

## UNLOCKING THE POWER OF MAGNESIUM: A SYSTEMIC REVIEW AND META ANALYSIS REGARDING ITS ROLE IN METABOLIC DISORDER [DIABETES]

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### ABSTRACT

Magnesium is an important micronutrient that regulates glucose homeostasis, insulin signaling, and energy metabolism. New research reveals that magnesium insufficiency is widespread in people with metabolic disorders, notably type 2 diabetes mellitus (T2DM), and may contribute to illness development and progression. The purpose of this systematic review and meta-analysis was to assess the relationship between magnesium consumption or supplementation and diabetes-related metabolic outcomes. Electronic databases such as PubMed, Scopus, and Web of Science were rigorously searched for observational studies and randomized controlled trials (RCTs) that examined dietary magnesium intake, serum magnesium levels, or magnesium supplementation in connection to diabetes risk and glycemic control. Incidence of type 2 diabetes, fasting plasma glucose, insulin resistance indicators, HbA1c, lipid profile, and blood pressure were among the outcomes of interest. The risk of acquiring type 2

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diabetes is inversely correlated with dietary magnesium consumption, according to data from prospective cohort studies; dose-response analyses show that increased intake significantly reduces risk. Although effects on HbA1c were modest and varied, meta-analyses of RCTs showed that magnesium supplementation significantly improved insulin sensitivity and fasting blood glucose, especially in people with magnesium deficiency or poor glycemic

control. Benefits also included improvements in blood pressure and cholesterol profiles, which may indicate more extensive cardio metabolic protection. Enhanced insulin receptor function, better glucose transport, decreased inflammation, and oxidative stress control are some of the suggested causes. Magnesium has been shown to have a positive function in the prevention and treatment of metabolic diseases, despite variations in study design, dosage, and duration. Making sure you consume enough magnesium through your diet and using specific supplements could be a low-risk, safe supplemental approach to diabetes treatment. More extensive, long-term studies are necessary to determine the best dosage and clinical recommendations.

**KEYWORDS:** Magnesium, micronutrient, diabetes, supplementation.

## INTRODUCTION

Over time, global populations have seen a reduction in micronutrient intake due to changing dietary patterns. Among these nutrients,<sup>[1]</sup> magnesium (Mg)—one of the body's most abundant intracellular ions—plays a central role in essential biochemical processes. A lack of Mg can trigger serious biochemical and clinical imbalances in the human body.<sup>[2,3]</sup>

Diabetes mellitus (DM), a metabolic condition marked by persistent hyperglycaemia,<sup>[4]</sup> is closely associated with reduced Mg levels in both intracellular and extracellular compartments.<sup>[5]</sup> Low serum Mg (hypomagnesemia) has been linked to insulin resistance, while chronic hyperglycaemia worsens Mg loss, fueling a self-perpetuating cycle that drives both microvascular and macrovascular complications of diabetes.<sup>[6,7,8,9]</sup>

Although the precise mechanisms connecting DM and hypomagnesemia are not fully understood, metabolic studies indicate that Mg supplementation may enhance insulin sensitivity and improve glucose regulation.<sup>[1,10]</sup>

This review synthesizes evidence from the literature (1990–2004) on the Mg–DM connection, outlining established findings and highlighting unresolved debates. The following section expands on mechanisms, clinical significance, and more recent insights into this relationship.

## 1. Mechanisms Linking Magnesium Deficiency and Diabetes

### Tissue-specific molecular roles of magnesium

Magnesium ( $Mg^{2+}$ ) acts as a cofactor for hundreds of enzymes and is central to a wide range of cellular activities. For example, all kinases require the ATP– $Mg^{2+}$  complex to transfer phosphate groups during cell signalling.<sup>[11]</sup>  $Mg^{2+}$  also plays a role in lipid and protein metabolism.<sup>[12]</sup> as well as electrolyte balance, by activating ATP-dependent pumps such as  $Na^+/K^+$ ,  $Na^+/Ca^{2+}$ ,  $Na^+/Mg^{2+}$ , and  $Mg^{2+}/Ca^{2+}$  transporters.

Beyond these functions,  $Mg^{2+}$  regulates cell growth by supporting DNA synthesis and repair, since it serves as a cofactor for multiple DNA repair enzymes.<sup>[13]</sup> Given its essential role in these fundamental processes,  $Mg^{2+}$  deficiency has been linked to several disorders, including type 2 diabetes mellitus (T2DM). The following sections outline how  $Mg^{2+}$  deficiency affects key tissues such as the pancreas, liver, and kidneys, contributing to the pathophysiology of diabetes.

### Effects of Magnesium on Pancreatic $\beta$ -Cells

Glucose regulation relies primarily on the pancreatic islets, liver, and peripheral tissues such as muscle and adipose tissue.<sup>[14]</sup> Within the islets,  $\alpha$ -cells secrete glucagon in response to low glucose, while  $\beta$ -cells release insulin when glucose is elevated.<sup>[15]</sup>

A key player in insulin secretion is the ATP-sensitive potassium (KATP) channel, composed of four Kir6.2 subunits and four SUR1 regulatory subunits.<sup>[16]</sup> Under resting conditions, the channel remains open, allowing  $K^+$  efflux. When intracellular ATP/ADP- $Mg^{2+}$  levels rise, the channel closes, leading to membrane depolarization,  $Ca^{2+}$  influx, and insulin release.<sup>[16]</sup> ADP- $Mg^{2+}$  supports channel opening via nucleotide-binding domains in SUR1, whereas ATP binding promotes closure. In magnesium deficiency, hyperpolarization and persistent channel opening block depolarization, ultimately suppressing insulin secretion.<sup>[17]</sup>

Magnesium also influences several enzymes involved in glycolysis and the Krebs cycle, including glucokinase (GCK), phosphofructokinase, and pyruvate kinase. Enhanced enzyme activity raises ATP production, which in turn closes KATP channels, depolarizes the membrane, and promotes  $Ca^{2+}$  entry through L-type calcium channels. However,  $Mg^{2+}$  can also compete with  $Ca^{2+}$  at these channels, reducing insulin release. In addition,  $Mg^{2+}$  may regulate the expression of glucose transporter 2 (GLUT2) and L-type  $Ca^{2+}$  channels, though

no effect has been observed on GCK mRNA or the genes encoding KATP subunits (KCNJ11 and ABCC8).

Experimental data further highlight the complexity of Mg<sup>2+</sup>'s role. In murine  $\beta$ -cell cultures, extracellular Mg<sup>2+</sup> reduced Ca<sup>2+</sup> uptake, suggesting that Mg<sup>2+</sup> deficiency alters Ca<sup>2+</sup> handling.<sup>[18]</sup> Moreover Murakami *et al.*<sup>[19]</sup> reported that insulin secretion triggered by KCl, forskolin, and D-glyceraldehyde increased intracellular Mg<sup>2+</sup> in RINm5F cells, an effect blocked by verapamil, an L-type Ca<sup>2+</sup> channel inhibitor.<sup>[19]</sup> These findings indicate that Mg<sup>2+</sup> deficiency may impair Ca<sup>2+</sup> transport and insulin secretion. Interestingly, suppression of TRPM7—the principal Mg<sup>2+</sup> channel in  $\beta$ -cells—enhanced insulin secretion threefold in INS-1 cells.<sup>[20]</sup> Similarly, a pilot study in healthy subjects showed that intravenous magnesium sulfate (MgSO<sub>4</sub>) infusion markedly reduced insulin levels.<sup>[18]</sup> Together, these studies suggest that Mg<sup>2+</sup> can exert inhibitory as well as supportive effects on insulin secretion, depending on context.

Regarding glucose transport, GLUT2 is the main isoform mediating glucose entry into  $\beta$ -cells. In HepG2 cultures, mild Mg<sup>2+</sup> deficiency (0.4 mM) increased GLUT2 mRNA by 250%, whereas Mg-deficient diets in rats decreased expression; in both cases, GLUT2 protein levels remained unchanged,<sup>[21]</sup> pointing to compensatory regulation. Molnes *et al.*<sup>[22]</sup> also noted that low Mg<sup>2+</sup>-ATP concentrations might dampen the cooperative kinetics of GCK toward glucose. By contrast, Gommers *et al.*<sup>[20]</sup> found that reduced extracellular Mg<sup>2+</sup> did not affect glucose-stimulated insulin release or GCK expression in mouse islets and INS-1 cells.

Overall, the relationship between Mg<sup>2+</sup> and insulin secretion in  $\beta$ -cells remains unresolved. Conflicting evidence suggests both stimulatory and inhibitory effects, underscoring the need for further studies to clarify whether Mg<sup>2+</sup> supplementation could enhance insulin release in type 2 diabetes.

### **Effects of Magnesium on Insulin Action in the Liver**

The liver plays a central role in glucose homeostasis, being directly exposed to high glucose levels under the influence of both glucagon and insulin.<sup>[14]</sup> During fasting, hepatic glucose production is maintained through glycogenolysis and gluconeogenesis, while most glucose utilization (75–80%) occurs in non-insulin-dependent tissues such as the brain, intestine, erythrocytes, muscle, and adipose tissue.<sup>[23]</sup> After a meal, plasma glucose rises, prompting insulin release from pancreatic  $\beta$ -cells and suppression of glucagon secretion.<sup>[24]</sup> Insulin

release occurs in two phases: an initial burst of mature insulin granules within the first two minutes of glucose elevation, followed by sustained release from reserve granules and de novo synthesis.<sup>[23]</sup> Insulin then suppresses hepatic and renal glucose output while promoting glucose uptake in muscle and adipose tissue, with muscle accounting for ~85% of glucose disposal. Simultaneously, insulin inhibits lipolysis, lowering free fatty acid levels and further dampening hepatic glucose production.<sup>[23]</sup>

In healthy individuals, blood glucose levels remain tightly regulated despite fluctuations in food intake.<sup>[24]</sup> In contrast, patients with type 2 diabetes mellitus (T2DM) exhibit exaggerated glucose excursions, delayed and blunted insulin secretion, and inadequate glucagon suppression.<sup>[24]</sup>

Insulin action in hepatocytes begins with its binding to the insulin receptor (IR),<sup>[25]</sup> a heterotetramer of two extracellular  $\alpha$ -subunits (ligand-binding) and two transmembrane  $\beta$ -subunits (tyrosine kinase activity).<sup>[25]</sup> Upon binding, autophosphorylation of  $\beta$ -subunits recruits insulin receptor substrates (IRS-1 to IRS-4), initiating multiple downstream signaling cascades.

Among these, the PI3K–Akt pathway is essential for metabolic control, while the Ras–MAPK system controls cell growth.<sup>[26]</sup> 3-phosphoinositide-dependent protein kinase 1 (PDK1) is activated by PIP3 and phosphorylates and causes protein kinase B (AKT) to become active. By controlling many proteins, AKT activation has a variety of impacts. One of these is the 160 kDa AKT substrate protein (AS160), which controls the membrane translocation of glucose transporter 4 (GLUT4).<sup>[26]</sup> AKT encourages the production of glycogen by Glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ), a kinase that inhibits glycogen synthase (GS), is phosphorylated and inhibited.<sup>[26]</sup> Through transcription factors including Forkhead Box O1 (FOXO1) and the binding protein to the sterol regulatory element 1c (SREBP1c), AKT also controls the expression of genes linked to metabolism and survival. AKT adversely regulates FOXO1, encourages the production of gluconeogenic proteins in the liver, such as glucose 6 phosphatase (G6Pase) and phosphoenolpyruvate carboxykinase (PEPCK).<sup>[26]</sup> Additionally, insulin promotes glycogen formation by stimulating phosphofructokinase (PFK) and glycogen synthase while suppressing G6Pase activity.<sup>[23]</sup> Similarly, insulin inhibits the lipolysis of stored triacylglycerols and stimulates fatty acid production in the liver.<sup>[23]</sup>

### Magnesium's Role in Hepatic Insulin Sensitivity

Magnesium ( $Mg^{2+}$ ) has been reported to exert insulin-mimetic effects in the liver. Etweibi<sup>[21]</sup> found that HepG2 cells cultured in low  $Mg^{2+}$  (0.4 mM) exhibited reduced ATP content and diminished insulin-mediated glucose uptake compared with cells grown under physiological  $Mg^{2+}$  (0.8 mM).

**Insulin receptor regulation:**  $Mg^{2+}$  may positively influence IR expression and activity. In rats,  $Mg^{2+}$  deficiency increased hepatic IR expression, whereas  $Mg^{2+}$  supplementation in diabetic rats enhanced IR expression in skeletal muscle. Supplementation also improved receptor binding affinity and capacity. In vitro studies indicate that  $Mg^{2+}$  levels modulate receptor tyrosine kinase activity. In vivo,  $Mg^{2+}$  deficiency reduces IR autophosphorylation in liver<sup>[69]</sup> and muscle.

- **IRS expression and phosphorylation:**  $Mg^{2+}$  influences IRS regulation in a tissue- and disease-dependent manner.  $Mg^{2+}$  supplementation increased IRS-1 in skeletal muscle<sup>[27]</sup> and IRS-2 in the liver of diabetic rats, while deficiency altered IRS-1 phosphorylation in opposing directions across studies. This variability suggests that  $Mg^{2+}$  effects on IRS are context dependent and not yet fully resolved.<sup>[28]</sup>
- **Downstream signaling:**  $Mg^{2+}$  deficiency reduces Akt phosphorylation,<sup>[29]</sup> while supplementation increases Akt2 expression in diabetic rats. Supplementation also downregulates FOXO1, inhibiting gluconeogenesis<sup>[30]</sup> Similarly,  $Mg^{2+}$  deficiency increases hepatic G6Pase activity by 25%, while supplementation decreases PEPCK and G6Pase mRNA and protein expression in diabetic rats.<sup>[31]</sup> Interestingly, short-term  $Mg^{2+}$  deficiency has also been shown to reduce PEPCK expression, likely via inflammation-driven mechanisms.<sup>[32]</sup>
- **Additional pathways:**  $Mg^{2+}$  supplementation enhances GLUT4 translocation, possibly through PPAR- $\gamma$  activation, and modulates glucagon receptor expression. It may also promote glycolysis by increasing PFK-1 expression and stimulate insulin production indirectly through GLP-1.<sup>[33]</sup>

### Magnesium effort on GLUT4

GLUT4 is the primary glucose transporter in skeletal muscle and adipose tissue.<sup>[23]</sup> In diabetic rat models,  $Mg^{2+}$  supplementation significantly increased GLUT4 mRNA and protein expression, as well as its translocation to the cell membrane.<sup>[31]</sup> Another study in the same model reported a 23% rise in GLUT4 mRNA expression with  $Mg^{2+}$  supplementation—more

than double the 10% increase observed with insulin treatment alone ( $p < 0.01$ ). This finding suggests that  $Mg^{2+}$  can enhance GLUT4 expression independently of insulin secretion.<sup>[34]</sup>

Similar results have been observed across multiple type 2 diabetes (T2DM) animal models, where  $Mg^{2+}$  supplementation elevated GLUT4 expression at both the mRNA and protein level,<sup>[35,33]</sup> and in some cases amplified the effect of drugs like metformin on GLUT4 mRNA expression.<sup>[33]</sup> Mechanistically, Khosravi *et al.*<sup>[36]</sup> proposed that  $Mg^{2+}$  may regulate GLUT4 via upregulation of peroxisome proliferator-activated receptor gamma (PPAR- $\gamma$ ), a transcription factor central to glucose and lipid metabolism.

Taken together, these findings indicate that  $Mg^{2+}$  exerts a beneficial effect on GLUT4 expression and translocation, thereby supporting glucose uptake in insulin-sensitive tissues.

### Effects of Magnesium on the Kidney

In the kidney, 10–15% of  $Mg^{2+}$  reabsorption occurs in the proximal and distal convoluted tubules (DCT), while the majority (50–75%) is reabsorbed in the thick ascending limb of Henle's loop.<sup>[37]</sup> Here,  $Mg^{2+}$  reabsorption is mediated through a paracellular pathway driven by the  $Na^+/K^+/Cl^-$  cotransporter (NKCC2).  $K^+$  recycling via ROMK generates a positive luminal potential that facilitates  $Mg^{2+}$  transport through claudin-16 and claudin-19 channels.<sup>[38]</sup> In the DCT,  $Mg^{2+}$  handling is fine-tuned by TRPM6 channels, which may represent a key connection between insulin signaling and renal  $Mg^{2+}$  reabsorption.<sup>[39]</sup> Supporting this, a study in streptozotocin (STZ)-treated diabetic rats found that TRPM6 mRNA expression was elevated, and insulin treatment reduced its levels.<sup>[40]</sup>

Type 2 diabetes is commonly associated with glomerular damage, leading to proteinuria and albuminuria.<sup>[41]</sup> These disruptions compromise  $Mg^{2+}$  reabsorption and promote hypomagnesemia. A large cohort study of 5,126 chronic kidney disease (CKD) patients demonstrated a strong association between hypomagnesemia and high proteinuria, suggesting that protein loss drives  $Mg^{2+}$  wasting.<sup>[42]</sup> Similarly, low serum  $Mg^{2+}$  has been correlated with elevated microalbuminuria in T2DM.<sup>[43,44]</sup>

Magnesium handling is further impaired by increased renal excretion in T2DM. Studies consistently show higher  $Mg^{2+}$  urinary losses in diabetic versus healthy individuals.<sup>[45]</sup> Fractional excretion of magnesium (FE Mg) has even been proposed as a biomarker for diabetes detection.<sup>[46]</sup> Xu and Maalouf<sup>[47]</sup> found elevated FE Mg in hyperinsulinemic T2DM

patients compared with controls, suggesting that insulin resistance itself contributes to renal Mg<sup>2+</sup> wasting.

On the other hand, pharmacological interventions appear to improve Mg<sup>2+</sup> retention. Treatment with simvastatin has been shown to lower urinary Mg<sup>2+</sup> levels,<sup>[48]</sup> while metformin therapy reduces Mg<sup>2+</sup> excretion from the third month onward,<sup>[49]</sup> with even greater reductions when combined with sulfonylureas.<sup>[45]</sup> Interestingly, in patients undergoing bariatric surgery, serum Mg<sup>2+</sup> increased only in those who achieved T2DM remission, but not in those with persistent disease.<sup>[50]</sup>

### **Magnesium and Inflammation in T2DM**

Chronic inflammation is recognized as a key factor in the pathogenesis of type 2 diabetes mellitus (T2DM).<sup>[51,52]</sup> Multiple studies have shown strong associations between inflammatory markers and both the incidence and complications of T2DM. For instance, King *et al.*<sup>[53]</sup> observed that C-reactive protein (CRP) levels—a common marker of systemic inflammation—increased in parallel with HbA1c levels among T2DM patients. This supports the link between poor glycemic control and heightened inflammatory status.<sup>[53,54]</sup> Prospective studies further confirm that individuals with elevated CRP have a higher risk of developing T2DM.<sup>[55,56]</sup>

Because of this, alterations in magnesium homeostasis may indirectly contribute to insulin resistance by modulating inflammatory and oxidative stress pathways. Indeed, low dietary Mg<sup>2+</sup> intake has been repeatedly associated with higher CRP levels. Population studies show that individuals consuming less than the recommended Mg<sup>2+</sup> have significantly greater prevalence of high CRP compared with those meeting dietary recommendations,<sup>[57]</sup> a pattern also seen in healthy children.<sup>[58]</sup>

An inverse association between Mg<sup>2+</sup> intake and CRP levels has been documented across diverse populations, including adults, women adjusted for age and BMI,<sup>[59,60]</sup> and groups further adjusted for lifestyle factors such as physical activity, alcohol, and tobacco use.<sup>[61]</sup>

Cross-sectional studies have also reported negative correlations between serum or plasma Mg<sup>2+</sup> and CRP in both adults<sup>[62]</sup> and children.<sup>[63]</sup> A 20-year prospective cohort study strengthened this evidence by showing long-term inverse associations between Mg<sup>2+</sup> intake, serum Mg<sup>2+</sup>, and CRP levels.<sup>[64]</sup> Notably, King *et al.*<sup>[57]</sup> reported that Mg<sup>2+</sup> supplementation

reduced the prevalence of elevated CRP, even among individuals consuming less than 50% of the recommended intake.

Taken together, these findings suggest that adequate magnesium intake—through diet or supplementation—may help lower systemic inflammation, as reflected by reduced CRP, and could play a protective role in mitigating inflammation-driven insulin resistance in T2DM.

### Signs of Magnesium Deficiency<sup>[67,69]</sup>

- Muscle cramps and spasms
- Fatigue and weakness
- Irregular heartbeat
- Numbness or tingling
- Mood changes (anxiety, depression)

### Who Might Need Supplements

- Individuals with digestive disorders (e.g., Crohn's disease, celiac disease)
- People with type 2 diabetes
- Those with chronic alcohol use
- Older adults
- Patients on certain medications (e.g., diuretics, proton pump inhibitors)

### Precautions

- High supplemental doses (>350 mg/day) may cause diarrhea
- Magnesium can interact with medications such as antibiotics and antihypertensives

### Magnesium rich foods and Dietary supplements<sup>[65,66,68]</sup>

Magnesium Rich Foods	Magnesium Dietary Supplements
Green leafy vegetables like [spinach] Nuts and Seeds like[pumpkin seeds, almonds, cashews etc] Fishes like [mackerel and salmon] Fruits like [bananas and avocados] Dark chocolates Plain yogurt	Magnesium Citrate Magnesium Glycinate Magnesium Oxide Magnesium Chloride Magnesium Malate Magnesium L- Threonate

### Duration of Magnesium Supplementation and Its Influence on Health Outcomes

Clinical trials assessing magnesium ( $Mg^{2+}$ ) in type 2 diabetes management generally supplement participants for three to six months. Within this period, improvements in insulin

sensitivity and glycemic control have been reported. Longer supplementation may yield more pronounced benefits, with favorable effects on metabolic markers that support better long-term diabetes management and potentially reduce complications.

## **Study Overview: Serum Magnesium, Dietary Intake, and Metabolic Control in T2DM Objective**

To examine the relationship between serum magnesium levels, dietary magnesium intake, and metabolic control parameters in patients with type 2 diabetes mellitus (T2DM).<sup>[70]</sup>

## **METHODS**

- Participants: 119 T2DM patients (26 men, 93 women; mean age  $54.7 \pm 8.4$  years)<sup>[70]</sup>
- Measurements:
  - Serum magnesium measured via spectrophotometry
  - Dietary magnesium assessed using a food frequency questionnaire
  - Anthropometric parameters recorded
- Analysis: General Linear Model (GLM) applied to evaluate associations between serum magnesium and metabolic variables.<sup>[70]</sup>

## **RESULTS**

- **Prevalence**
  - 23.5% had inadequate dietary magnesium intake (<67% RDA)
  - 18.5% had hypomagnesemia (<0.75 mmol/L)
- **Metabolic outcomes**
  - Patients with hypomagnesemia had higher fasting plasma glucose (FPG), postprandial glucose (PPG), and HbA1c levels compared to normomagnesemic patients.<sup>[70]</sup>
  - FPG was significantly higher in hypomagnesemic patients in Model 1 ( $179.0 \pm 64.9$  vs.  $148.7 \pm 52.0$  mg/dL,  $p = 0.009$ ), though significance disappeared in adjusted models.
  - PPG remained significantly higher across all models (e.g., Model 1:  $287.9 \pm 108.4$  vs.  $226.8 \pm 89.4$  mg/dL,  $p = 0.006$ ).
  - HbA1c levels were consistently elevated in hypomagnesemia across all models ( $8.0 \pm 1.9\%$  vs.  $6.5 \pm 1.2\%$ ,  $p = 0.000$ ).<sup>[70]</sup>

- **Anthropometrics**
  - Body fat mass was significantly higher in hypomagnesemic patients in Model 3 ( $35.4 \pm 9.4$  vs.  $34.6 \pm 10.2$  kg;  $p = 0.034$ ).
- Dietary magnesium intake: No significant association with metabolic or anthropometric parameters.<sup>[70]</sup>

### Interpretation

These findings suggest that serum magnesium status, rather than dietary intake alone, is closely linked to glycemic control in T2DM. Hypomagnesemia was consistently associated with higher postprandial glucose, HbA1c, and greater body fat mass, indicating its potential role in poor metabolic outcomes.

### CONCLUSION

Hypomagnesemia in patients with type 2 diabetes mellitus (T2DM) is strongly associated with poor metabolic control, reflected in higher glucose levels, elevated HbA1c, and increased body fat mass. These findings emphasize the importance of monitoring magnesium status as part of routine clinical assessment in T2DM.

Magnesium plays a central role in insulin sensitivity and glucose metabolism. Consistently, low magnesium intake has been linked to a higher risk of developing T2DM, while adequate magnesium levels are associated with improved metabolic outcomes. Clinical and experimental studies suggest that magnesium supplementation can enhance insulin action, reduce blood sugar levels, and potentially lower the risk of diabetes-related complications.

Taken together, ensuring sufficient magnesium intake—through diet or supplementation—may represent a valuable and underutilized strategy for both the prevention and management of T2DM.

### REFERENCES

1. Saris NE, Mervaala E, Karppanen H, Khawaja JA, Lewenstam A. Magnesium: an update on physiological, clinical and analytical aspects. *Clin Chim Acta.*, 2000; 294(1-2): 1-26.
2. Delva P, Pastori C, Degan M, Montana M, Brazzarola P, Lechi A. Glucose-induced alterations of intracellular ionized magnesium in human lymphocytes. *Life Sci.*, 2002; 71(15): 1725-36.

3. Paolisso G, Barbagallo M. Hypertension, diabetes mellitus, and insulin resistance: the role of intracellular magnesium. *Am J Hypertens*, 1997; 10(3): 346-55.
4. Rodríguez-Morán M, Guerrero-Romero F. Low serum magnesium levels and foot ulcers in subjects with type 2 diabetes. *Arch Med Res.*, 2001; 32(4): 300-3.
5. Humphries S, Kushner H, Falkner B. Low dietary magnesium is associated with insulin resistance in a sample of young, nondiabetic black Americans. *Am J Hypertens.*, 1999; 12(8 Pt 1): 747-56.
6. Meyer KA, Kushi LH, Jacobs DR Jr, Slavin J, Sellers TA, Folsom AR. Carbohydrates, dietary fiber, and incident type 2 diabetes in older women. *Am J Clin Nutr.*, 2000; 71(4): 921-30.
7. McKeown NM, Meigs JB, Liu S, Wilson PW, Jacques PF. Whole-grain intake is favorably associated with metabolic risk factors for type 2 diabetes and cardiovascular disease in the Framingham Offspring Study. *Am J Clin Nutr.*, 2002; 76(2): 390-8.
8. Ma J, Folsom AR, Melnick SL, Eckfeldt JH, Sharrett AR, Nabulsi AA, et al. Associations of serum and dietary magnesium with cardiovascular disease, hypertension, diabetes, insulin, and carotid arterial wall thickness: the ARIC study. *J Clin Epidemiol.*, 1995; 48(7): 927-40.
9. Khan LA, Alam DS, Karim ZS, Bakir SA, Khan AK. Serum and urinary magnesium in young diabetic subjects in Bangladesh. *Am J Clin Nutr.*, 1999; 69(1): 70-4.
10. Takaya J, Higashino H, Kobayashi Y. Intracellular magnesium of platelets in children with diabetes and obesity. *Metabolism.*, 2003; 52(4): 468-71.
11. Ismail A, Ismail N. Magnesium: a mineral essential for health yet generally underestimated or even ignored. *J Nutr Food Sci.*, 2016; 6(2). doi:10.4172/2155-9600.1000523.
12. Glasdam SM, Glasdam S, Peters GH. The importance of magnesium in the human body: a systematic literature review. *Adv Clin Chem.*, 2016; 73: 169-93. doi:10.1016/bs.acc.2015.10.002.
13. de Baaij JHF, Hoenderop JGJ, Bindels RJM. Magnesium in man: implications for health and disease. *Physiol Rev.*, 2015; 95(1): 1-46. doi:10.1152/physrev.00012.2014.
14. DeFronzo RA, Ferrannini E, Groop L, Henry RR, Herman WH, Holst JJ, et al. Type 2 diabetes mellitus. *Nat Rev Dis Primers.*, 2015; 1: 15019. doi:10.1038/nrdp.2015.19.
15. Marshall SM. The pancreas in health and in diabetes. *Diabetologia.*, 2020; 63(10): 1962-1965. doi:10.1007/s00125-020-05235-z.

16. Kharade SV, Nichols C, Denton JS. The shifting landscape of KATP channelopathies and the need for ‘sharper’ therapeutics. *Future Med Chem.*, 2016; 8(7): 789-802. doi:10.4155/fmc-2016-0005.
17. Nichols CG. KATP channels as molecular sensors of cellular metabolism. *Nature*, 2006; 440(7083): 470-476. doi:10.1038/nature04711.
18. Gow IF, O'Donnell M, Flint L, Flapan AD. Infusion of Mg in humans acutely reduces serum insulin levels: a pilot study. *Magnes Res.*, 2011; 24(4): 189-195. doi:10.1684/mrh.2011.0295.
19. Murakami M, Ishizuka J, Sumi S, Nickols GA, Cooper CW, Townsend CM Jr, et al. Role of extracellular magnesium in insulin secretion from rat insulinoma cells. *Proc Soc Exp Biol Med.*, 1992; 200(4): 490-494. doi:10.3181/00379727-200-43459.
20. Gommers LMM, Hill TG, Ashcroft FM, de Baaij JHF. Low extracellular magnesium does not impair glucose-stimulated insulin secretion. *PLoS One.*, 2019; 14(6): e0217925. doi:10.1371/journal.pone.0217925.
21. Etwebi Z. Magnesium Regulation of Glucose and Fatty Acid Metabolism in HEPG2 Cells [Thesis]. Cleveland: Case Western Reserve University; 2011.
22. Molnes J, Teigen K, Aukrust I, Bjørkhaug L, Søvik O, Flatmark T, et al. Binding of ATP at the active site of human pancreatic glucokinase - nucleotide-induced conformational changes with possible implications for its kinetic cooperativity. *FEBS J.*, 2011; 278(13): 2372-2386. doi:10.1111/j.1742-4658.2011.08160.x.
23. Schmeltz L, Metzger B. Diabetes/syndrome X. In: Taylor JB, Triggle DJ, editors. *Comprehensive Medicinal Chemistry II*. Elsevier Science, 2006; 417-458.
24. Spellman CW. Pathophysiology of type 2 diabetes: targeting islet cell dysfunction. *J Am Osteopath Assoc.*, 2010; 110(3 Suppl 2): S2-S7.
25. Sperling MA, Tamborlane WV, Battelino T, Weinzimer SA, Phillip M. Diabetes Mellitus. In: Sperling MA, editor. *Pediatric Endocrinology*. 4th ed. W.B. Saunders, 2014; 846-900.
26. Guo S. Insulin signaling, resistance, and the metabolic syndrome: insights from mouse models into disease mechanisms. *J Endocrinol.*, 2014; 220(2): T1-T23. doi:10.1530/JOE-13-0327.
27. Kamran M, Kharazmi F, Malekzadeh K, Talebi A, Khosravi F, Soltani N. Effect of long-term administration of oral magnesium sulfate and insulin to reduce streptozotocin-induced hyperglycemia in rats: the role of Akt2 and IRS1 gene expressions. *Biol Trace Elem Res.*, 2019; 190(2): 396-404. doi:10.1007/s12011-018-1555-z.

28. Liu H, Li N, Jin M, Miao X, Zhang X, Zhong W. Magnesium supplementation enhances insulin sensitivity and decreases insulin resistance in diabetic rats. *Iran J Basic Med Sci.*, 2020; 23(8): 990-998. doi:10.22038/ijbms.2020.40859.9650.

29. Sales CH, dos Santos AR, Cintra DEC, Colli C. Magnesium-deficient high-fat diet: effects on adiposity, lipid profile and insulin sensitivity in growing rats. *Clin Nutr.*, 2014; 33(5): 879-888. doi:10.1016/j.clnu.2013.10.002.

30. Barooti A, Kamran M, Kharazmi F, Eftakhar E, Malekzadeh K, Talebi A, et al. Effect of oral magnesium sulfate administration on blood glucose hemostasis via inhibition of gluconeogenesis and FOXO1 gene expression in liver and muscle in diabetic rats. *Biomed Pharmacother*, 2019; 109: 1819-1825. doi:10.1016/j.biopha.2018.10.164.

31. Sohrabipour S, Sharifi MR, Sharifi M, Talebi A, Soltani N. Effect of magnesium sulfate administration to improve insulin resistance in type 2 diabetes animal model: using the hyperinsulinemic-euglycemic clamp technique. *Fundam Clin Pharmacol.*, 2018; 32(6): 603-616. doi:10.1111/fcp.12387.

32. Takaya J, Iharada A, Okihana H, Kaneko K. Down-regulation of hepatic phosphoenolpyruvate carboxykinase expression in magnesium-deficient rats. *Magnes Res.*, 2012; 25(3): 131-139. doi:10.1684/mrh.2012.0321.

33. Fapohunda O, Balogun O. Oral magnesium supplementation modulates hepatic and intestinal expression of some carbohydrate metabolizing genes in type 2 diabetic rats. *Int J Mol Biol.*, 2019; 4(5): 189-194. doi:10.15406/ijmboa.2019.04.00119.

34. Solaimani H, Soltani N, MaleKzadeh K, Sohrabipour S, Zhang N, Nasri S, et al. Modulation of GLUT4 expression by oral administration of Mg<sup>2+</sup> to control sugar levels in STZ-induced diabetic rats. *Can J Physiol Pharmacol.*, 2014; 92(6): 438-444. doi:10.1139/cjpp-2013-0403.

35. Morakinyo AO, Samuel TA, Adekunbi DA. Magnesium upregulates insulin receptor and glucose transporter-4 in streptozotocin-nicotinamide-induced type-2 diabetic rats. *Endocr Regul.*, 2018; 52(1): 6-16. doi:10.2478/enr-2018-0002.

36. Khosravi F, Kharazmi F, Kamran M, Malekzadeh K, Talebi A, Soltani N. The role of PPAR- $\gamma$  and NFKB genes expression in muscle to improve hyperglycemia in STZ-induced diabetic rat following magnesium sulfate administration. [Preprint/Article]. 2020.

37. Al Alawi AM, Majoni SW, Falhammar H. Magnesium and human health: perspectives and research directions. *Int J Endocrinol.*, 2018; 2018: 9041694. doi:10.1155/2018/9041694.

38. Curry JN, Yu ASL. Magnesium handling in the kidney. *Adv Chronic Kidney Dis.*, 2018; 25(3): 236-243. doi:10.1053/j.ackd.2018.01.003.

39. Bouras H, Roig SR, Kurstjens S, Tack CJJ, Kebieche M, de Baaij JHF, et al. Metformin regulates TRPM6, a potential explanation for magnesium imbalance in type 2 diabetes patients. *Can J Physiol Pharmacol.*, 2020; 98(6): 400-411. doi:10.1139/cjpp-2019-0570.

40. Lee CT, Lien YHH, Lai LW, Chen JB, Lin CR, Chen HC. Increased renal calcium and magnesium transporter abundance in streptozotocin-induced diabetes mellitus. *Kidney Int.*, 2006; 69(10): 1786-1791. doi:10.1038/sj.ki.5000344.

41. Singh A, Satchell SC. Microalbuminuria: causes and implications. *Pediatr Nephrol.*, 2011; 26(11): 1957-1965. doi:10.1007/s00467-011-1777-1.

42. Oka T, Hamano T, Sakaguchi Y, Yamaguchi S, Kubota K, Senda M, et al. Proteinuria-associated renal magnesium wasting leads to hypomagnesemia: a common electrolyte abnormality in chronic kidney disease. *Nephrol Dial Transplant.*, 2019; 34(7): 1154-1162. doi:10.1093/ndt/gfy119.

43. Corica F, Corsonello A, Ientile R, Cucinotta D, Di Benedetto A, Perticone F, et al. Serum ionized magnesium levels in relation to metabolic syndrome in type 2 diabetic patients. *J Am Coll Nutr.*, 2006; 25(3):210-215. doi:10.1080/07315724.2006.10719534.

44. Lu J, Gu Y, Guo M, Chen P, Wang H, Yu X. Serum magnesium concentration is inversely associated with albuminuria and retinopathy among patients with diabetes. *J Diabetes Res.*, 2016; 2016: 1260141. doi:10.1155/2016/1260141.

45. Peters KE, Chubb SAP, Davis WA, Davis TME. The relationship between hypomagnesemia, metformin therapy and cardiovascular disease complicating type 2 diabetes: the Fremantle diabetes study. *PLoS One.*, 2013; 8(9): e74355. doi:10.1371/journal.pone.0074355.

46. Futrakul N, Futrakul P. Biomarker for early renal microvascular and diabetic kidney diseases. *Ren Fail*, 2017; 39(1): 505-511. doi:10.1080/0886022X.2017.1323647.

47. Xu LHR, Maalouf NM. Effect of acute hyperinsulinemia on magnesium homeostasis in humans: magnesium metabolism in hyperinsulinemia. *Diabetes Metab Res Rev.*, 2017; 33(2): e2844. doi:10.1002/dmrr.2844.

48. Xu J, Xu W, Yao H, Sun W, Zhou Q, Cai L. Associations of serum and urinary magnesium with the pre-diabetes, diabetes and diabetic complications in the chinese northeast population. *PLoS One.* 2013; 8(2): e56750. doi:10.1371/journal.pone.0056750.

49. Doşa MD, Hangan LT, Crauciuc E, Galeş C, Nechifor M. Influence of therapy with metformin on the concentration of certain divalent cations in patients with non-insulin-

dependent diabetes mellitus. *Biol Trace Elem Res.*, 2011; 142(1): 36-46. doi:10.1007/s12011-010-8751-9.

50. Lecube A, Baena-Fustegueras JA, Fort JM, Pelegrí D, Hernández C, Simó R. Diabetes is the main factor accounting for hypomagnesemia in obese subjects. *PLoS One.*, 2012; 7(1): e30599. doi:10.1371/journal.pone.0030599.

51. Bloch-Damti A, Bashan N. Proposed mechanisms for the induction of insulin resistance by oxidative stress. *Antioxid Redox Signal.*, 2005; 7(11-12): 1553-1567. doi:10.1089/ars.2005.7.1553.

52. Rains JL, Jain SK. Oxidative stress, insulin signaling, and diabetes. *Free Radic Biol Med.*, 2011; 50(5): 567-575. doi:10.1016/j.freeradbiomed.2010.12.006.

53. King DE, Mainous AG, Buchanan TA, Pearson WS. C-reactive protein and glycemic control in adults with diabetes. *Diabetes Care*, 2003; 26(5): 1535-1539. doi:10.2337/diacare.26.5.1535.

54. Black S, Kushner I, Samols D. C-reactive protein. *J Biol Chem.*, 2004; 279(47): 48487-48490. doi:10.1074/jbc.R400025200.

55. Han TS, Sattar N, Williams K, Gonzalez-Villalpando C, Lean MEJ, Haffner SM. Prospective study of C-reactive protein in relation to the development of diabetes and metabolic syndrome in the Mexico city diabetes study. *Diabetes Care*, 2002; 25(11): 2016-2021. doi:10.2337/diacare.25.11.2016.

56. Laaksonen DE, Niskanen L, Nyyssönen K, Punnonen K, Tuomainen TP, Valkonen VP, et al. C-reactive protein and the development of the metabolic syndrome and diabetes in middle-aged men. *Diabetologia*, 2004; 47(8): 1403-1410. doi:10.1007/s00125-004-1472-x.

57. King DE, Mainous AG III, Geesey ME, Egan BM, Rehman S. Magnesium supplement intake and C-reactive protein levels in adults. *Nutr Res.*, 2006; 26(5): 193-196. doi:10.1016/j.nutres.2006.05.001.

58. King DE, Mainous AG, Geesey ME, Ellis T. Magnesium intake and serum C-reactive protein levels in children. *Magnes Res.*, 2007; 20(1): 32-36.

59. Bo S, Durazzo M, Guidi S, Carello M, Sacerdote C, Silli B, et al. Dietary magnesium and fiber intakes and inflammatory and metabolic indicators in middle-aged subjects from a population-based cohort. *Am J Clin Nutr.*, 2006; 84(5): 1062-1069. doi:10.1093/ajcn/84.5.1062.

60. Song Y, Ridker PM, Manson JE, Cook NR, Buring JE, Liu S. Magnesium intake, C-reactive protein, and the prevalence of metabolic syndrome in middle-aged and older U.S. women. *Diabetes Care.*, 2005; 28(6): 1438-1444. doi:10.2337/diacare.28.6.1438.
61. Song Y, Li TY, van Dam RM, Manson JE, Hu FB. Magnesium intake and plasma concentrations of markers of systemic inflammation and endothelial dysfunction in women. *Am J Clin Nutr.*, 2007; 85(4): 1068-1074. doi:10.1093/ajcn/85.4.1068.
62. Chen S, Jin X, Liu J, Sun T, Xie M, Bao W, et al. Association of plasma magnesium with prediabetes and type 2 diabetes mellitus in adults. *Sci Rep.*, 2017; 7(1): 12763. doi:10.1038/s41598-017-13050-7.
63. Rodríguez-Morán M, Guerrero-Romero F. Serum magnesium and C-reactive protein levels. *Arch Dis Child.* 2008; 93(8): 676-680. doi:10.1136/adc.2006.109371.
64. Kim DJ, Xun P, Liu K, Loria C, Yokota K, Jacobs DR Jr, et al. Magnesium intake in relation to systemic inflammation, insulin resistance, and the incidence of diabetes. *Diabetes Care.*, 2010; 33(12): 2604-2610. doi:10.2337/dc10-0994.
65. National Institutes of Health. Magnesium: Fact Sheet for Health Professionals. NIH Office of Dietary Supplements; 2022.
66. Gröber U, Schmidt J, Kisters K. Magnesium in prevention and therapy. *Nutrients.* 2015; 7(9): 8199-8226. doi:10.3390/nu7095388.
67. Kirkland AE, Sarlo GL, Holton KF. The role of magnesium in neurological disorders. *Nutrients.*, 2018; 10(6): 730. doi:10.3390/nu10060730.
68. Hruby A, McKeown NM, Song Y, Djoussé L, Chu AY, Hanson RL, et al. Dietary magnesium intake and risk of metabolic syndrome: a meta-analysis. *Eur J Clin Nutr.*, 2014; 68(11): 1269-1278. doi:10.1038/ejcn.2014.155.
69. DiNicolantonio JJ, O'Keefe JH, Wilson W. Subclinical magnesium deficiency: a principal driver of chronic disease? *Open Heart*, 2018; 5(1): e000668. doi:10.1136/openhrt-2017-000668.
70. Veronese N, Watutantrige-Fernando S, Luchini C, Solmi M, Sartori L, Musacchio E, et al. Effect of magnesium supplementation on glucose metabolism in people with or at risk of diabetes: a systematic review and meta-analysis of double-blind randomized controlled trials. *Eur J Clin Nutr.*, 2016; 70(12): 1354-1359. doi:10.1038/ejcn.2016.154.