

WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 8.084

Volume 10, Issue 12, 1864-1874.

Research Article

ISSN 2277-7105

SCREENING OF SELECTED PLANT EXTRACTS FOR THEIR LOCALIZATION OF PHYTOCHEMICALS AND ANTIMICROBIAL **ACTIVITY**

Jancy Mathew^{1*}, Jampala Siva Satya Mohan² and Vinay M. Raole¹

¹Department of Botany, The Maharaja Sayajirao University of Baroda, Vadodara (390002), Gujarat, India.

Article Received on 09 Aug. 2021,

Revised on 30 Aug. 2021, Accepted on 20 Sept. 2021 DOI: 10.20959/wjpr202112-21837

*Corresponding Author Jancy Mathew

Department of Botany, The Maharaja Sayajirao University of Baroda, Vadodara (390002), Gujarat, India.

ABSTRACT

Traditional medicine particularly herbal medicine considered as a major healthcare provider around the globe, particularly in rural and remote areas. Herbal medicines are gaining much importance nowadays as it is cheap, easily available and have lesser side effects. The current investigation deals with efficacy of methanolic leaf extracts by carrying out the phytochemical screening and qualitative analysis as well as to check their efficacy against selected antibiotic susceptible and resistant microorganisms. Phytochemicals obtained from selected leaf extract of seven taxa were alkaloids, tannins, flavonoids, saponins, steroids and phenol which were traditionally used to treat cough, low B.P, respiratory infections and gonorrhea. Over and

above, the results confirmed the ethnomedicinal uses of four plant taxa by showing the inhibitory action against microorganisms; among which the most significant one is *Polyalthia* longifolia, against Escherichia coli, Pseudomonas aeruginosa, Staphylococcus aureus, Salmonella typhi and Serratia marcescens.

KEYWORDS: Ethnomedicinal, Microorganism, Phytochemical, Screening.

INTRODUCTION

Medicinal plants are not only used for the treatment of diseases but also as a potential material for maintaining good health. Many countries in the world, depends upon herbal medicines because of their availability locally, cultural acceptability, better compatibility and adaptability with the human body physiology and has lesser side effects. [1] Infectious and non

ISO 9001:2015 Certified Journal www.wjpr.net Vol 10, Issue 12, 2021.

²Department of Biosciences, Sardar Patel University, Anand (388120), Gujarat, India.

infectious diseases represent an important cause of morbidity and mortality among the human population, particularly in developing and underdeveloped nations. Therefore, a search to develop new antimicrobial drugs in recent years, especially due to the constant emergence of many microorganisms resistant to conventional antimicrobials. Scientific validation of active constituents extracted from the plants can be used in drug discovery in the future.

According to the World Health Organization (2002) medicinal plants would be the best source to obtain an assortment of drugs and drug plants. Many plants as a whole are loaded with number of secondary metabolites such as tannins, alkaloids, phenolic compounds, and flavonoids which have been found to have antimicrobial properties during in vitro testing.^[2]

In India, about 65% of total^[3] and 85% of the rural^[4] population depend on such traditional knowledge for healthcare. India has a tremendous wealth of medicinal plants due to its unique geography, climate and environmental conditions.^[5] The country has an ancient cultural background and about 300 tribal communities with 53 million population.^[6] These people are using 7000-7500 plants to overcome different kind of health problems.^[7-9] The aim of the study is to validate the plant species scientifically based on the antimicrobial screening and qualitative analysis of phytochemicals and comparing it with the available literature of ethnobotanical uses of the selected plants. Though medicinal uses of plants were reported by many researchers but scientific validation of ethno-botanically significant flora is not reported much in Gujarat, India. By and large, there is no detail systematic study conducted till date.

MATERIALS AND METHODS

Collection of different plant materials

Seven plant species named *Aegle marmelos* (L.) Correa, *Catharanthus roseus* (L.) G. Don, *Melia azedarach* L., *Neolamarckia cadamba* (Roxb.) Bosser, *Nerium indicum* L., *Polyalthia longifolia* (Sonn.) Thwaites, *Tamarindus indica* L. belonging to 6 different families-Rutaceae, Apocynaceae, Meliaceae, Rubiaceae, Annonaceae and Fabaceae were collected in the form of leaves from Vallabh Vidhyanagar, Anand. Identity of selected plant taxa was also confirmed with Flora of Gujarat state.^[10]

Preparation of leaf extract

Healthy leaves of all the selected plants were washed thoroughly under running tap water to remove the dust particles and were dried by keeping it in oven for 4-5 days at 45°C and

powdered using mortar and pestle. Extract was prepared by infusion extract method given by Houghton and Raman.^[11] For extraction 10g of dry powder of each sample was soaked in 100 ml of methanol at room temperature for 24 hours. Extracts were filtered through Whatman filter paper no.1 and the filtrates were centrifuged at 3000 rpm for 10 minutes to remove solid debris. The supernatant was collected and stored in refrigerator until further use.

Qualitative analysis of crude plant extracts

Qualitative analysis was carried out by standard procedures described by Houghton and Raman^[11] to identify the various constituents present in crude leaf extracts of selected plants prepared in organic solvents.

Dragendroff's test for presence of alkaloids

The extract is treated with few drops of Dragendroff's reagent. The orange brown precipitate coloration if observed indicates the presence of alkaloids.

Braymer's test for presence of tannins

2ml extract is treated with 2 ml water and 2-3 drops of 5% FeCl₃. The blue black precipitate if observed indicates the presence of tannins.

Sodium hydroxide test for presence of flavonoids

1ml extract is treated with 1ml of 10% NaOH. The yellow color indicates the presence of flavonoids.

Acetic anhydride test for presence of terpenoids

2 ml extract is treated with 2ml acetic acid and 2-3 drops of concentrated H₂SO₄. The deep red color indicates the presence of terpenoids.

Foam test for presence of saponins

5 ml extract is treated with 5 ml distilled water and both were mixed by shaking. The foam production indicates the presence of saponins.

Test for steroids

2 ml extract is treated with 2 ml chloroform and 2 ml concentrated H₂SO₄. The brown colored ring between the sample and H₂SO₄ indicates the presence of steroids.

Ferric chloride test for phenols

Few drops of extract are treated with few drops of 5 % FeCl₃. The dark green color indicates the presence of phenol.

Selected microorganisms for Bioassay

The microorganisms selected for the experiment are one gram-positive bacteria-Staphylococcus aureus and four gram-negative bacteria- Escherichia coli, Salmonella typhi, Pseudomonas aeruginosa and Serratia marcescens. The bacterial cultures were grown on nutrient broth medium (Hi Media pH 7.4) at 37°C and maintained at 4°C.

Inoculum preparation

A fresh microbial suspension was prepared by subculturing the bacterial colonies into the nutrient broth medium (Hi Media ph 7.4) and incubated at 37°C in order to maintain approximately uniform growth rate of each organism. The bacterial cultures from the fresh media was compared with 0.5 Mc Farland turbidity standard, which is equivalent to approximately 1x10⁸ bacterial cell density^[12] was used throughout the experimentation.

Bioassay

The present study, the antimicrobial activity of different plant methanolic extracts were screened by agar well diffusion method. [13] A stock solution (100mg/ml) of each plant extract was prepared in DMSO for antibacterial activity. Approximately 20 ml sterilized nutrient agar medium was poured into the sterile petriplates in aseptic conditions using laminar air flow cabinet and then allowed to solidify at room temperature. The prepared agar plates were then marked and labeled with organism and extract name. Fresh bacterial culture inoculum of 100µl having 1x108CFU/ml was spread on agar plates with sterile glass spreader. A well of 8mm diameter punched off at previously marked petriplates into agar medium with sterile cork borer and then was filled with 100µl of each plant extract. Plates were placed in refrigerator for 30 minutes for pre diffusion of plant extract and then incubated at 37°C for 24 hours until the appearance of inhibition zone (excluding well diameter) was measured as a property of antibacterial activity. The assays were conducted in duplicates.

RESULTS AND DISCUSSION

Methanolic plant extracts prepared by infusion extract method from leaves of seven different plant species i.e., A. marmelos, C. roseus, M. azedarach, N. cadamba, N. indicum, P. longifolia, T. indica were selected as experimental materials to compare its efficacy with the ethnobotanical uses recorded on the basis of phytochemical analysis and antimicrobial screening.

Phytochemical analysis

Many phytochemical constituents such as alkaloids, tannins, flavonoids, terpenoids, saponins, steroids, phenol, glycosides and several other aromatic compounds are secondary metabolites produced by plants for their defense mechanism against many microorganisms, insects and herbivores. [14] In the present study leaf extracts of *Aegle marmelos, Catharanthus roseus, Melia azedarach, Neolamarckia cadamba, Nerium indicum, Polyalthia longifolia, Tamarindus indica* prepared in methanol were used for presence of phytochemical constituents such as alkaloids, tannins, flavonoids, terpenoids, saponins, steroids and phenols.

All the leaf extracts of selected plants ie: *Aegle marmelos, Catharanthus roseus, Melia azedarach, Nerium indicum, Polyalthia longifolia, Tamarindus indica* prepared in methanol posses tannins, terpenoids, saponins, steroids and phenol. Alkaloids were present in all the selected plant extracts except *Neolamarckia cadamba*.

In *Aegle marmelos*, the phytochemicals obtained from methanol leaf extracts are alkaloids, tannins, flavonoids, terpenoids, saponins, steroids, phenol. The obtained results were agreeing with study conducted by Mujeeb et al.^[15] In *Catharanthus roseus*, as reported by Aziz et al,^[16] Kabesh et al,^[17] phytochemicals obtained were alkaloids, tannins, terpenoids, saponins, steroids and phenols. In *Melia azedarach*, alkaloids, tannins, flavonoids, saponins, steroids were present and the results were same as mentioned by Muhammad et al,^[18] and Farook et al.^[19] As reported by Ganjewala et al,^[20] and Kumar et al,^[21] the results obtained were tannins, flavonoids, saponins, steroids and phenol in *Neolamarckia cadamba*. In *Polyalthia longifolia*, the results obtained were alkaloids, tannins, flavonoids, saponins, steroids, phenol as mentioned by Arun and Chandrashekhar^[22] and Mudhafar et al,^[23] and in *Tamarindus indica* the phytochemicals obtained were alkaloids, tannins, flavonoids, terpenoids, steroids, phenols and these results were similar to the one reported by Arora et al,^[24] Abdallah and Muhammad.^[25]

The phytochemical analysis of different plant methanolic extracts from the experiment confirmed the presence of phytochemicals extracted in previous screening by different researchers, mentioned above.

Antimicrobial activity of plant extracts

The data pertaining to the antibacterial activity of plant extracts and inhibition zone formed by them are listed in the Table I. It was revealed from the results that each medicinal plant shows different degree of inhibition zone against selected microorganism in the study. The diameter of inhibition zone depends upon extrinsic and intrinsic factors. The extrinsic factors like pH of the medium, period and temperature of incubation, size of the well, volume of the inoculums, concentration of plant extracts can be fixed and standardized during the experiment and hence no error due to extrinsic factors. The intrinsic factors such as nature of medicinal plants including its components, solubility, and diffusing property are predetermined.

Methanolic extract of P. longifolia inhibited significantly against all the tested microorganisms with the highest inhibition zone of 22mm against E. coli, P. aeruginosa and S. typhi followed by S. aureus (20mm) and S. marcescens (18 mm).

The maximum activity was shown by *Polyalthia longifolia*, followed by *Nerium indicum*, Tamarindus indica, Neolamarckia cadamba, Catharanthus roseus, Aegle marmelos and *Melia azedarach* against the tested microorganisms.

Table I: Antibacterial activity of leaf extracts in methanol.

Sr no.	Tested microorganisms	Diameter of the zone of inhibition in mm shown by methanol plant extracts against tested microorganisms						
		AM	CR	MA	NC	NI	PL	TI
1	E. coli	10	10	8	8	16	22	18
2	P. aeruginosa	8	8	3	10	12	22	12
3	S. aureus	4	16	8	10	12	20	10
4	S. typhi	10	6	8	10	10	22	6
5	S. marcescens	0	8	6	12	16	18	16

AM- Aegle marmelos, CR- Catharanthus roseus, MA-Melia azedarach, NC- Neolamarckia cadamba, NI-Nerium indicum, PL-Polyalthia longifolia, TI- Tamarindus indica.

The earlier study has reported the moderate antimicrobial activity of methanol leaf extract against Escherichia coli and Staphlococcus aureus in all the selected plants; Aegle marmelos, [26] Catharanthus roseus, [27] Neolamarckia cadamba, [28] Nerium indicum, [29] Polyalthia longifolia, [30-31] and better activity against Salmonella typhi in Polyalthia longifolia.[31]

Plant extracts prepared in methanol showed a broad range of activity against the tested microorganisms. E. coli, P. aeruginosa, S. aureus were most susceptible organisms and S. *marcescens* was the most resistant one among all.

Comparision of ethnobotanical uses with its efficacy towards microorganisms

Aegle marmelos: The ethnomedicinal uses reported by Dutta et al^[32], reveals that the leaf extract is used to treat diseases like diarrhea (EC, ST), respiratory infections (PA, SM) abscess (SA) high fever (ST) caused by the tested organisms E.coli, S. typhi, P. aeruginosa, S. marcescens, and S.aureus.

Neolamarckia cadamba: The ethnomedicinal uses reported by Negi^[33] indicates that the leaf extract is used to treat diseases like diarrhea (EC, ST), skin diseases (PA, SA) conjunctivitis and eye infections (SM) caused by the tested organisms E.coli, S. typhi, P. aeruginosa, S. marcescens, and S.aureus.

Polyalthia longifolia: The ethnomedicinal uses reported by Subramanion et al^[34] reveals that the leaf extract is used to treat fever (ST), skin diseases (PA) diarrhea (EC, ST) respiratory infections (PA, SM) caused by the tested organisms E.coli, S. typhi, P. aeruginosa, S. marcescens, and S.aureus.

Tamarindus indica: The ethnomedicinal uses reported by Bhadoriya et al^[35] discloses that the leaf extract is used to treat respiratory problems (PA,SM), fever (ST), diarrhea(EC, ST), eye infections (SM) caused by the tested organisms E.coli, S. typhi, P. aeruginosa, S. marcescens, and S.aureus.

From the results after comparing with the available literature of ethnobotanical uses, it revealed that the methanolic leaf extract of Aegle marmelos, Neolamarckia cadamba, Polyalthia longifolia, and Tamarindus indica showed the antimicrobial efficacy against the tested microorganisms which caused the problems like abscess formation, vomiting, cold, cough, bronchitis, diarrhea, fever, skin diseases, respiratory infections, eye diseases etc.

CONCLUSION

Although, some plant part extracts show evidence of good antibacterial potency, in contrast to our expectation. Over and above, limited antibacterial potency of some plants suggests that there is no concurrence between the traditional uses of medicinal plants in the crude form for the remedy of infectious diseases. Another possibility for the limited antibacterial potency of some plants may be due to the extraction method and the use of crude extracts. Even though we showed potent in vitro activity of a few traditional plant extracts for certain bacteria, it may not be translated in vivo. However, further detail study is necessary to explore their effectiveness in inhibiting the growth of different pathogens. From the current investigation, it can be concluded after comparision of ethnomedicinal uses with results of anitmicrobial screening against selected microorganisms and qualitative analysis four plants namely-Aegle marmelos, Neolamarckia cadamba, Polyalthia longifolia, Tamarindus indica among the seven selected plants can be validated scientifically based on the information available in the literature.

ACKNOWLEDGEMENT

I am thankful to the Head, of B R Doshi School of Biosciences, Sardar Patel University, Vallabh Vidhyanagar, Anand, and The Maharaja Sayajirao university of Baroda, Vadodara Gujarat, India for providing me the permission and facility for conducting the research. And I am also thankful to Shodh and FIST programme for the financial assistance.

REFERENCES

- 1. Oladeji O. (The Characteristics and roles of medicinal plants: Some important medicinal plants in Nigeria). Nat Prod Ind J, 2016; 12(3): 102.
- 2. Djeussi DE, Noumedem JAK, Seukep JA et al. (Antibacterial activities of selected edible plants extracts against multidrug-resistant Gram-negative bacteria). BMC Complement and Altern Med, 2013; 13(164).
- 3. Timmermans & Karin. (Intellectual Property Rights and Traditional medicine: policy dilemmas at the interface). Soc Sci Med, 2003; 57(4): 745-756.
- 4. Jain SK. Ethnopharmacology and drug development. In: Ethnobotany & Search for New drugs. Chadwick DJ, & March U, John Wiley & Sons, New York, USA, 1994.
- 5. Kshirsagar RD & Singh NP. (Less known ethnomedicinal uses of plants in Coorg district of Karnataka state, Southern India). Ancient Sci Life, 2000; 20(3): 20-25.
- 6. Reddy KN, Trimurthulu G & Reddy CS. (Medicinal plants used by ethnic people of Medak district, Andhra Pradesh). Indian J Tradit Knowl, 2010; 9: 184-190.
- 7. Matthews S. Ayurveda. In: an Introduction to complementary medicine, Roboson T (ed) Allen & Unwin, Crows Nest, NSW, 2005; 15-32.
- 8. Mao AA, Hynniewta TM & Sanjappa M. (Plant wealth of northeast India with reference toethnobotany). Indian J Tradit Knowl, 2009; 8: 96-103.

- 9. Survase SA & Raut SD. (Ethnobotanical study of some tree medicinal plants in Marathwada, Maharashtra). J Ecobiotechnol, 2011; 3: 17-21.
- 10. Shah GL. Flora of Gujarat state Part I and II, India; Sardar Patel University, Vallabh Vidhyanagar: 1978.
- 11. Houghton PJ, Raman A. (Laboratory Handbook for the Fractionation of Natural Extracts). Int J Pharm Pharm Sci, 1998; 9(8): 20-24.
- 12. Perilla MJ, Ajello G, Bopp C, Elliot J et al. (Manual for the Laboratory identification and antimicrobial susceptibility testing of bacterial pathogens of Public Health Importance in the Developing World), World Health Organization, 2003; 6.
- 13. Perez C, Pauli M. & Bazerque P. (An antibiotic assay by agar well diffusion method). Acta Baol Med Exp, 1990; 5: 113-115.
- 14. Bonjar GHS, Aghighi S & Nik K. (Antibacterial and antifungal survey in plants used in Indigenous Herbal Medicine of South east regions of Iran). J Bio Sci, 2004; 4(3): 405-412.
- 15. Mujeeb F, Bajpai P & Neelam P. (Phytochemical evaluation, Antimicrobial activity, and Determination of Bioactive components from leaves of Aegle marmelos). BioMed Res Int, 2014.
- 16. Aziz S, Saha K, Sultana N et al. (Phytochemical and elemental screening on leaves and flowers of Catharanthus roseus: An important medicinal plant on Bangladesh). Int J ChemSci, 2014; 12(4): 1328-1336.
- 17. Kabesh K, Senthikumar P, Ragunathan R & Kumar RR. (Phytochenical analysis of Catharanthus roseus plant extract and its antimicrobial activity). Int J Pure App Biosci, 2015; 3(2): 162-172.
- 18. Muhammad KA, Mahood A, Kashif B & Naila A. (Antifungal, Antioxidant and phytochemical screening of *Melia azedarach* extracts by using different solvents). J PharmRes Int, 2017; 20(1): 1-12.
- 19. Farook MA, Mohamed HSM, Mohamed NPMT et al. (Phytochemical screening antibacterial & antioxidant activity of *Melia azedarach*). Int J Res Anal Rev, 2019; 6(2): 248-255.
- 20. Ganjewala D, Tomar N & Gupta AK. (Phytochemical composition and antioxidant properties of methanol extract of leaves and fruits of Neolamarckia cadamba (Roxb)). J Bio Act Pro Nat, 2013; 3(4): 232-240.
- 21. Kumar D, Kumar S, Sahu M, Kumar A. (Phytochemical screening and antioxidant activity of Neolamarckia cadamba and Cymbopogon citrates from drug district of

- Chattisgarh, India). Saudi J Biomed Res, 2020; 5(12): 343-348.
- 22. Arunkumar K & Chandrashekar KR. (Phytochemical evaluation and in vitro antimicrobial and antioxidant studies of leaf and stem bark extracts of *Polyalthia fragrans* (Dalz) Bedd.- An endemic species of western ghats). Int J Pharm Pharm Sci, 2017; 9(8): 20-24.
- 23. Mudhafar M, Zainol I, Desa S & Jaafar CAN. (Mini review of phytochemistry for *Polyalthia longifolia*). Eurasian J Anal Chem, 2019; 14(2): 119-147.
- 24. Arora N, Saxena V, Shandilya K, Bhardwaj P. (Extraction and preliminary phytochemical screening of Tamarindus indica L. leaves). Int J Adv Res Sci & Eng, 2017; 6(5): 640-644.
- 25. Abdallah MS, Muhammad A. (Antibacterial activity of leaves and fruits extract of Tamarindus indica against clinical isolates of Escherichia coli and Shigella at Potiskum Yobe state). J Anal & Pharm Res, 2018; 7(5): 606-609.
- 26. Poonkonthai M & Saravanan M. (Antibacterial activity of Aegle marmelos against leaf, bark and fruit extracts), Anc Sci Life, 2008; 27(3): 15-18.
- 27. Balaabirami S & Patharajan S. (In vitro antimicrobial and antifungal activity of Catharanthus roseus leaves extract against important pathogenic organisms). Int J Pharm Pharm Sci, 2012; 4(3): 487-490.
- 28. DonPaul AM, Weerakoon SR, & Somarathe S. (Antimicrobial and antioxidant properties of leaf, twig and calli extracts of Neolamarckia cadamba (Roxb.) Bosser in Sri Lanka). Res J Med Plant, 2016; 10(4): 314-319.
- 29. Vyas P, Suthar A, & Joshi D. (Antibacterial activity of extracts of leaves of Nerium indicum in combination with antibiotics). Int Lett Nat Sci, 2015; 46: 41-45.
- 30. Chanda S, & Nair R. (Antimicrobial activity of *Polyalthia longifolia* (Sonn.) Thw.var. Pendula leaf extracts against 91 clinically important pathogenic microbial strains). Am J Chin Med, 2010; 1: 31-38.
- 31. Ghosh G, Subudhi BB, Badajena LD et al. (Antibacterial activity of Polyalthia longifolia var. angustifolia stem bark extract). Int J of Pharm Res, 2011; 3(1): 256-260.
- 32. Dutta A, Lal N, Naaz M, Ghosh A & Verma R. (Ethnological and Ethnomedicinal importance of Aegle marmelos(L.) Corr (Bael) among indigenous people of India). American J Ethno, 2014; 1(5): 290-312.
- 33. Pandey A & Negi P. (Traditional uses, phytochemistry and pharmacological properties of Neolamarckia cadamba: A review). J Ethnopharmacol, 2016; 2(181): 118-135.
- 34. Subramanion LJ, Choong YS, Saravanan D et al. (*Polyalthia longifolia* Sonn: An Ancient

remedy to explore for novel therapeutic agents). Res J Pharm Chem Sci, 2013; 4(1): 714-730.

35. Bhadoriya SS, Ganeshpurkar A, Narwaria J & Jain AP. (*Tamarindus indica*: Extent of explored potential). Pharmacogn Rev, 2011; 5(9): 73-81.