoticum* and their cytotoxic activity. *Rec. Nat. Prod.*, 2018; *12*: 611-618. DOI: 10.25135/rnp.53.17.11.187
- 58. Gómez-Serranillos, M.P., Palomino, O.M., Carretero, E., Villar, A., Analytical study and analgesic activity of oripavine from *Papaver somniferum* L. *Phytother. Res.*, 1998; *12*: 346-349. DOI: 10.1002/(SICI)1099-1573(199808)12:5<346::AID-PTR307>3.0.CO;2-A
- 59. Güler, D.A., Aydin, A., Koyuncu, M., Parmaksiz, I., Anticancer activity of *Papaver somniferum*. J. Turk. Chem. Soc., 2016; 3: 349-366. DOI: 10.18596/jotcsa.43273
- 60. Sharopov, F., Valiev, A., Gulmurodov, I., Sobeh, M., Satyal, P., Wink, M., Alkaloid content, antioxidant and cytotoxic activities of various parts of *Papaver somniferum*. *Pharm. Chem. J.*, 2018; 52: 459-463. DOI: 10.1007/s11094-018-1839-9
- 61. Bulduk, I., Taktak, F., Isolation and Characterization of Antitumor Alkaloid from Poppy Capsules (*Papaver somniferum*). *J. Chem.*, 2013; 493870. DOI: 10.1155/2013/493870
- 62. Shahzadi, K., Rasool, D., Irshad, F., khalid, A., Aslam, S., Nazir, A., Identification and characterization of anti-epileptic compounds from *Papaver somniferum* using quantification techniques (GC-MS, FTIR), integrated network pharmacology, molecular

- docking, and molecular dynamics simulations. *bioRxiv*., 2024; 2024-07. DOI: 10.1101/2024.07.29.605558
- 63. Bibi, F., Rauf, A., Khan, M.A., Ashfaq, U.A., Awan, M.Q., Masoud, M.S., Zarmeena, R., Bhinder, M.A., Afandi, M.H., Bhatti, R., Network Pharmacological Study of *Papaver somniferum* to Explore the Potential Compounds to Treat Epilepsy. *J. Liaquat Univ. Med. Health Sci.*, 2024; 24: 25-32. DOI: 10.22442/jlumhs.2024.01154
- 64. Ismaili, A., Sohrabi, S.M., Azadpour, M., Heydari, R., Rashidipour, M., Evaluation of the antimicrobial activity of alkaloid extracts of four *Papaver* species. *Herb. Med. J.*, 2017; 2: 146-152. DOI: 10.22087/hmj.v0i0.638
- 65. Thetchinamoorthi, R., Kalidass Subramaniam, K., Phytochemical Profile, Antibacterial Efficacy, and Antioxidant Potential of Eight Ethnomedicinal Plants: A Solvent-Dependent Study. *Rev. Electron. Vet.*, 2024; 25: 4094-4102. [Journal Website]
- 66. Dilek, M., Gültepe, A., Öztaşan, N., 2018. Determination of essential oil composition and investigation of antimicrobial properties of poppy (*Papaver Somniferum* L.) flower. *AKU J. Sci. Eng.*, 2018; *18*: 786-795. DOI: 10.5578/fmbd.67616
- 67. Abbasi, E., Mirzaei, F., Mashayekhi, S., Khodadadi, I., Komaki, A., Faraji, N., Vafaii, S.A., Effects of opium on hippocampal-dependent memory, antioxidant enzyme levels, oxidative stress markers, and histopathological changes of rat hippocampus. *Biochem. Biophys. Rep.*, 2025; 42: 101993. DOI: 10.1016/j.bbrep.2025.101993
- 68. Abd Elaleem, K.G., Saeed, B.A., Ahamed, H.A., Phytochemical Screening and Antioxidant Activity Evaluation of *Papaver Somniferum* L Seed Extract from Eastern Sudan. *Int. J. Sci. Res.*, 2017; 6: 1391-1394. DOI: 10.21275/ART20178134
- 69. Chmelová, D., Ondrejovič, M., Havrlentová, M., Kraic, J., Evaluation of polar polyphenols with antioxidant activities in *Papaver somniferum L. J. Food Nutr. Res.*, 2018; *57*: 98–107. [ResearchGate]
- Krošlák, E., Maliar, T., Nemeček, P., Viskupičová, J., Maliarová, M., Havrlentová, M., Kraic, J., Antioxidant and Proteinase Inhibitory Activities of Selected Poppy (*Papaver somniferum* L.) Genotypes. *Chem. Biodivers*, 2017; 14: e1700176. DOI: 10.1002/cbdv.201700176
- 71. Kirkan, B., Ozer, M.S., Sarikurkcu, C., Copuroglu, M., Cengiz, M., Tepe, B., Can the stalks of *Papaver somniferum* L. be an alternative source of bioactive components?. *Ind. Crops Prod.*, 2018; *115*: 1-5. DOI: 10.1016/j.indcrop.2018.02.023
- 72. Mika, E.S., Studies on the growth and development and morphine content of opium poppy. *Bot. Gaz.*, 1955; *116*: 323-339. DOI: 10.1086/335877

- 73. Sengupta, A., Mazumder, U.K., Triglyceride composition of *Papaver somniferum* seed oil. *J. Sci. Food Agric.*, 1976; 27: 214-218. DOI: 10.1002/jsfa.2740270303
- 74. Wickström, K., Widen, C.J., Pyysalo, H., Salemink, C.A., Alkaloid formation of *Papaver somniferum* in Finland. *Annal. Bot. Fennici*, 1984; 21: 201-208. [Journal Website]
- 75. Yasmin, R., Naru, A.M., Biochemical analysis of *Papaver somniferum* (opium poppy). *Biochem. Soc. Trans.*, 1991; 19: 436S. DOI: 10.1042/bst019436s
- 76. Bajpai, S., Prajapati, S., Luthra, R., Sharma, S., Naqvi, A., Kumar, S., Variation in the seed and oil yields and oil quality in the Indian germplasm of opium poppy *Papaver somniferum*. *Gen. Resour. Crop Evol.*, 1999; 46: 435-439. DOI: 10.1023/A:1008753604907
- 77. Bilková, A., Bezakova, L., Bilka, F., Psenak, M., An amine oxidase in seedlings of *Papaver somniferum* L. *Biol, Plant.*, 2005; 49: 389-394. DOI: 10.1007/s10535-005-0013-x
- 78. Özcan, M.M., Atalay, Ç., Determination of seed and oil properties of some poppy (*Papaver somniferum* L.) varieties. *Grasas aceites*, 2006; *57*: 169-174. DOI: 10.3989/gya.2006.v57.i2.33
- 79. Dittbrenner, A., Mock, H.P., Börner, A., Lohwasser, U., Variability of alkaloid content in *Papaver somniferum* L. *J. Appl. Bot. Food Qual*, 2009; 82: 103-107. [ResearchGate]
- 80. Ivan, S., Jozef, F., Content of heavy metals in poppy seeds (*Papaver somniferum* L.). *Adv. Environ. Biol.*, 2011; 5: 315-319. [ResearchGate]
- 81. Khan, M.R., Khawar, Z., Latif, M., Biochemical Investigation of Oil of *Papaver somniferum*. *Asian J. Chem.*, 2012; 24: 4476-4478. [ResearchGate]
- 82. Rahimi, A., Arslan, N., Ahmed, H. A. Variation in Oil, Protein Content and Fatty Acid Composition of Twelve Turkish Opium Poppy (*Papaver somniferum* L.) Lines. *J. Med. Plants By-prod.*, 2012; *1:* 49-54. DOI: 10.22092/jmpb.2012.108447
- 83. Meos, A., Saks, L., Raal, A., Content of alkaloids in ornamental *Papaver somniferum* L. cultivars growing in Estonia. *Proc. Estonian Acad. Sci.*, 2017; 66: 34-39. DOI: 10.3176/proc.2017.1.04
- 84. Carlin, M.G., Dean, J.R., Ames, J.M., Opium Alkaloids in Harvested and Thermally Processed Poppy Seeds. *Front. Chem.*, 2020; 8: 737. DOI: 10.3389/fchem.2020.00737
- 85. Lainer, J., Dawid, C., Dunkel, A., Gläser, P., Wittl, S., Hofmann, T., Characterization of Bitter-Tasting Oxylipins in Poppy Seeds (*Papaver somniferum* L.). *J. Agric. Food Chem.*, 2020; 68: 10361-10373. DOI: 10.1021/acs.jafc.9b06655

- 86. Dąbrowski, G., Czaplicki, S., Konopka, I., Composition and quality of poppy (*Papaver somniferum* L.) seed oil depending on the extraction method. *LWT*., 2020; *134*: 110167. DOI: 10.1016/j.lwt.2020.110167
- 87. Luhmer, K., Schulze-Kaysers, N., Feuereisen, M., Wirth, L., Maretzky, F., Wüst, M., Blum, H., Dörr, E., Pude, R., Fatty Acid Composition, Tocopherols, Volatile Compounds, and Sensory Evaluation of Low Morphine Yielding Varieties of Poppy (*Papaver somniferum* L.) Seeds and Oils. *J. Agric. Food Chem.*, 2021; 69: 3439-3451. DOI: 10.1021/acs.jafc.0c07183
- 88. Májer, P., Németh, É.Z., Alkaloid Accumulation and Distribution within the Capsules of Two Opium Poppy (*Papaver somniferum* L.) Varieties. *Plants.*, 2024; *13*: 1640. DOI: 10.3390/plants13121640
- 89. Muhizi, S., Kim, I.H., Effect of dietary poppy (*Papaver somniferum* L.) seed meal supplementation on growth performance, nutrient digestibility, faecal microbiota and blood profile in growing-finishing pigs. *J. Anim. Feed Sci.*, 2021; *30*: 58-63. DOI: 10.22358/jafs/133958/2021
- 90. Sakenin Chelav, H., Khashaveh, A., Insecticidal activity of Poppy (*Papaver somniferum* L.) seed oil against cowpea weevil (*Callosobruchus maculatus* F.) in stored cowpea, *Arch. Phytopathol. Plant Protec*, 2013; 46: 2314-2322. DOI: 10.1080/03235408.2013.792599
- 91. Gupcsó, K., Kókai, Z., Bálint, M., Tavaszi-Sárosi, S., Németh, É.Z., Studies on Sensory and Phytochemical Characteristics of Poppy (*Papaver somniferum* L.) Varieties for Their Oil Utilisation. *Foods.*, 2023; *12*: 3165. DOI: 10.3390/foods12173165
- 92. Moneo, I., Alday, E., Ramos, C., Curiel, G., Occupational asthma caused by *Papaver somniferum*. *Allergol*. *Immunopathol*, 1993; 21: 145-148. [PubMed]
- 93. Avci, F.G., Atas, B., Aksoy, C.S., Kurpejovic, E., Toplan, G.G., Gurer, C., Guillerminet, M., Orelle, C., Jault, J.M., Akbulut, B.S., Repurposing bioactive aporphine alkaloids as efflux pump inhibitors. *Fitoterapia*, 2019; *139*: 104371. DOI: 10.1016/j.fitote.2019.104371
- 94. Bayazeid, O., Yalçin, F.N., Biological targets of 92 alkaloids isolated from *Papaver* genus: a perspective based on in silico predictions. *Med. Chem. Res.*, 2021; *30*: 574-585. DOI: 10.1007/s00044-020-02663-9
- 95. Ziegler, J., Voigtländer, S., Schmidt, J., Kramell, R., Miersch, O., Ammer, C., Gesell, A., Kutchan, T.M., Comparative transcript and alkaloid profiling in *Papaver* species

- identifies a short chain dehydrogenase/reductase involved in morphine biosynthesis. *Plant J.*, 2006; 48: 177-192. DOI: 10.1111/j.1365-313X.2006.02860.x
- 96. Laughlin, J.C., The effect of time of harvest on the yield components of poppies (*Papaver somniferum* L.). *J. Agric. Sci.*, 1980; 95: 667-676. DOI: 10.1017/S0021859600088067
- 97. Laughlin, J.C., Munro, D., The effect of fungal colonization on the morphine production of poppy (*Papaver somniferum* L.) capsules. *J. Agric. Sci.*, 1982; 98: 679-687. DOI: 10.1017/S0021859600054484
- 98. Hofman, P.J., Menary, R.C., Alkaloid losses from the capsules of *Papaver somniferum* L. during kiln drying and storage under commercial conditions in Tasmania. *J. Stored Prod. Res.*, 1985; 2: 135-139. DOI: 10.1016/0022-474X(85)90007-4
- 99. Luthra, R., Singh, N., Changes in fatty acid composition accompanying the deposition of triacylglycerols in developing seeds of opium poppy (*Papaver somniferum* L.). *Plant Sci.*, 1989; 60: 55-60. DOI: 10.1016/0168-9452(89)90043-5
- 100. Subrahmanyam, K., Verma, R.K., Naqvi, A.A., Singh, D.V., Effect of Forms of Sulphur on Yield and Quality of Seed, Oil and Alkaloids of Opium Poppy (*Papaver somniferum* L.). Acta Hortic, 1992; 306: 431-435. DOI: 10.17660/ActaHortic.1992.306.57
- 101. Laughlin, J.C., Chung, B., Nitrogen and Irrigation Effects on the Yield of Poppies (*Papaver somniferum* L.). Acta Hortic., 1992; *306*: 466-473. DOI: 10.17660/ActaHortic.1992.306.63
- 102. Chizzola, R., Micronutrient composition of Papaver somniferum L. grown under low cadmium stress condition. *J. Plant Nutr.*, 2001; 24: 1663-1677. DOI: 10.1081/pln-100107305
- 103. Grothe, T., Lenz, R., Kutchan, T.M., Molecular characterization of the salutaridinol 7-O-acetyltransferase involved in morphine biosynthesis in opium poppy *Papaver somniferum*. *J. Biol. Chem.*, 2001; 276: 30717-30723. DOI: 10.1074/jbc.M102688200
- 104. Németh, É., Bernáth, J., Sztefanov, A., Petheö, F., New results of poppy (*Papaver somniferum* L.) breeding for low alkaloid content in Hungary. *Acta Hortic*, 2002; *576*: 151-158. DOI: 10.17660/ActaHortic.2002.576.22
- 105. Balažová, A., Bilka, F., Blanáriková, V., Pšenák, M., Changes in Sanguinarine Content and Polyphenoloxidase Activity Due to a FungalElicitor in Suspension Cultures of the Opium Plant *Papaver somniferum* L. Čes. Slov. Farm, 2002; 51: 182-185. [ResearchGate]

- 106. Fisinger, U., Grobe, N., Zenk, M.H., Thebaine synthase: a new enzyme in the morphine pathway in Papaver somniferum. *Nat. Prod. Commun.*, 2007; 2: 249-253. DOI: 10.1177/1934578X0700200305
- 107. Khan, R., Khan, M.M., Singh, M., Nasir, S., Naeem, M., Siddiqui, M.H., Mohammad, F., Gibberellic acid and triacontanol can ameliorate the opium yield and morphine production in opium poppy (*Papaver somniferum* L.). *Acta Agric. Scand. Sect. B-Soil Plant Sci.*, 2007; 57: 307-312. DOI: 10.1080/09064710600982811
- 108. Mahdavi-Damghani, A., Kamkar, B., Al-Ahmadi, M.J., Testi, L., Muñoz-Ledesma, F.J., Villalobos, F.J., Water stress effects on growth, development and yield of opium poppy (*Papaver somniferum* L.). *Agric. Water Manag.*, 2010; 97: 1582-1590. DOI: 10.1016/j.agwat.2010.05.011
- 109. Holkova, I., Bilka, F., Rauova, D., Bezakova, L., Purification and properties of lipoxygenase from opium poppy seedlings (*Papaver somniferum L.*). *Turk. J. Biol.*, 2016; *40:* 772-780. DOI: 10.3906/biy-1507-149
- 110. Kara, N., The effects of autumn and spring sowing on yield, oil and morphine contents in the Turkish poppy (*Papaver somniferum* L.) cultivars. *Turk. J. Field Crops.*, 2017; 22: 39-46. DOI: 10.17557/tjfc.301829
- 111. Holková, I., Rauová, D., Mergová, M., Bezáková, L., Mikuš, P., Purification and Product Characterization of Lipoxygenase from Opium Poppy Cultures (*Papaver somniferum* L.). Molecules, 2019; *24*: 4268. DOI: 10.3390/molecules24234268
- 112. Verma, R.K., Kumar, R., Sankhla, M.S., Forensic identifications of *Papaver Somniferum* (Opium) for fatal poisoning. *Int. Medico-Legal Rep. J.*, 2019; *1*: 110-118. DOI: 10.2139/ssrn.3483979
- 113. Satranský, M., Fraňková, A., Kuchtová, P., Pazderů, K., Capouchová, I., Oil content and fatty acid profile of selected poppy (*Papaver somniferum* L.) landraces and modern cultivars. *Plant Soil Environ*, 2021; 67: 579-587. DOI: 10.17221/316/2021-PSE
- 114. Yazici, L., Influence of different sowing times on yield and biochemical characteristics of different opium poppy (*Papaver somniferum* L.) genotypes. *J. King Saud Univ. Sci.*, 2022; *34*: 102337. DOI: 10.1016/j.jksus.2022.102337
- 115. Neugschwandtner, R.W., Dobos, G., Wagentristl, H., Lošák, T., Klimek-Kopyra, A., Kaul, H-P., Yield and Yield Components of Winter Poppy (*Papaver somniferum* L.) Are Affected by Sowing Date and Sowing Rate under Pannonian Climate Conditions. *Agriculture*, 2023; *13*: 997. DOI: 10.3390/agriculture13050997

- 116. Aghaali, Z., Naghavi, M.R., Zargar, M., Promising approaches for simultaneous enhancement of medicinally significant Benzylisoquinoline alkaloids in opium poppy. *Front. Plant Sci.*, 2024; *15*: 1377318. DOI: 10.3389/fpls.2024.1377318
- 117. Carr, S.C., Rehman, F., Hagel, J.M., Chen, X., Ng, K.K., Facchini, P.J., Two ubiquitous aldo-keto reductases in the genus *Papaver* support a patchwork model for morphine pathway evolution. *Commun. Biol.*, 2024; 7: 1410. DOI: 10.1038/s42003-024-07100-w
- 118. Varga, I., Moslavac, T., Flanjak, I., Iljkic, D., Pospišil, M., Loncaric, Z., Antunovic, M., White-Seeded Culinary Poppy (*Papaver somniferum* L.) Se Biofortification: Oil Quality, Fatty Acid Profile, and Seed Yield. *Plants.*, 2025; *14:* 95. DOI: 10.3390/plants14010095
- 119. Apaydin, D., Investigation of γ-irradiation effects on physicochemical characteristics, fatty acid and sterol profiles of three different poppy (*Papaver somniferum* L.) seed varieties. *Grasas Aceites*, 2025; 76: 2226. DOI: 10.3989/gya.0871241.2226
- 120. Diaz-Barcena, A., Giraldo, P., Exploring the research evolution of *Papaver somniferum* and *Cannabis sativa*: A bibliometric comparative analysis. *Ind. Crops Prod.*, 2023; 203: 117143. DOI: 10.1016/j.indcrop.2023.117143
- 121. Beycioğlu, T., Bibliometric Analysis of Poppy Alkaloids and Morphine Biosynthesis. *Black Sea J. Agric.*, 2025; 8: 11-12. DOI: 10.47115/bsagriculture.1692279
- 122. Kunwar, R.M., Bussmann, R.W., Ethnobotany in the Nepal Himalaya. *J. Ethnobiol, Ethnomed*, 2008; *4*: 24. DOI: 10.1186/1746-4269-4-24
- 123. Stermitz, F.R., Rapoport, H., The Biosynthesis of Opium Alkaloids. Alkaloid Interconversions in *Papaver somniferum* and *P. orientale. J. Am. Chem. Soc.*, 1961; 83: 4045-4050. DOI: 10.1021/ja01480a022
- 124. Victor, L., Are traditional medicinal plants and ethnobotany still valuable approaches in pharmaceutical research? *Bol. Latinoam. Caribb. Plantas Med. Aromát*, 2011; *10:* 3-10. [ResearchGate]
- 125. Vijayaraghavan, K., Nalini, S.K., Prakash, N.U., Madhankumar, D., One step green synthesis of silver nano/microparticles using extracts of *Trachyspermum ammi* and *Papaver somniferum*. *Colloids Surf. B Biointerfaces*, 2012; 94: 114-117. DOI: 10.1016/j.colsurfb.2012.01.026
- 126. Shehzad, A., Tariq, F., Al Saidi, A.K., Khan, K.A., Khan, S.A., Alshammari, F.H., Ul-Islam, M., Green-synthesized silver nanoparticle-infused PVA hydrogels: A sustainable solution for skin repair. *Results Chem.*, 2025; *17*: 102584. DOI: 10.1016/j.rechem.2025.102584

- 127. Wali, M., Sajjad, A.S., Sumaira, S., Muhammad, N., Safia, H., Muhammad, J., Green synthesis of gold nanoparticles and their characterizations using plant extract of *Papaver somniferum. Nano Sci. Nano Technol.*, 2017; 11: 118. [ResearchGate]
- 128. Lakhali, H., Baştaş, S., Türkben, A.B., Ceyhan, A.A., Eco-friendly green synthesis of Co3O4-NiO nano catalysts from Papaver somniferum biomass for efficient NaBH4 Hydrolysis: Advancing circular bioeconomy and clean hydrogen energy 2025; 202: 108240. DOI: conversion. Biomass Bioenergy, 10.1016/j.biombioe.2025.108240
- 129. Muhammad, W., Khan, M.A., Nazir, M., Siddiquah, A., Mushtaq, S., Hashmi, S.S., Abbasi, B.H., *Papaver somniferum* L. mediated novel bioinspired lead oxide (PbO) and iron oxide (Fe₂O₃) nanoparticles: In-vitro biological applications, biocompatibility and their potential towards HepG2 cell line. *Mater. Sci. Eng. C.*, 2019; *103*: 109740. DOI: 10.1016/j.msec.2019.109740
- 130. Muhammad, W., Ullah, N., Haroon, M., Abbasi, B.H., Optical, morphological and biological analysis of zinc oxide nanoparticles (ZnO NPs) using *Papaver somniferum* L. *RSC Adv.*, 2019; *9*: 29541-29548. DOI: 10.1039/c9ra04424h
- 131. Kadhim, A.A., Abbas, N.R., Kadhum, H.H., Albukhaty, S., Jabir, M.S., Naji, A.M., Hamzah, S.S., Mohammed, M.K., Al-Karagoly, H., Investigating the effects of biogenic zinc oxide nanoparticles produced using *Papaver somniferum* extract on oxidative stress, cytotoxicity, and the induction of apoptosis in the THP-1 cell line. *Biol. Trace Elem. Res.*, 2023; 201: 4697-4709. DOI: 10.1007/s12011-023-03574-7
- 132. Küçüker, H., Aydıner, F., Fatal Poppy Capsule Toxicity. *Eur. J. Gen. Med.*, 2011; *8:* 335-337. DOI: 10.29333/ejgm/82766
- 133. Bozan, G., Ulukapi, H., Oncul, U., Levent, S., Dinleyici, E.C., A Fatal Case of Opioid Intoxication After Raw Poppy Plant Ingestion. *Cureus.*, 2021; 13: e13176. DOI: 10.7759/cureus.1317
- 134. Tan, Y.P., Alexander, P.D., Knowles, S., Acute cardiotoxicity following 'poppy seed tea' consumption. *Anaesth. Rep.*, 2021; 9: e12130. DOI: 10.1002/anr3.12130
- 135. Hoummani, H., Benlamkaddem, S., Achour, S., Poppy intoxication in children: The interest of early administration of naloxone. *Toxicol. Anal. Clin.*, 2025; *37*: 111-114. DOI: 10.1016/j.toxac.2024.09.014
- 136. Yoshimatsu, K., Kiuchi, F., Shimomura, K., Makino, Y., A rapid and reliable solidphase extraction method for high-performance liquid chromatographic analysis of

- opium alkaloids from *Papaver* plants. *Chem. Pharm. Bull.*, 2005; 53: 1446-1450. DOI: 10.1248/cpb.53.1446
- 137. Brook, K., Bennett, J., Desai, S.P., The Chemical History of Morphine: An 8000-year Journey, from Resin to de-novo Synthesis. J. Anesth. Hist., 2017; 3: 50-55. DOI: 10.1016/j.janh.2017.02.001
- 138. Teunissen, A.J., van Gastel, L., Stolker, R.J., Koopman, S.A., The use of intrathecal morphine in non-abdominal surgery: a scoping review. BJA Open., 2025; 14: 100387. DOI: 10.1016/j.bjao.2025.100387
- 139. Megens, A.A., Artois, K., Vermeire, J., Meert, T., Awouters, F.H., Comparison of the analgesic and intestinal effects of fentanyl and morphine in rats. J. Pain Symptom Manag., 1998; 15: 253-257. DOI: 10.1016/s0885-3924(97)00371-0
- 140. Song, X., Zhang, Y., Liu, Y., Chen, G., Zhao, L., Enhanced Analgesic Efficacy and Reduced Side Effects of Morphine by Combination with PD-1 Agonist. ACS Chem. Neurosci, 2025; 16: 490-499. DOI: 10.1021/acschemneuro.4c00732
- 141. Tatum, A.L., Seevers, M.H., Collins, K.H., Morphine addiction and its physiological interpretation based on experimental evidences. J. Pharmacol. Exp. Therap., 1929; 36: 447-475. DOI: 10.1016/S0022-3565(25)04096-0
- 142. Listos, J., Łupina, M., Talarek, S., Mazur, A., Orzelska-Górka, J., Kotlińska, J., The Mechanisms Involved in Morphine Addiction: An Overview. Int. J. Mol. Sci., 2019; 20: 4302. DOI: 10.3390/ijms20174302
- 143. Glare, P., Walsh, D., Sheehan, D., The adverse effects of morphine: a prospective survey of common symptoms during repeated dosing for chronic cancer pain. Am. J. Hosp. Palliat. Care., 2006; 23: 229-235. DOI: 10.1177/1049909106289068
- 144. Dunlap, C.E., Leslie, F.M., Effect of ascorbate on the toxicity of morphine in mice. Neuropharmacol, 1985; 24: 797-804. DOI: 10.1016/0028-3908(85)90015-2
- 145. Gonçalves, F., Morphine toxicity in renal failure. J. Opioid Manag., 2006; 2: 174-176. DOI: 10.5055/jom.2006.0027
- 146. Jalili, C., Makalani, F., Roshankhah, S., Sohrabi, K., Salahshoor, M.R., Protective Effect of Resveratrol Against Morphine Damage to Kidneys of Mice. Int. J. Morphol., 2017; 35: 1409-1415. DOI: 10.4067/S0717-95022017000401409
- 147. Noufal, Y., Kringel, D., Toennes, S.W., Dudziak, R., Lötsch, J., Pharmacological data science perspective on fatal incidents of morphine treatment. *Pharmacol. Ther.*, 2023; 241: 108312. DOI: 10.1016/j.pharmthera.2022.108312

- 148. Olukiran, O.S., Ogundipe, O.J., Adelodun, S.T., Hamed, M.A., Babatunde, O.S., Alese, O.O., Akomolafe, R.O., 2025. Sex and Dose Differences in Morphine Administration on Renal Function, Inflammatory and Apoptotic Markers in Male and Female Wistar Rats. *Curr. Res. Physiol*, 2025; 8: 100162. DOI: 10.1016/j.crphys.2025.100162
- 149. Dutra, R.C., Campos, M.M., Santos, A.R., Calixto, J.B., Medicinal plants in Brazil: Pharmacological studies, drug discovery, challenges and perspectives. *Pharmacol. Res.*, 2016; *112*: 4-29. DOI: 10.1016/j.phrs.2016.01.021
- 150. Goldberg, A.A., Shapero, M., A comparative study of the spasmolytic activities of Octaverine, Perparine and Papaverine. *J. Pharm. Pharm.*, 1954; 6: 171-177. DOI: 10.1111/j.2042-7158.1954.tb10933.x
- 151. Aggarwal, M., Leser, G.P., Lamb, R.A., Repurposing Papaverine as an antiviral agent against influenza viruses and paramyxoviruses. *J. Virol.*, 2020; *94:* e01888-19. DOI: 10.1128/JVI.01888-19
- 152. Valipour, M., Irannejad, H., Emami, S., Papaverine, a promising therapeutic agent for the treatment of COVID-19 patients with underlying cardiovascular diseases (CVDs). *Drug Dev. Res.*, 2022; 83: 1246-1250. DOI: 10.1002/ddr.21961
- 153. Pan, D., Yang, J., Liu, M., Xu, Y., Feng, J., Li, H., Luo, W., He, B., Xiao, S., Yang, X., Tang, Y., Efficacy of Papaverine to Prevent Radial Artery Spasm During Transradial Cerebral Angiography (PASS): rationale and design. *Stroke Vasc. Neurol.*, 2025; 1-6. DOI:10.1136/svn-2024-003659
- 154. Leem, Y.H., Park, J.S., Park, J.E., Kim, D.Y., Kang, J.L., Kim, H.S., Papaverine inhibits α-synuclein aggregation by modulating neuroinflammation and matrix metalloproteinase-3 expression in the subacute MPTP/P mouse model of Parkinson's disease. *Biomed. Pharmacother*, 2020; *130*: 110576. DOI: 10.1016/j.biopha.2020.110576
- 155. Ashrafi, S., Alam, S., Sultana, A., Raj, A., Emon, N.U., Richi, F.T., Sharmin, T., Moon, M., Park, M.N., Kim, B., Papaverine: a miraculous alkaloid from opium and its multimedicinal application. *Molecules*, 2023; 28: 3149. DOI: 10.3390/molecules28073149
- 156. Levine, S.B., Althof, S.E., Turner, L.A., Risen, C.B., Bodner, D.R., Kursh, E.D., Resnick, M.I., Side effects of self-administration of intracavernous Papaverine and phentolamine for the treatment of impotence. *J. Urol.*, 1989; *141:* 54-57. DOI: 10.1016/s0022-5347(17)40585-4

- 157. Kahramaner, Z., Erdemir, A., Türkoğlu, E., Coşar, H., Sütçüoğlu, S., Özer, E.A., Papaverine intoxication in a newborn: an unusual case report. Turk. J. Pediatr., 2014; 56: 532-534. [Google Scholar]
- 158. Davila, J.C., Reddy, C.G., Davis, P.J., Acosta, D., Toxicity assessment of Papaverine hydrochloride and papaverine-derived metabolites in primary cultures of rat hepatocytes. In Vitro Cell. Dev. Biol., 1990; 26: 515-524. DOI: 10.1007/BF02624095
- 159. Zhou, X., Alambyan, V., Ostergard, T., Pace, J., Kohen, M., Manjila, S., Ramos-Estebanez, C., Prolonged intracisternal Papaverine toxicity: index case description and proposed mechanism of action. World Neurosurg, 2018; 109: 251-257. DOI: 10.1016/j.wneu.2017.09.196
- 160. Al-Sharshahi, Z.F., Hoz, S.S., Almurayati, M.E., Kareem, Z.M., Ameen, Z., Intracisternal Papaverine toxicity in anterior circulation aneurysm clipping surgery: A literature review. Rom. Neurosurg, 2020; 34: 386-390. DOI: 10.33962/roneuro-2020-060
- 161. Vincent, P.G., Bare, C.E., Gentner, W.A., Thebaine content of selections of *Papaver* bracteatum Lindl. at different ages. J. Pharm. Sci., 1977; 66: 1716-1719. DOI: 10.1002/jps.2600661215
- 162. Palevitch, D., Levy, A., Domestication of *Papaver bracteatum* as a source of Thebaine. Acta Hortic, 1992; 306: 33-52. DOI: 10.17660/ActaHortic.1992.306.2
- 163. Onoyovwe, A., Hagel, J.M., Chen, X., Khan, M.F., Schriemer, D.C., Facchini, P.J., Morphine biosynthesis in opium poppy involves two cell types: sieve elements and laticifers. Plant Cell., 2013; 25: 4110-4122. DOI: 10.1105/tpc.113.115113
- 164. Kodaira, H., Spector, S., Transformation of thebaine to Oripavine, Codeine, and Morphine by rat liver, kidney, and brain microsomes. Proc. Natl. Acad. Sci. USA., 1988; 85: 1267-1271. DOI: 10.1073/pnas.85.4.1267
- 165. Li, X., Krysiak-Baltyn, K., Richards, L., Jarrold, A., Stevens, G.W., Bowser, T., Speight, R.E., Gras, S.L., High-Efficiency Biocatalytic Conversion of Thebaine to Codeine. ACS Omega., 2020; 5: 9339-9347. DOI: 10.1021/acsomega.0c00282
- 166. Carlin, M.G., Dean, J.R., Bookham, J.L., Perry, J.J., Investigation of the acid/base behaviour of the opium alkaloid Thebaine in LC-ESI-MS mobile phase by NMR spectroscopy. R. Soc. Open Sci., 2017; 4: 170715. DOI: 10.1098/rsos.170715
- 167. Cassella, G., Wu, A.H., Shaw, B.R., Hill, D.W., The analysis of Thebaine in urine for the detection of Poppy seed consumption. J. Anal. Toxicol., 1997; 21: 376-383. DOI: 10.1093/jat/21.5.376

- 168. Lee, S., Park, Y., Han, E., Choi, H., Chung, H., Oh, S.M., Chung, K.H., Thebaine in hair as a marker for chronic use of illegal opium poppy substances. *Forensic Sci. Int.*, 2011; 204: 115-118. DOI: 10.1016/j.forsciint.2010.05.013
- 169. BfR., Poppy seeds in food: The content of opium alkaloid Thebaine should be reduced as much as possible, 2018, Opinion No 039/2018. DOI: 10.17590/20191120-092911
- 170. Sloan, J.W., Brooks, J.W., Eisenman, A.J., Martin, W.R., Comparison of the effects of single doses of Morphine and Thebaine on body temperature, activity, and brain and heart levels of catecholamines and serotonin. *Psychopharmacologia*, 1962; *3*: 291-301. DOI: 10.1007/BF00411369
- 171. Eisenreich, A., Sachse, B., Gürtler, R., Dusemund, B., Lindtner, O., Schäfer, B., What do we know about health risks related to Thebaine in food?. *Food Chem.*, 2020; *309*: 125564. DOI: 10.1016/j.foodchem.2019.125564
- 172. Navarro, G., Elliott, H.W., The effects of Morphine, Morphine and Thebaine on the EEG and behavior of rabbits and cats. *Neuropharmacology*, 1971; *10*: 367-377. DOI: 10.1016/0028-3908(71)90065-7
- 173. Penafiel, R., Yoo, D., Turner, C., Brown, J.A., McDonald, C., Tran, J., Shaw, V., Roberts, D.M., Toxicokinetics of Thebaine in those consuming non-food grade poppy seeds as a tea. *Clin. Toxicol*, 2023; *61*: 644-648. DOI: 10.1080/15563650.2023.2271163