

EVALUATING THE CLINICAL EFFECTIVENESS AND SAFETY OF TULSI: A COMPREHENSIVE REVIEW

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ABSTRACT

Tulsi, or holy basil, is a venerated medicinal herb that has profound historical significance in Ayurvedic medicine. This study consolidates data from more than one hundred investigations on the pharmacological advantages of Tulsi, emphasizing its adaptogenic, anti-inflammatory, and immunomodulatory characteristics, among others. Despite considerable in vitro and animal studies indicating its therapeutic potential, there is a significant deficiency of complete human clinical trials assessing the safety and efficacy of Tulsi as a singular therapy. This paper seeks to critically evaluate these human clinical trials to offer an educated view of the therapeutic efficacy and safety of Tulsi.

KEYWORDS: Pharmacological, Immunomodulatory, Adaptogenic.

INTRODUCTION

Tulsi in Hindi or Tulasi in Sanskrit (holy basil in English) is a highly esteemed culinary and medicinal fragrant plant belonging to the Lamiaceae family. It is native to the Indian subcontinent and has been utilized in Ayurvedic treatment for over 3000 years. In the Ayurvedic system, Tulsi is frequently referred to as an "Elixir of Life" due to its therapeutic properties and its efficacy in addressing several prevalent health issues. Tulsi leaf extracts are documented in the Indian Materia Medica for treating bronchitis, rheumatism, and pyrexia.^[1] Additional documented therapeutic applications encompass the management of epilepsy, asthma or dyspnea, hiccups, cough, dermatological and hematological disorders, parasite infections, neuralgia, headaches, wounds, and inflammation^[2] and oral conditions.^[3] The juice extracted from the leaves has been utilized as a remedy for earache^[4], The tea infusion has been utilized for the treatment of stomach and hepatic problems.^[5] The roots and stems

were historically employed to remedy insect and snake bites, as well as malaria.^[5]

Three varieties of tulsi are frequently delineated. *Ocimum tenuiflorum* (or *Ocimum sanctum* L.) comprises two botanically and phytochemically different cultivars: Rama or Sri tulsi (green leaves) and Krishna or Shyama tulsi (purplish leaves)^[6,7], *Ocimum gratissimum* is a third kind of tulsi, referred to as Vana or wild/forest tulsi, characterized by its dark green leaves.^[8,9] The many forms of tulsi have significant variety in shape and phytochemical composition, including secondary metabolites; nonetheless, they may be differentiated from other *Ocimum* species by their yellow pollen and elevated eugenol content^[10], and smaller chromosome number.^[11] Despite being distinct species with *Ocimum tenuiflorum* having six times less DNA than *Ocimum gratissimum*^[11], they are traditionally employed identically to address comparable diseases.^[5] This review consistently used the term tulsi to denote both *Ocimum tenuiflorum* and *Ocimum gratissimum*.

Tulsi has been extensively studied, with over one hundred papers in the past decade addressing its pharmacological properties and many medicinal uses. Numerous in vitro and animal investigations demonstrate that tulsi leaf has significant pharmacological properties, including adaptogenic effects^[12,14], metabolic^[15,17], immunomodulatory^[18,20], anticancer^[21,23], anti-inflammatory^[24,25], antioxidant^[26,27], hepatoprotective^[28,29], radioprotective^[30,31], antimicrobial^[32,35], and antidiabetic effects^[36,38] that have been extensively reviewed previously.^[39,45]

Preclinical studies have demonstrated that tulsi increases swimming survival times in mice and prevents stress-induced ulcers in rats^[46] exhibiting antistress effects akin to those of antidepressant medications.^[47] Similarly, recent studies report leaf extracts from ethanolic and aqueous tulsi to protect rats from stress-induced cardiovascular changes.^[47,49] Research utilizing animal models has demonstrated that tulsi leaf extract has anticonvulsant and anxiolytic properties.^[50,51] Numerous animal studies published over the last fifty years indicate that the consumption of tulsi leaves improves glucose and lipid profiles in both normal and diabetes-induced animal models.^[36,38,52–58] The intramammary infusion of Tulsi aqueous leaf extract has demonstrated a potential impact in enhancing the immunological response in bovine animals.^[59]

Alongside the comprehensive literature detailing in vitro and animal experiments, investigations into the use of tulsi within a polyherbal formulation in people have been

extensively evaluated. Currently, there are no comprehensive evaluations assessing the therapeutic effectiveness and safety of tulsi as a standalone herbal intervention in humans. This study aimed to describe and critically evaluate human clinical studies of tulsi to assess its clinical effectiveness and safety based on existing data.^[60]

Approximately 160 species within the genus *Ocimum*, such as *Ocimum sanctum*, *Ocimum americanum*, *Ocimum basilicum* (Ban Tulsi), *Ocimum camphor*, *Ocimum canum* (Dulal Tulsi), *Ocimum gratissimum* (Ram Tulsi), *Ocimum kilimandscharicum*, *Ocimum micranthum*, and *Ocimum tenuiflorum* (Krishna Tulsi), are extensively distributed across the warm regions of the globe and possess recognized medicinal properties.^[62–65]

Sanctum is a fragrant herb and an invasive weed. It is an upright, extensively branched perennial or biennial subshrub, measuring 30–75 cm in height, characterized by hairy stems and simple, opposite, aromatic green leaves. The leaves are ovate, oblong, obtuse, or acute, measuring up to 5 cm in length, and are typically somewhat serrated. Flowers are little and purple-hued, arranged in elongated racemes with closely spaced whorls. The fruits are little, and the seeds have a reddish-yellow hue.^[66] The plant is cultivated across India, from the Himalayas (up to 1800 meters above sea level) to the Andaman and Nicobar Islands.^[67] It propagates by seeds and may be cultivated effortlessly across diverse soils and weather conditions.

Sanctum is renowned for its remarkable therapeutic properties in ancient folklore and several indigenous medical systems, including Ayurveda, Unani, Siddha, Greek, and Roman practices.^[64,69] Distinguished for its powerful fragrance and astringent flavor. In Ayurveda, *Sanctum* is referred to as 'the elixir of life' and is said to enhance longevity. *Sanctum* is conventionally used in several forms, including cold or hot infusions of dried leaves (herbal tea), dry powder, fresh leaves, alcoholic tinctures, and oil (ghee) preparations, as well as compositions including stem, root, and seeds, utilized both systemically and topically.^[70] As per the Indian Materia Medica, which encompasses the Ayurvedic and Unani medical traditions. *Sanctum* possesses numerous medicinal properties, including antipyretic, antiseptic, antiemetic, alexipharmic, carminative, diaphoretic, demulcent, expectorant, and stimulant effects. Consequently, it has been advocated for the treatment of various ailments, such as malaria fever, bronchitis, catarrh, gastric and genitourinary disorders, rheumatism, and dermatological conditions.^[71] The leaves of the *sanctum* are utilized for throat and chest disorders, as well as for alleviating colds, coughs, and indigestion. The juice extracted from

fresh leaves, blossoms, and thin roots serves as an antidote for snake bites and scorpion stings.^[72] The leaves are utilized as a seasoning in salads and various culinary applications. The essential oil derived from the leaves is utilized to relieve joint discomfort and possesses significant insecticidal and larvicidal properties. The seeds possess anticoagulant qualities and are beneficial for both constipation and diarrhea.^[73] Consequently, every component of the plant possesses medicinal properties and is utilized for the treatment of many ailments grounded in traditional knowledge.^[68]

Recent scientific investigations have demonstrated that various components of sanctum, including leaves, stem, root, flowers, and seed, exhibit a multitude of biological and pharmacological activities, such as antioxidant, anti-inflammatory, antiallergic, immunomodulatory, anticoagulant, antimicrobial, antistress, antiulcer, wound-healing, anticataract, analgesic, antipyretic, antihypertensive, antidiabetic, antifertility, central nervous system depressant, cardioprotective, gastroprotective, hepatoprotective, renoprotective, radioprotective, chemopreventive, and anticancer properties.^[61,66,68,70,73–76] Despite several recent studies providing a comprehensive summary of the pharmacological characteristics of sanctum, the application of this herb for chemoprevention or treatment of oncological disorders has not been thoroughly and critically examined in the literature. This essay seeks to deliver, for the first time, a thorough and critical examination of current anticancer and cancer prevention research on sanctum.

Ocimum sanctum and cancer

This section emphasizes research that illustrates the impact of sanctum extracts and phytoconstituents in preclinical in vitro and in vivo cancer models, as well as in clinical contexts.

In-vitro studies

The use of an ethanolic extract of sanctum leaves to HFS-1080 human fibrosarcoma cells demonstrated cytotoxicity at a dosage of 50 mg/ml. Mechanistic investigations have identified induced apoptosis ask, indicating reduction or downregulation; m, indicating rise or overexpression.

Reduced intracellular glutathione (GSH) and elevated amounts of lipid peroxidation products.^[77] 6-alkylamines are powerful mutagenic, carcinogenic, and cytotoxic molecular lesions caused by both exogenous and endogenous alkylating agents. Enhanced clearance of

these lesions by the augmentation of methylguanine–DNA methyltransferase (MGMT) activity appears to be an effective chemopreventive approach. Both ethanolic and aqueous extracts of sanctum leave dose-dependently increased amounts of MGMT mRNA and protein, as well as its demethylation activity in HT29 human colon cancer cells. The activity and protein expression of glutathione S-transferase-pi (GSTP1) were elevated after treatment with each extract.^[78] These findings highlight the potential of using sanctum to mitigate alkylation- induced carcinogenesis. Magesh et al.^[79] examined the anticancer efficacy of an ethanolic extract of sanctum leaves on A549 human non-small-cell lung carcinoma cells. The extract demonstrated cytotoxicity, elevated the sub-G1 population, and generated apoptotic bodies. Furthermore, the extract-cleaved poly (ADP-ribose) polymerase (PARP) liberated cytochrome c into the cytosol, activated caspase-9 and caspase-3 proteins, elevated the ratio of pro- apoptotic protein Bax to anti-apoptotic protein Bcl-2 and concurrently inhibited the phosphorylation of AKT and extracellular signal-regulated kinase (ERK). A subsequent study by the same researchers utilizing mouse Lewis lung carcinoma cells demonstrated that the extract markedly inhibited tumor cell adhesion and invasion, along with the activity of matrix metalloproteinase-9 (MMP-9), but not MMP-2, underscoring the critical role of MMP-9 in the antimetastatic efficacy of sanctum. The anticancer activity of vicenin-2, an active component of the sanctum, has been evaluated against several human prostate cancer cell lines, including LNCaP (androgen-dependent), PC-3, and DU-145 (androgen-independent).^[80]

Morphology

Distribution	Grow up to 30 - 60 cm in height.
Territory	Found throughout the Indis.
Habit	Annual herb.
Mool	Thin, wired, branches, hairs, soft, color black to brown from external or dull and violet internal.
Tana	Stiff, greenly, wooded, branches are hairy, external color pinkish-brown to black, internal color dull yellowish, fracture: stringy and slightly aromatic odor.
Patra	Leaves are 2.5 to 5cm long 1.6 to 3.2 cm wide, elliptically egg-shaped, dumb or acute apex, and hairy on both sides. The petiole is thin and hairy; the odor is aromatic.
Chaal	Color: Greyish brown from the outside and pale pink from the inside, longitudinal cracks are present.
Pushpa	Purplish or crimson-colored, calyx elliptical or campanulated 3-4 mm blipped, odor is aromatic, taste is pungent.
Phal	Four nutlets, each containing one seed, membranous, color is dull brown or reddish with small black patches, odor is aromatic, taste is pungent.
Beej	The shape is oval, the color is brown, mucilaginous when soaked in water, the odorless, taste is pungent.

Extract and the Part of Tulsi Plant Used For Pharmacological Activities

Therapeutic activity	Extract used	Part used
Anti-stress	Ethanolic	Whole plant (dried)
Anti-inflammatory	Methanolic/aqueous	Leaves
Anti-fungal	Methanolic/aqueous	Leaves
Anti-fertility	Benzene	Leaves
Hepatoprotective	Ethanolic/aqueous	Whole plant (aerial)
Anti-diabetic	Ethanolic/aqueous	Leaves
Anti-ulcer	Ethanolic/aqueous	Leaves
Anti-microbial	Ethanolic	Leaves
Anti-psychotic	Methanolic/ leaves paste	Leaves
Anti-cancer	Ethanolic	Root

CONCLUSION

In conclusion, Tulsi, or holy basil, is a revered plant with a rich history of medicinal and culinary use on the Indian subcontinent. It has been employed in Ayurvedic medicine for over 3000 years and is recognized for its medicinal properties in treating various health concerns. Tulsi has been thoroughly examined, revealing many pharmacological qualities such as adaptogenic, immunomodulatory, antioxidant, and antibacterial activities. Recent studies indicate that Tulsi possesses antistress, anticonvulsant, and anxiolytic qualities, along with potential advantages in enhancing glycemic and lipid profiles.

Numerous kinds of Tulsi exist, each exhibiting distinct phytochemical compositions and morphological characteristics, however all are historically employed to treat analogous ailments. Moreover, additional species in the *Ocimum* genus, including *Ocimum sanctum*, are acknowledged for their therapeutic virtues and have been employed in many medical systems throughout for millennia. *Sanctum* possesses a diverse array of therapeutic characteristics, including antipyretic, anti-inflammatory, expectorant, and stimulating actions. It has been utilized to address different conditions, including malaria, rheumatism, and gastrointestinal issues.

Recent scientific studies have demonstrated that constituents of Tulsi and *Sanctum* possess various biological and pharmacological actions, including anticancer effects. Research indicates that extracts from these plants can trigger apoptosis in cancer cells, diminish intracellular glutathione levels, and augment the activity of certain enzymes associated with chemoprevention. Preclinical studies indicate that Tulsi extracts exhibit cytotoxic effects on cancer cells and may impede tumor cell adhesion and invasion. The active constituents of Tulsi and *Sanctum* have demonstrated potential in suppressing the proliferation and spread of several

cancer cell lines.

Tulsi and Sanctum possess a longstanding history of traditional application and have significant potential in contemporary scientific study about their therapeutic attributes, particularly as anticancer agents. Additional study is required to comprehensively elucidate the mechanisms of action and prospective therapeutic implications of these plants in cancer therapy and prevention.

REFERENCES

1. K. Nadkarni and A. Nadkarni, Indian Materia Medica with Ayurvedic, Unani-Tibbi, Siddha, Allopathic, Homeopathic, Naturopathic & Home Remedies, vol. 2, Popular Prakashan Private Ltd, Bombay, India, 1982.
2. A. P. Committee, The Ayurvedic Pharmacopoeia of India, Part I, Volume IV, Government of India, Ministry of Health and Family Welfare, Department of Ayurveda, Yoga & Naturopathy, Unani, Siddha, and Homoeopathy (AYUSH), New Delhi, India, 1st edition, 2016.
3. S. S. Hebbar, V. H. Harsha, V. Shripathi, and G. R. Hegde, "Ethnomedicine of Dharwad district in Karnataka, India—plants used in oral health care," Journal of Ethnopharmacology, 2004; 94(2-3): 261–266.
4. H. J. Dadysett, "On the various domestic remedies, with their effects, used by the people of India for certain diseases of the ear," The Lancet, 1899; 154(3968): 781–782.
5. R. Chopra and I. Chopra, "Glossary of Indian Medicinal Plants," Council of Scientific & Industrial Research, New Delhi, India, 1992.
6. S. K. Kothari, A. K. Bhattacharya, S. Ramesh, S. N. Garg, and S. P. S. Khanuja, "Volatile constituents in oil from different plant parts of methyl eugenol-rich *Ocimum tenuiflorum* L.f. (syn. *O. sanctum* L.) grown in South India," Journal of Essential Oil Research, 2005; 17(6): 656–658.
7. J. A. Parrotta, Healing Plants of Peninsular India, CABI, Oxfordshire, UK, 2001.
8. C. Orwa, A. Mutua, R. Kindt, R. Jamnadass, and A. Simons, Agroforestry Database: A Tree Species Reference and Selection Guide Version 4.0, World Agroforestry Centre ICRAF, Nairobi, Kenya, 2009.
9. S. Bhamra, M. Heinrich, C. Howard, M. Johnson, and A. Slater, "DNA authentication of tulsi (*Ocimum tenuiflorum*) using the nuclear ribosomal internal transcribed spacer (ITS) and the chloroplast intergenic spacer trnH-psbA," Planta Medica, 2015; 81(16): PW_20.

10. T. Chowdhury, A. Mandal, S. C. Roy, and D. De Sarker, "Diversity of the genus *Ocimum* (Lamiaceae) through morphomolecular (RAPD) and chemical (GC–MS) analysis," *Journal of Genetic Engineering and Biotechnology*, 2017.
11. K. Carovic-Stanko, Z. Liber, V. Besendorfer, et al., "Genetic' relations among basil taxa (*Ocimum* L.) based on molecular markers, nuclear DNA content, and chromosome number," *Plant Systematics and Evolution*, 2010; 285(1): 13–22.
12. E. Jothie Richard, R. Illuri, B. Bethapudi et al., "Anti-stress activity of *Ocimum sanctum*: possible effects on hypothalamic–pituitary–adrenal axis," *Phytotherapy Research*, 2016; 30(5): 805–814.
13. M. P. Venu Prasad and F. Khanum, "Antifatigue activity of Ethanolic extract of *Ocimum sanctum* in rats," *Research Journal of Medicinal Plant*, 2012; 6(1): 37–46.
14. I. Tabassum, Z. N. Siddiqui, and S. J. Rizvi, "Effects of *Ocimum sanctum* and *Camellia sinensis* on stress-induced anxiety and depression in male albino *Rattus norvegicus*," *Indian Journal of Pharmacology*, 2010; 42(5): 283–288.
15. T. Suanarunsawat, G. Anantasomboon, and C. Piewbang, "Antidiabetic and anti-oxidative activity of fixed oil extracted from *Ocimum sanctum* L. leaves in diabetic rats," *Experimental and Therapeutic Medicine*, 2016; 11(3): 832–840.
16. I. Husain, R. Chander, J. K. Saxena, A. A. Mahdi, and F. Mahdi, "Antidyslipidemic effect of *Ocimum sanctum* leaf extract in Streptozotocin-induced diabetic rats," *Indian Journal of Clinical Biochemistry*, 2015; 30(1): 72–77.
17. T. Raja, R. R. N. Reddy, and M. B. Priyadharshini, "An evaluation of the anti-hyperglycemic activity of *Ocimum sanctum* Linn (leaves) in Wister rats," *The Pharma Innovation Journal*, 2016; 5(1): 1–3.
18. F. J. Sutili, D. M. Gatlin, W. Rossi, B. M. Heinzmann, and B. Baldisserotto, "In vitro effects of plant essential oils on nonspecific immune parameters of red drum, *Sciaenops ocellatus* L.," *Journal of Animal Physiology and Animal Nutrition*, 2016; 100(6): 1113–1120.
19. D. Panprommin, W. Kaewpunnin, and D. Insee, "Effects of holy basil (*Ocimum sanctum*) extract on the growth, immune response and disease resistance against *Streptococcus agalactiae* of Nile tilapia (*Oreochromis niloticus*)," *International Journal of Agriculture and Biology*, 2016; 18(4): 677–682.
20. S. Godhwani, J. L. Godhwani, and D. S. Was, "*Ocimum sanctum*- a preliminary study evaluating its immunoregulatory profile in albino rats," *Journal of Ethnopharmacology*, 1988; 24(2-3): 193–198.

21. K. Aruna and V. M. Sivaramakrishnan, "Anticarcinogenic effects of some Indian plant products," Food and Chemical Toxicology, 1992; 30(11): 953–956.
22. S. Banerjee, R. Prashar, A. Kumar, and A. R. Rao, "Modulatory influence of alcoholic extract of Ocimum leaves on carcinogen metabolizing enzyme activities and reduced glutathione levels in mouse," Nutrition and Cancer, 1996; 25(2): 205–217.
23. C.-C. Lin, P.-Y. Chao, C.-Y. Shen, et al., "Novel target genes responsive to apoptotic activity by ocimum gratissimum in human osteosarcoma cells," American Journal of Chinese Medicine, 2014; 42(3): 743–767.
24. S. Godhwani, J. L. Godhwani, and D. S. Vyas, "Ocimum sanctum: an experimental study evaluating its anti-inflammatory, analgesic and antipyretic activity in animals," Journal of Ethnopharmacology, 1987; 21(2): 153–163.
25. Y. Tanko, G. M. Magaji, M. Yerima, R. A. Magaji, and A. Mohammed, "Antinociceptive and anti-inflammatory activities of aqueous leaves extract of Ocimum gratissimum (Labiata) in Rodents," African Journal of Traditional, Complementary and Alternative Medicines, 2008; 5(2): 141–146.
26. A. C. Akinmoladun, E. Ibukun, E. Afor, E. Obuotor, and E. Farombi, "Phytochemical constituent and antioxidant activity of extract from the leaves of Ocimum gratissimum," Scientific Research and Essays, 2007; 2(5): 163–166.
27. M. A. Kelm, M. G. Nair, G. M. Strasburg, and D. L. DeWitt, "Antioxidant and cyclooxygenase inhibitory phenolic compounds from Ocimum sanctum Linn.," Phytomedicine, 2000; 7(1): 7–13.
28. H.-C. Chang, Y.-W. Chiu, Y.-M. Lin et al., "Herbal supplement attenuation of cardiac fibrosis in rats with CCl₄-induced liver cirrhosis," Chinese Journal of Physiology, 2014; 57(1): 41–47.
29. R. Chattopadhyay, S. Sarkar, S. Ganguly, C. Medda, and T. Basu, "Hepatoprotective activity of Ocimum sanctum leaf extract against paracetamol-induced hepatic damage in rats," Indian Journal of Pharmacology, 1992; 24(3): 163.
30. P. U. Devi and A. Ganasoundari, "Radioprotective effect of leaf extract of Indian medicinal plant Ocimum sanctum," Indian Journal of Experimental Biology, 1995; 33(3): 205–208.
31. P. U. Devi, A. Ganasoundari, B. S. S. Rao, and K. K. Srinivasan, "In vivo radioprotection by Ocimum flavonoids: survival of mice," Radiation Research, 1999; 151(1): 74–78.
32. M. S. Rahman, M. M. Khan, and M. A. Jamal, "Anti-bacterial evaluation and minimum inhibitory concentration analysis of Oxalis corniculata and Ocimum santum against

- bacterial pathogens,” *Biotechnology*, 2010; 9(4): 533–536.
33. G. Prasad, A. Kumar, and A. K. Singh, “Antimicrobial activity of essential oils of some *Ocimum* species and clove oil,” *Fitoterapia*, 1986; 57(6): 429–432.
34. M. P. Prasad, K. Jayalakshmi, and G. G. Rindhe, “Antibacterial activity of *Ocimum* species and their phytochemical and antioxidant potential,” *International Journal of Microbiology Research*, 2012; 4(8): 302–307.
35. C. V. Nakamura, T. Ueda-Nakamura, E. Bando, A. F. Negrao~ Melo, D.A.GarciaCortez, and B.P.DiasFilhoFilho, “Antibacterial activity of *Ocimum gratissimum* L. essential oil,” *Memorias do Instituto Oswaldo Cruz*, 1999; 94(5): 675–678.
36. R. R. Chattopadhyay, “Hypoglycemic effect of *Ocimum sanctum* leaf extract in normal and streptozotocin diabetic rats,” *Indian Journal of Experimental Biology*, 1993; 31(11): 891–893.
37. J. C. Aguiyi, C. I. Obi, S. S. Gang, and A. C. Igweh, “Hypoglycaemic activity of *Ocimum gratissimum* in rats,” *Fitoterapia*, 2000; 71(4): 444–446.
38. V. Rai, U. Iyer, and U. V. Mani, “Effect of Tulasi (*Ocimum sanctum*) leaf powder supplementation on blood sugar levels, serum lipids, and tissues lipids in diabetic rats,” *Plant Foods for Human Nutrition*, 1997; 50(1): 9–16.
39. N. Srinivas, K. Sali, and A. Bajoria, “Therapeutic aspects of Tulsi unraveled: a review,” *Journal of Indian Academy of Oral Medicine and Radiology*, article no. 17, 2016; 28(1).
40. M. S. Baliga, S. Rao, M. P. Rai, and P. D’Souza, “Radioprotective effects of the Ayurvedic medicinal plant *Ocimum sanctum* Linn. (Holy Basil): a memoir,” *Journal of Cancer Research and Therapeutics*, 2016; 12(1): 20–27.
41. H. R. D. Fonseka, W. M. S. S. K. Kulathunga, A. Peiris, and L. D. A. M. Arawwawala, “A review on the therapeutic potentials of *Ocimum sanctum* Linn: in the management of diabetes Mellitus (Madhumeha),” *Journal of Pharmacognosy and Phytochemistry*, 2015; 4(3).
42. M. M. Cohen, “Tulsi—*Ocimum sanctum*: a herb for all reasons,” *Journal of Ayurveda and Integrative Medicine*, 2014; 5(4): 251–259.
43. A. A. Kamyab and A. Eshraghian, “Anti-inflammatory, gastrointestinal and hepatoprotective effects of *Ocimum sanctum* Linn: an ancient remedy with new application,” *Inflammation and Allergy - Drug Targets*, 2013; 12(6): 378–384.
44. S. Vishwabhan, V. K. Birendra, and S. Vishal, “A review on ethnomedical uses of *Ocimum sanctum* (Tulsi),” *International Research Journal of Pharmacy*, 2011; 2: 1–3.

45. P. Pattanayak, P. Behera, D. Das, and S. K. Panda, "Ocimum sanctum Linn. A reservoir plant for therapeutic applications: an overview," *Pharmacognosy Reviews*, 2010; 4(7): 95–105.
46. K. P. Bhargava and N. Singh, "Anti-stress activity of *Ocimum sanctum* Linn," *The Indian Journal of Medical Research*, 1981; 73: 443–451.
47. M. R. Sakina, P. C. Dandiya, M. E. Hamdard, and A. Hameed, "Preliminary psychopharmacological evaluation of *Ocimum sanctum* leaf extract," *Journal of Ethnopharmacology*, 1990; 28(2): 143–150.
48. S. Sood, D. Narang, M. K. Thomas, Y. K. Gupta, and S. K. Maulik, "Effect of *Ocimum sanctum* Linn. on cardiac changes in rats subjected to chronic restraint stress," *Journal of Ethnopharmacology*, 2006; 108(3): 423–427.
49. E. Mitra, D. Ghosh, A. K. Ghosh, et al., "Aqueous Tulsi leaf (*Ocimum sanctum*) extract possesses antioxidant properties and protects against cadmium-induced oxidative stress in rat heart," *International Journal of Pharmacy and Pharmaceutical Sciences*, 2014; 6(1): 500–513.
50. C. O. Okoli, A. C. Ezike, O. C. Agwagah, and P. A. Akah, "Anticonvulsant and anxiolytic evaluation of leaf extracts of *Ocimum gratissimum*, a culinary herb," *Pharmacognosy Research*, 2010; 2(1): 36–40.
51. F. C. M. Okomolo, J. T. Mbafor, E. N. Bum, et al., "Evaluation of the sedative and anticonvulsant properties of three Cameroonian plants," *African Journal of Traditional, Complementary and Alternative Medicines*, 2011; 8(5): 181–190.
52. M. L. Dhar, M. M. Dhar, B. N. Dhawan, B. N. Mehrotra, and C. Ray, "Screening of Indian plants for biological activity: I," *Indian Journal of Experimental Biology*, 1968; 6(4): 232–247.
53. J. Giri, B. Suganthi, and G. Meera, "Effect of Tulasi (*Ocimum sanctum*) on diabetes mellitus," *The Indian Journal of Nutrition and Dietetics*, 1987; 24: 193–198.
54. A. Sarkar, S. C. Lavana, D. N. Pandey, and M. C. Pant, "Changes in the blood lipid profile after administration of *Ocimum sanctum* (Tulsi) leaves in the normal albino rabbits," *Indian Journal of Physiology and Pharmacology*, 1994; 38(4): 311–312.
55. S. Gupta, P. K. Mediratta, S. Singh, K. K. Sharma, and R. Shukla, "Antidiabetic, antihypercholesterolaemic and antioxidant effect of *Ocimum sanctum* (Linn) seed oil," *Indian Journal of Experimental Biology*, 2006; 44(4): 300–304.
56. R. Patil, R. Patil, B. Ahirwar, and D. Ahirwar, "Isolation and characterization of anti-diabetic component (bioactivity—guided fractionation) from *Ocimum sanctum* L.

- (Lamiaceae) aerial part,” Asian Pacific Journal of Tropical Medicine, 2011; 4(4): 278–282.
57. A.Chandra,A.A.Mahdi,R.K.Singh,F.Mahdi, and R. Chander, “Effect of Indian herbal hypoglycemic agents on antioxidant capacity and trace elements content in diabetic rats,” Journal of Medicinal Food, 2008; 11(3): 506–512.
58. S. S. Reddy, R. Karuna, R. Baskar, and D. Saralakumari, “Prevention of insulin resistance by ingesting aqueous extract of *Ocimum sanctum* to fructose-fed rats,” Hormone and Metabolic Research, 2008; 40(1): 44–49.
59. R. Mukherjee, P. K. Dash, and G. C. Ram, “Immunotherapeutic potential of *Ocimum sanctum* (L) in bovine subclinical mastitis,” Research in Veterinary Science, 2005; 79(1): 37–43.
60. P. K. Kundu and P. Chatterjee, “Meta-analysis of Diabecon tablets: efficacy and safety,” Indian Journal of Clinical Practice, 2010; 20(9): 653.
61. Kumar V, Andola HC, Lohani H, Chauhan N. Pharmacological review on *Ocimum sanctum* Linnaeus: a queen of herbs. J Pharm Res., 2011; 4: 366–368.
62. Kirtikar KR, Basu BD. The *ocimum sanctum* in Indian medicinal plants. Allahabad: LB Basu, 1965.
63. Pandey BP. Anita in economic botany. New Delhi: Chand and Company Limited, 1990.
64. Gupta SK, Prakash J, Shrivastava S. Validation of traditional claim of Tulsi, *Ocimum sanctum* Linn. as a medicinal plant. Indian J Exp Biol., 2002; 40: 765–773.
65. Ravi P, Elumalai A, Cinna Eswaraiah M, Kasarla R. A review on Krishna Tulsi, *Ocimum tenuiflorum* Linn. Int J Res Ayurveda Pharm, 2012; 3: 291–293.
66. Pandey G, Madhuri S. Pharmacological activities of *Ocimum sanctum* (Tulsi): a review. Int J Pharm Sci Rev Res., 2010; 5: 61–66.
67. Anonymous. The Wealth of India, Raw Materials Series. New Delhi: Council of Scientific and Industrial Research, 1991.
68. Mondal S, Mirdha BR, Mahapatra SC. The science behind sacredness of Tulsi (*Ocimum sanctum* Linn.). Indian J Physiol Pharmacol, 2009; 53: 291–306.
69. Garodia P, Ichikawa H, Malani N, Sethi G, Aggarwal BB. From ancient medicine to modern medicine: Ayurvedic concepts of health and their role in inflammation and cancer. J Soc Integr Oncol., 2007; 5: 25–37.
70. Mohan L, Ambedkar MV, Kumari M. *Ocimum sanctum* Linn. (Tulsi) – an overview. Int J Pharm Sci Rev Res., 2011; 7: 51–53.
71. Nadkarni GB, Nadkarni KM. Indian Materia Medica. Bombay: Popular Book Depot.,

1954.

72. Kirtikar KR, Basu BD. Indian Medicinal Plant. Dehradun: Bishen Singh Mahendra Pal Singh, 1975.
73. Khosla MK. Sacred Tulsi (*Ocimum sanctum* L.) in traditional medicine and pharmacology. *Anc Sci Life.*, 1995; 15: 53–61.
74. Uma Devi P. Radioprotective, anticarcinogenic, and antioxidant properties of the Indian holy basil, *Ocimum sanctum* (Tulasi). *Indian J Exp Biol.*, 2001; 39: 185–190.
75. Singh S, Taneja M, Majumdar DK. Biological activities of *Ocimum sanctum* L. fixed oil – an overview. *Indian J Exp Biol.*, 2007; 45: 403–412.
76. Pattanayak P, Behera P, Das D, Panda SK. *Ocimum sanctum* Linn. A reservoir plant for therapeutic applications: an overview. *Pharmacogn Rev.*, 2010; 4: 95–105.
77. Karthikeyan K, Gunasekaran P, Ramamurthy N, Govindasamy S. Anticancer activity of *Ocimum sanctum*. *Pharm Biol.*, 1999; 37: 285–290.
78. Niture SK, Rao US, Srivenugopal KS. Chemopreventive strategies targeting the MGMT repair protein: augmented expression in human lymphocytes and tumor cells by ethanolic and aqueous extracts of several Indian medicinal plants. *Int J Oncol.*, 2006; 29: 1269–1278.
79. Magesh V, Lee JC, Ahn KS, Lee HJ, Lee HJ, Lee EO, et al. *Ocimum sanctum* induces apoptosis in A549 lung cancer cells and suppresses the in vivo growth of Lewis lung carcinoma cells. *Phytother Res.*, 2009; 23: 1385–1391.
80. Kim SC, Magesh V, Jeong SJ, Lee HJ, Ahn KS, Lee HJ, et al. Ethanol extract of *Ocimum sanctum* exerts anti-metastatic activity through the inactivation of matrix metalloproteinase-9 and enhancement of antioxidant enzymes. *Food Chem Toxicol.*, 2010; 48: 1478–1482.