

# WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 8.084

Volume 11, Issue 2, 2368-2377.

Research Article

ISSN 2277-7105

## VENATION PATTERN IN EIGHT SPECIES OF LINDERA OF FAMILY LAURACEAE

#### Ulka Chodankar\*

S.V. K. M's Mithibai College of Arts, Chauhan Institute of Science and Amrutben Jivanlal College of Commerce and Economics (Autonomous), Affiliated to University of Mumbai, Vile Parle - West Mumbai 400056, India.

Article Received on 21 Dec. 2021.

Revised on 11 Jan. 2022. Accepted on 01 Feb. 2022 DOI: 10.20959/wjpr20222-23125

### \*Corresponding Author Ulka Chodankar

S.V. K. M's Mithibai College of Arts, Chauhan Institute of Science and Amrutben Jivanlal College of Commerce and Economics (Autonomous), Affiliated to University of Mumbai, Vile Parle - West Mumbai 400056, India.

#### **ABSTRACT**

Lindera belongs to family Lauraceae. The family is economically important. The existing classifications of Lauraceae are based on floral androecial characters. The separation of species of *Lindera* is quite difficult based on morphological characters in the family and so the leaf venation of eight species of Lindera is studied and a key to venation is made so that the species can be separated based on venation pattern. Anatomy is a very important tool which can be used to other identify the species along with the morphological characters. The species studied are Lindera assamica, L. bifaria, L. caudata, L. griffithi, L. latifolia, L. melastomacea, L. nessiana and L. pulcherrima. The types of venation found is pinnate camptodromous with festooned brochidodromous secondaries, acrodromous with perfect basal secondaries and is acrodromous with perfect supra-basal secondaries. There is a variation in angle of divergence of secondary veins and pattern of tertiary veins. The highest vein order also ranges from 4<sup>0</sup> to

6<sup>0</sup>. Presence or absence of tracheoids has also been studied along with the angle of origin of tertiary veins from exmedial to admedial sides is also showing a range which can be used to separate the species.

**KEYWORDS:** Pinnate camptodromous, acrodromous, festooned brochidodromous, basal and suprabasal.

#### INTRODUCTION

Lindera has about 80 species in the world. Fifteen species are found in India. The genus Lindera belongs to tribe II Litsaeaceae which is divided into 4 sections: Section I: Aperula: to which belong L. assamica and L. latifolia which have persistent leaves, umbels, long pedicelled with 4 bracts, involucriform, 6-12 flowers and 9 stamens. Section II: Polydenia: in which leaves are persistent, with the exception of L. venosa, umbels which are sessile or siubsessile with 4-8 bracts and 9-12 stamens and L. bifaria belongs to this section. Section III: Daphnidium: to which belong L. caudata, L. pulcherrima and L. melastomacea which have persistent leaves, triple nerved. Section IV: Sassafrimorpha to which L. nessiana and L. griffithi belong. They have deciduous leaves. L. nessiana is triple or quintriple nerved and L. griffithi is undetermined section species. The separation of species is quite difficult and so the leaf venation is studied and a key to venation is made so that the species can be separated based on venation pattern (Hooker, 1883; Kanjilal & Das 1939). So the study of venation will be helpful as an aid to identify the species.

#### MATERIAL AND METHODS

The plant material for the present work was personally collected from Shillong- Meghalaya; Kodaikanal, Kolli Hills-Tamilnadu. The duplicates of herbarium were collected from the herbarium section of B.S.I. Eastern Circle and A.R.I., Pune. The identification of fresh material was checked with the help of Standard Herbaria from B.S.I. Shillong and B.S.I. Yercaud and A.R.I. Herbarium, Pune.

1. For the study of leaf architecture, the mature leaves either fresh, dried or preserved were first cleared by keeping them in 5% sodium hydroxide solution at room temperature for 1-2 days. The decoloured leaves were washed and transferred to 5% sodium hypochlorite till they were transparent. For more clarity the leaves were washed and put into the solution of trichloroacetic acid and phenol (2:1 by weight) for a few minutes at room temperature. They were then thoroughly washed to remove acid traces and were stained with aqueous Saffranine by keeping them in it for 10-15 minutes. The leaves were then transferred to 50% Glycerine and mounted in Glycerine jelly. (Payne, 1969; Mohan Ram and Nayyar, 1968) and the standard terminology is used in anatomical studies as given by Hickey and Wolfe. 1975; Melville, 1976; Hickey, 1973, 1979; Dilcher, 1974.

The microphotographs showing different anatomical features were removed by using Nikon Camera at various magnifications as mentioned in the plates.

#### **OBSERVATIONS**

Eight species of *Lindera* are studied namely, *L. assamica*, *L. bifaria*, *L. caudata*, *L. griffithi*, L. latifolia, L. melastomacea, L. nessiana and L. pulcherrima (Photoplates I-III)

Pinnate camptodromous with festooned brochidodromous type of venation is observed in six species namely: L. assamica, L. bifaria, L. griffithi, L. latifolia, L. nessiana and L. pulcherrima. Highest vein order of the leaf is 4°, the angle of divergence of secondary veins is acute moderate, pattern of tertiary veins is percurrent, angle of origin of tertiary veins OO/AO/OA/RR/OR/RO and tracheoids present.

Highest vein order of the leaf is 5°, the angle of divergence of secondary veins is acute narrow, pattern of tertiary veins is percurrent, angle of origin of tertiary veins OO/OA/OR/AR/RO/RA in L. bifaria and angle of origin of tertiary veins OA/OR/RR/RO/AA/OO in L. pulcherrima tracheoids present.

Highest vein order of the leaf is 6°, the angle of divergence of secondary veins is acute narrow, pattern of tertiary veins is random reticulate in three species L. assamica, L. latifolia and L. nessiana. The angle of origin of tertiary veins RA/RR/OA/RO/OR in L. latifolia, OR/OO/RR/RA/AR in L. nessiana and RR/AR/OA/OR/AO in L. assamica and tracheoids absent.

Two species of Lindera namely L. caudata and L. melastomacea show acrodromous type of venation with perfect suprabasal position, pattern of tertiary veins is random reticulate and angle of origin of tertiary veins is OO/OR/OA/RR/AR and tracheoids absent in L. caudata. L. melastomacea shows acrodromous type of venation with perfect basal position, pattern of tertiary veins is random reticulate and angle of origin of tertiary veins is OO/OR/OA/RR/RA and tracheoids present.

**SUMMARY**: Based on the pattern of venation a key to the species could be made.

#### **KEY TO VENATION**

Venation is pinnate camptodromous with festooned brochidodromous secondaries.

Angle of divergence of secondary veins is acute narrow

Pattern is of tertiary veins is random reticulate

Highest vein order 5<sup>0</sup>

Tracheoids absent

Angle or origin exmedial to

admedial side is RR/AR/OA/OR/AO -----Lindera assamica

Highest vein order 6<sup>0</sup>

Tracheoids present

Angle or origin exmedial to

admedial side is RR/AR/OA/OR/AO --- Lindera nessiana

Venation is pinnate camptodromous with festooned brochidodromous secondaries.

Angle of divergence of secondary veins is acute narrow

Pattern is of tertiary veins is percurrent

Highest vein order 5<sup>0</sup>

Tracheoids absent

Angle or origin exmedial to

admedial side is OO/OA/OR/AR/RR/RO/RA ----Lindera bifaria

Angle or origin exmedial to

admedial side is OA/OR/RR/RO/OO-----Lindera pulcherrima

Highest vein order 6<sup>0</sup>

Tracheoids present

Angle or origin exmedial to

admedial side is RA/RR/OA/RO/OR ------Lindera latifolia

Angle of divergence is acute moderate

Highest vein order 4<sup>0</sup>

Tracheoids present

Angle or origin exmedial to

admedial side is OO/AO/OA/RR/OR/RO -----Lindera griffithii

Venation is acrodromous with perfect basal secondaries.

Angle of divergence of secondary veins is acute narrow

Pattern is of tertiary veins is random reticulate

Highest vein order 4<sup>0</sup>

Tracheoids present

Chodankar.

Angle or origin exmedial to admedial side is OR/OA/RR/OO/RA -----Lindera melastomacea

Venation is acrodromous with perfect supra-basal secondaries.

Angle of divergence of secondary veins is acute moderate

Pattern is of tertiary veins is random reticulate

Highest vein order 5<sup>0</sup>

Tracheoids absent

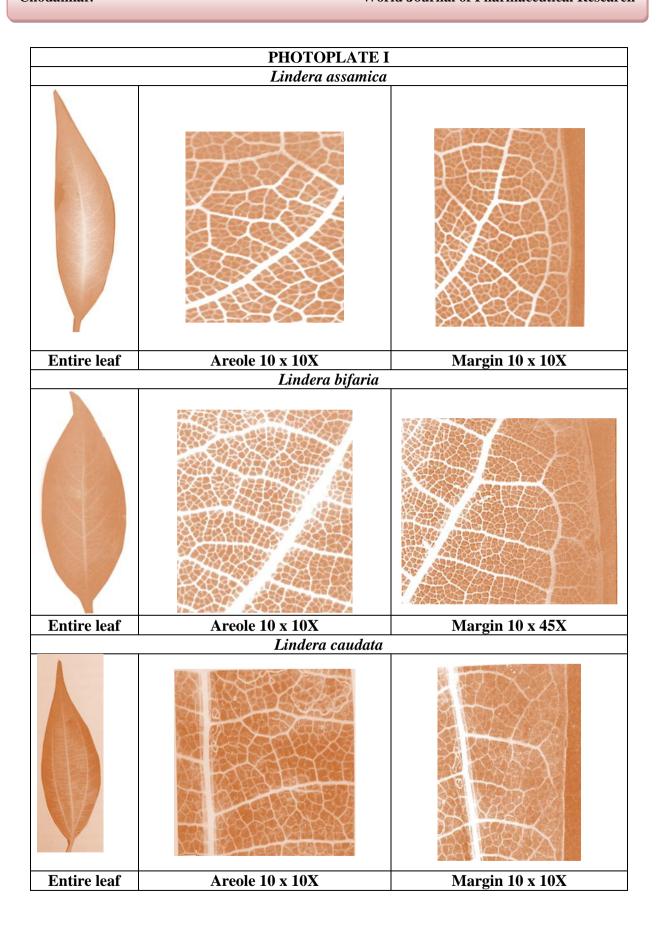
Angle or origin exmedial to

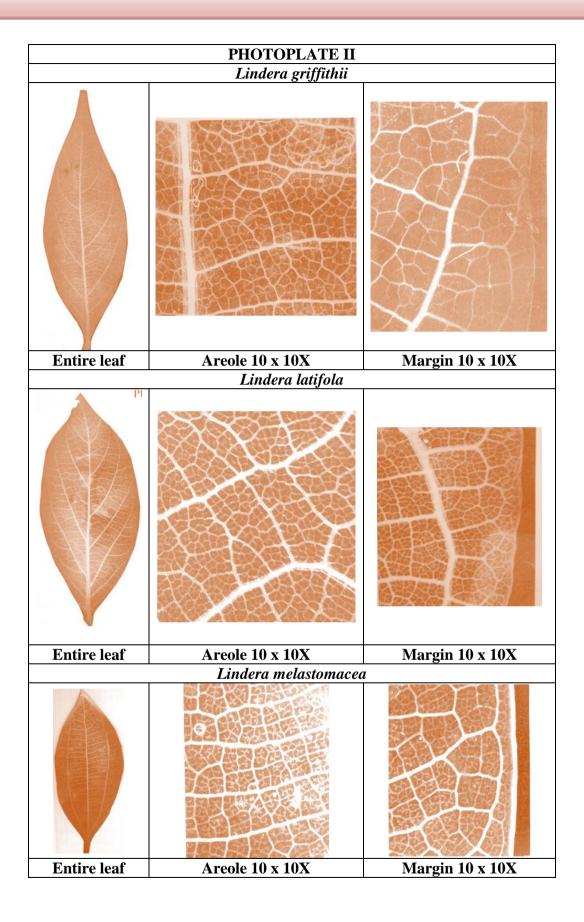
admedial side is OO/OR/OA/RR/AR -----Lindera caudata

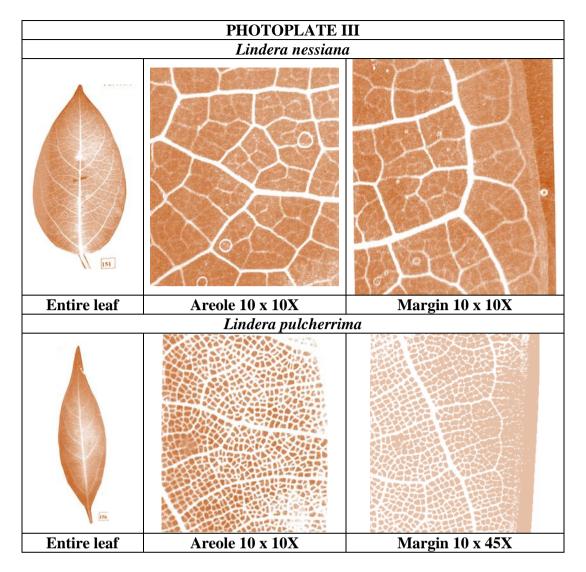
[R- Right angle; A- Acute angle; O- Obtuse angle]

#### DISCUSSION

Leaf architectural study is found to be useful for taxonomic purpose as suggested by Dilcher (1974) and Hall and Melville (1951). The veinlet termination number is used as a technique for testing the purity of fragments of a particular leaf type for pharmacognostical properties. Further contributions in use of absolute veinislet number were made by Hall and Melville (1954). Venation features of angiosperm leaves have also been added by Gupta (1961) and Meyerhoff (1952). Descriptive terminology and leaf architecture usage has been presented by Klucking (1962), Madler and Strauss (1971) and Ferguson, (1974). An outline of leaf architecture of dicot leaves is also presented earlier which has been revised and the architecture of dicotyledonous angiospermous leaves has been classified. Tracheoids can be used as a diagnostic feature in certain species of Lauraceae Mouten, (1967) & Kim and Moon Hong Kim (1984. Leaf venation patterns have been studied in *Litsaea* by Vaidya (2015) and anatomical studies in Schleichera oleosa too have been carried out (20) Vaidya and Guleria (2015). The venation patterns of six species of *Phoebe* and eleven species of *Cinnamomum* have been reported by Chodankar and Vaidya (2021a; 2021b).







#### **CONCLUSION**

The eight species of *Lindera* are studied namely, *L. assamica*, *L. bifaria*, *L. caudata*, *L. griffithi*, *L. latifolia*, *L. melastomacea*, *L. nessiana* and *L. pulcherrima* could be separated based on venation pattern study and can be used as an aid to the identification of the eight species. Anatomy can be used to help in the correct identification of a species.

#### **REFERENCES**

- 1. Chodankar, U and M. Vaidya: Patterns of Leaf Architecture In Six Species Of *Phoebe* From Family Lauraceae In WJPR, 2021a; 10(3): 1772-1778.
- 2. Chodankar, U and M. Vaidya: Study Of Leaf Architecture In Eleven Species Of *Cinnamomum* Blume. Of Family Lauraceae In WJPR, 2021b; 10(2): 1190-1199.
- 3. Dilcher, D.L. The Study of Angiosperm Leaf remains. The Botanical Rev., 1974; 40: 1-157.

- 4. Ferguson, D.K. On the taxonomy of recent and fossil species of *Laurus* (Lauraceae). J. Link. Soc. Lond. Bot., 1974; 68: 51-72.
- 5. Gupta, R. Correlation of tissues in leaves. I. Absolute vein-islet numbers and absolute veinlet termination numbers, Ann Bot., 1961; 47: 684-698.
- 6. Hall, J. P. and Melville, C. Veinlet termination number a new character for the differentiation of leaves. J. Pharm. Pharmac., 1951; 3: 934-941.
- 7. Hall, J. P. and Melville, C. Veinlet termination numbers some further observation. J. Pharm. Pharmac., 1954; 6: 129-133.
- 8. Hickey L.J. A revised classification of the architecture of dicoty ledonous leaves. In: Metcalfe and Chalk. Anatomy of dicotyledons. Clarendon Press, Oxford, 1979.
- 9. Hickey, L.J. Classification of the architecture of dicotyledonous leaves. Amer. J. Bot., 1973; 60: 17-35.
- 10. Hickey, L.J. and J.A. Wolfe. The basis of angiosperm Phylogeny: venation. Ann. Mlssouri Bot. Gard., 1975; 62: 538-589.
- 11. Hooker, J.D. Flora of British India. Reeve and Co., London, 1883; 5: 116-189.
- 12. Kanjilal, U.N. and Das A.K. Flora of Assam, 1939; 4: 46-93.
- 13. Kim Kyung Sik and Moon Hong Kim. Venation Pattern of Lauraceae Korean plants. Korean J. Bot., 1984; 27(1): 15-24.
- 14. Klucking, E. P. An oligocene flora from the Western Cascades, with an analysis of leaf structure, Ph. D. Thesis, University of California, Berkeley, Dept. of Geology, 1962.
- 15. Madler, K. and Strauss, A. Em system der blattformen mit spezieller anwendung fur die bestimmung Noegener blattreste. Bot. J., 1971; 90: 562-574.
- 16. Melville, R. The terminology of leaf architecture of Apocynaceae. Taxon, 1976; 25: 549-561.
- 17. Meyerhoff, A.A. (1952). A study of leaf venation of Betulaceae with its applications to Paleobotany, Ph. D. Thesis, Stanford University.
- 18. Mohan Ram, H.Y. and Nayyar Vijaylaxmi. Leaf clearing techinque with a wide range of applications; Proc. Indian Acad. Sci. (Plant Sci.) B., 1978; 87: 125-127.
- 19. Mouten, J. A. Architecture de la nervation Foliare. 92. Congress national des societetes savantes. Strashourg et Columr, 1967; III: 165-176.
- 20. Payne, W.W. A quick method for clearing leaves. Ward's Bulletin, 1969; 8(61): 4,5.
- 21. Vaidya, M. Study of leaf architecture in some species of *Litsaea* Lamk. Of family Lauraceae in Journal of anatomy, 2015; 115: 182-184.

22. Vaidya, M and H. Guleria. Anatomical Studies of Schleichera oleosa (Lour.) Oken. in WJPR, 2015; 4(12): 1178-1188.