

PURIFICATION AND IMPURITY CONTROL IN APIS

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ABSTRACT

A majority spectroscopic and chromatographic method can be performed alone or by the conjunction with the help of method to determine the contaminants. Impurities can be found and characterised using a variety of techniques like atomic absorption spectroscopy (AAS), HPLC, HPTLC. Traditional HPLC in particular form of liquid chromatography, has been thoughtfully utilised throughout the field of impurity profiling. Opportunities for continuous separation based on polymer membrane wettability are created by the ongoing processing of pharmaceuticals. In-line liquid-liquid extraction is made possible by the dual utilization of hydrophobic and hydrophilic membranes in the synthesis of four crucial APIs. Phase separation of aqueous–organic

reaction streams depends on a secondary membrane with opposing wetting properties. Many purification techniques can be used to purify pharmaceutical drugs at all scales, addressing some of the more difficult problems associated with drug purification. Pharmaceutical medications method such as single-use manufacture, chromatography, sterile sampling, ultrafiltration, process development, and sterile filtration.

KEYWORDS: In-line liquid-liquid extraction is made possible by the dual utilization of hydrophobic and hydrophilic membranes in the synthesis of four crucial APIs.

INTRODUCTION

This research aims to explore the various purification techniques and impurity control strategies employed in the manufacturing of APIs. By elucidating the principles and methodologies underlying purification processes, this study seeks to enhance understanding of how impurities are identified, quantified, and controlled throughout the manufacturing process. Furthermore, the regulatory landscape governing impurity limits and quality

standards for APIs will be examined, emphasizing the need for compliance with regulatory guidelines to ensure product safety and efficacy.^[1]

The purification of synthetic API's and semi-synthetic API's can be done by various methods are as follows.

1. Crystallization: That involves the precipitation of the API from a solvent by cooling or evaporation, resulting in the formation of pure crystals which can then be separated from impurities.
2. Sublimation: This technique involves the conversion of a solid directly into a vapour without passing through a liquid phase. It's particularly useful for purifying volatile substances.
3. Distillation: It's especially effective for purifying liquids and volatile compounds. Chromatography: This encompasses various techniques such as HPLC, HPTLC, GC component have been removed chromatography according to how well they bind to mobile & stationary phase.^[2]
4. Ultrafiltration: Involves passing a solution through a semipermeable membrane to separate particles based on size. It's commonly used for removing macromolecules and particulate impurities from solutions.
5. Nano filtration: This technique employs membranes with nanoscale pores to separate substances based on size and molecular weight. It's particularly useful for purifying water and removing small organic molecules and ions.^[3]

Purification processes aim to remove these impurities to ensure the final product meets regulatory standards for safety and efficacy. Control measures include stringent process optimization, analytical testing, and adherence to Good Manufacturing Practices (GMP)^[4]

The excipient and the active pharmaceutical ingredient (API) are the two basic parts of all pharmacological medications. The excipients are Chemically inert materials that transport the active pharmaceutical ingredient (API) to the desired location within an individual's body; the API itself possesses the medicinal properties.^[5]

Classification of purification

Purification control in API (Active Pharmaceutical Ingredient) manufacturing typically falls under critical quality attributes (CQAs) and is classified based on various parameters such as

1. Physical Purification

- Crystallization: Utilized to isolate and purify API crystals from the reaction mixture.
- Filtration: Removes solid impurities or particulate matter from the API solution or suspension.
- Distillation: Separates components based on differences in boiling points, commonly used to purify solvents or remove volatile impurities.

2. Chemical Purification

- Chemical Reactions: Further chemical transformations to remove impurities or convert them into more easily separable forms.
- Complexation: Formation of complexes with specific ligands to selectively remove impurities.
- Precipitation: Addition of reagents to induce the precipitation of impurities, followed by filtration or centrifugation.^[6,7]

3. Chromatographic Purification

- A. Column Chromatography: Separates components on the basis of mobile&stationary phase
- B. High-Performance liq Chromatography (HPLC): Offers higher resolution and faster separation, commonly used for the purification of APIs with complex impurity profiles.
- C. Thin layer chromatography Used for preliminary separate out and observe the impurities^[8]

4. Biological Purification

- A. Biological Assays: Utilized to assess the biological activity and purity of APIs, particularly for biologically derived substances such as peptides or proteins.
- B. Filtration through Biological Filters: Removal of microbial contaminants through filtration using specialized biological filters.

5. Process Optimization

- A. Parameter Optimization: It include temperature, pH, flow rate, along with pressure to enhance purification efficiency and yield.
- B. Scale-Up Considerations: Ensuring scalability of purification processes from laboratory-scale to industrial-scale production while maintaining product quality and purity^[9]

Classification of Impurities

Organic impurities

In all APIs, the most prevalent kind of contaminants are organic ones. They are created while the API is being synthesized. The associated impurities in the synthesis process can be obtained from material, product, catalyst.^[10]

Inorganic impurities

Inorganic impurities are typically introduced during the drug substance's manufacturing or exist as impurities in the excipients. The majority of inorganic contaminants are recognized and classified as follows

Heavy metal: These comprise substances that can be harmful to people even at extremely low concentrations, such as vanadium, antimony, bismuth etc.^[11]

Inorganic reagents and catalysts: These are substances that are utilized in the API's manufacturing process and could include contaminants. Sulphuric acid, platinum, and sodium hydroxide are a few examples.^[12]

Residual solvent: The airborne produce during the producing excipients, drug product preparation are known as residual solvents. They are dangerous for people's health. Based on their degree of toxicity, residual solvents are categorized by ICH into the following three groups.^[13]

Class 1 solvent: These solvents are recognized as the most hazardous and are thought to be a serious threat for people wellness. C₆H₆, CCl₄, C₂H₄Cl₂, and are a few examples.

Class II solvent: These solvents have a moderate level of toxicity and could be harmful to people's health if exposure is not well managed. Methylene chloride, isopropanol, and ethanol are a few examples.^[14]

Method of isolation and identification of API impurities

For governments and the pharmaceutical industry, recognising and determining control for contaminants in active API was a crucial undertaking.

To standardise technical stipulation for pharmaceutical record in the US, Japan the International meeting on Harmonisation was founded in the since 1990. It is advised to

observed & characterise some contaminants 0.10% increase level in accordance ICH criteria. It is the most significant analytical tasks in the developed is the detection of contaminants in APIs.^[15]

High Performance Liquid Chromatography. HPLC also separates & quantifies impurities forms to the stationary and mobile phase. HPLC are highly sensitive and can detect impurities at low concentrations.

Gas Chromatography: They are used for evaluating volatile contaminants found in APIs. It divides substances into mobile gas phases and stationary phases according on how they partition. Remaining solvents and volatile organic contaminants are frequently evaluated by gas chromatography.^[16]

Mass spectrometry-liquid chromatography: Liquid chromatography's separation powers are combined with mass spectrometry's detection and identification powers to create LC-MS. It offers mass spectral information as well as chromatographic separation and is utilized for the identification and measurement of contaminants in APIs.^[17]

Nuclear Magnetic Resonance Spectroscopy: NMR spectroscopy is used to determine the structural makeup of contaminants in active pharmaceutical ingredients (APIs). It enables the detection and impurities by giving information about the molecular structure and connectivity of molecules.

Ultra violet Visible Spectroscopy: The quantitative examination of contaminants in APIs that absorb light in the UV-visible region is done using spectroscopy. It may offer quick and affordable impurity analysis and is frequently used for chromophore impurity analysis.

Thin Layer Chromatography: This is a straightforward and reasonably priced chromatographic method for testing contaminants in APIs qualitatively.

These is often used for preliminary screening of impurities before more sophisticated analytical methods are employed.^[18]

Titration Methods: Various titration methods, such as acid-base titrations, are used for the quantification of impurities in APIs. These methods rely on the reaction between the impurity and a titrant, allowing for the determination of impurity concentrations.^[19]

Method of purification for API

To produce high-quality medicines, reaction by-products like metals and color bodies must be eliminated from processes that synthesize the API through a sequence of chemical reaction stages. The procedures that are most frequently recommended for getting rid of residual metal catalysts include distillation, crystallization, and precipitation. Precipitation and crystallization techniques result in solid material that may be physically removed by selecting a filter step, whereas distillation processes collect the pure API and leave behind the non-volatile components in the residue. Moreover, chromatography and activated carbon powder treatments employ charge and adsorptive methods to eliminate contaminants.^[20]

Reaction by-products, such metals and color bodies, must be removed from processes that synthesize the API through a sequence of chemical reaction steps in order to generate high-quality medications. The best methods for getting rid of residual metal catalysts include distillation, crystallization, and precipitation. Precipitation and crystallization techniques result in solid material that may be physically removed by selecting a filter step, whereas distillation processes collect the pure API and leave behind the non-volatile components in the residue. Moreover, chromatography and activated carbon powder treatments use charge and adsorptive methods to eliminate contaminant.^[21]

Three main areas of focus are downstream processing, host cell impurity removal during clarity, and API bioprocessing. The requirement for expensive chromatography columns should decrease when charged membranes are used as encapsulated capsules.

The utilization of single-use systems in bioprocessing process stages is usually not hindered by chemical compatibility issues, as seen in synthetic API procedures. However, alkali-resistant capsules are required in the manufacturing of bioprocessing if caustic solutions are used for confirmed dehydrogenation procedures.

Bioprocessing companies are achieving higher titres for their target API proteins, which are usually fragments of monoclonal antibodies. This has the consequence of enabling the production of the required amounts of product for clinical trials to be conducted using smaller bioreactors with less demanding facility design constraints. Because of these reduced throughput requirements, single-use technology for the bioreactor and purification process has become more popular.^[22]

Crystallization: Crystallization is a popular technique for API purification. In order to create pure crystals, it entails dissolving the crude API in an appropriate solvent and letting it gently cool or evaporate. Impurities are often excluded from the crystal lattice, resulting in purified API crystals.

Chromatography: APIs are separated and purified from contaminants using techniques such as including flash chromatography and High-Performance Liquid Chromatography (HPLC). In contrast to flash chromatography, which uses a column packed with a stationary phase and a solvent as the mobile phase, HPLC separates substances based on their differential association with a stationary phase and a mobile phase.^[23]

Distillation: Distillation is utilized to purify APIs by separating them from impurities based on differences in boiling points. This method is particularly useful for purifying volatile APIs or removing volatile impurities.

Filtration: Filtration methods include filtration by vacuum filtration, gravity, and membrane analyzing are used to extract particle matter or solid contaminants from liquid API suspensions or solutions.

Extraction: Liquid-liquid extraction is utilized to purify APIs by transferring them from one solvent phase to another. This method exploits differences in solubility to selectively extract APIs from impurities present in the solution.

Precipitation: Precipitation involves the addition of a precipitating agent to induce the formation of insoluble impurities, which can then be separated from the API solution by filtration or centrifugation.

Recrystallization: Recrystallization is a purification technique where a saturated solution of the API in a suitable solvent is allowed to cool slowly, resulting in the formation of pure crystals. This method is often used to remove impurities that are less soluble than the API in the chosen solvent.^[24]

Sublimation: Processes known as sublimation and desublimation are used on materials that have a tendency to break down or polymerize when heated above their melting point. Through the addition of thermal energy, a product can be moved straight from its solid state into its gaseous state without going through a liquid phase. We refer to this reversing process

as de-sublimation. Purification of active pharmaceutical ingredients (APIs) or pharmacological compounds is the most popular use of sublimation and desublimation. Since volatile APIs, complex APIs, and medicinal substances sublime during heating while nonvolatile impurities do not, sublimation is typically employed to separate volatile organic compounds from nonvolatile impurities.^[25]

Regulatory guidelines for impurities

Q1 RECOMMENDATION IS FOR STABILITY

Q1A (R2): Stability Examination of Novel Medicinal Ingredients and Items

Q1B: Examining Photostability of Novel Drug Substances and Items

Q1C: Testing for Stability of Novel Dosage Forms:

Q1D: Matrixing and Bracketing Designs for New Drug Substance and Product Stability Testing

Q1E: Assessment of Stability Information

Q1F: Stability Data Package for Climatic Zones III and IV Registration Applications.^[26,27]

FOR ANALYTICAL VALIDATION, THE Q2 GUIDELINE

Q2 (R1): Analytical Procedure Validation (Text and Methodology)

IMPURITIES ARE THE GOALS OF Q3.

Q3A (R2): Contaminants in Novel Medicinal Ingredients

Q3B (R2): Contaminants in Novel Pharmaceuticals

Q3C (R4): Impurities: Residual Solvent Guidelines.^[28,29]

Extraction method for impurities control in API

One common extraction method used for impurity control in Active Pharmaceutical Ingredients is liquid-liquid extraction (LLE). Here's how it works

1. Principle: Liquid-liquid extraction exploits differences in solubility between the API and impurities in two immiscible liquid phases. By selectively partitioning the API and impurities between these phases, impurities can be effectively separated from the API.



2. Selection of Solvents: Choose two immiscible solvents, typically one polar and one non-polar, that have different affinities for the API and impurities. Common solvent pairs include water and organic solvents like ether, chloroform, or dichloromethane.



3. Extraction Process: Mix the API solution containing impurities with the selected solvent in a separating funnel. Shake or agitate the mixture to facilitate the transfer of the API and impurities between the two solvent phases. Allow the phases to separate based on their densities, forming distinct layers. Collect the organic (non-polar) phase containing the impurities, leaving behind the aqueous (polar) phase containing the purified API. Optionally, repeat the extraction process to further purify the API or remove additional impurities.



4. Washing and Drying: After extraction, wash the organic phase with a suitable solvent to remove any remaining traces of impurities or water. Then, dry the organic phase using an appropriate drying agent to remove any residual moisture



5. Concentration and Recovery: Concentrate the purified API by evaporating the solvent under reduced pressure or using other suitable techniques. Recover the API in its desired form, such as a solid or liquid, ready for further processing or analysis.



6. Validation and Control: Validate the extraction method to ensure its effectiveness in removing impurities and preserving the quality of the API. Monitor process parameters such as solvent ratio, extraction time, and agitation speed to maintain consistency and control impurity levels^[30]

Liquid-solid extraction: a common method used for impurity control in API research. Here's how it works

1. Principle: Liquid-solid extraction involves the transfer of target compounds (the API) and impurities from a liquid phase to a solid phase. The solid phase acts as an adsorbent, selectively retaining the target compounds while allowing impurities to remain in the liquid phase.



2. Selection of Solid Adsorbent: Choose a solid adsorbent material that exhibits affinity for the API while excluding impurities. Activated carbon, silica gel, alumina, and different resins are a few common adsorbents. Preparations of Sample: Prepare the sample containing the API and impurities by dissolving it in a suitable solvent. The sample is then mixed with the solid adsorbent material.



3. Adsorption: Allow the mixture of sample and solid adsorbent to equilibrate, facilitating the adsorption of the API onto the surface of the solid phase. Impurities remain in the liquid phase.



4. Separation Use centrifugation or filtration to eliminate the solid adsorbent material in the liquid phase. The solid phase containing the adsorbed API is retained, while the liquid phase containing impurities is discarded.



5. Washing: Wash the solid adsorbent material with one or more solvents to remove any remaining impurities or contaminants that may be adsorbed non-specifically.^[31]



6. Elution: Elute the API from the solid adsorbent material using a suitable solvent or solvent mixture that disrupts the interactions between the API and the adsorbent. The eluted API is collected in a clean vessel.



7. Concentration and Recovery: Concentrate the eluted API by evaporating the solvent under reduced pressure or using other suitable techniques. Recover the purified API in its desired form, such as a solid or liquid, ready for further processing or analysis.



8. Validation and Control: Validate the liquid-solid extraction method to ensure its effectiveness in purifying the API and removing impurities. Optimize parameters such as solvent composition, adsorbent material, adsorption time, washing conditions, and elution protocol to achieve the desired purification level. Monitor the process to maintain consistency and control impurity levels.^[32]

Soxhlet extraction method is commonly used for impurity control in API research, especially when dealing with solid samples. Here's how it works

1. Principle: The Soxhlet extraction method involves continuous extraction and refluxing of a solvent through a solid sample to remove target compounds (the API) and impurities. It operates on the principle of solubility and differential partitioning of compounds between the sample and the solvent.



2. Setup: The setup consists of a Soxhlet extractor, a round-bottom flask containing the solvent, a condenser, and a collection flask. The solid sample is placed in a porous thimble inside the Soxhlet extractor.



3. Heating and Refluxing: The solvent in the round-bottom flask is heated to its boiling point, causing it to vaporize and rise through the Soxhlet extractor. As the solvent vapor contacts the solid sample, it dissolves the target compounds and impurities. The dissolved compounds are

carried upward with the solvent vapor.



4. Condensation and Collection: As the solvent vapor reaches the condenser, it condenses back into liquid form due to cooling. The condensed solvent then drips back into the Soxhlet extractor, continuously rinsing the solid sample



5. Extraction Cycle: In the Soxhlet extractor, the solvent progressively builds up until it reaches a certain level. At this point, the solvent and dissolved compounds are siphoned off into the round-bottom flask by gravity, completing one extraction cycle. The process repeats automatically until the desired level of extraction is achieved.



6. Purification and Concentration: The collected solvent containing the dissolved API and impurities is then subjected to further purification steps, such as filtration, evaporation, or chromatography, to separate and concentrate the API from impurities.



7. Validation and Control: Validate the Soxhlet extraction method to ensure its effectiveness in purifying the API and removing impurities. To reach the appropriate degree of purification, adjust variables including sample-to-solvent ratio, temperature, extraction duration, and solvent choice. Monitor the process to maintain consistency and control impurity levels^[33]

Extraction method for purification control in API

Membrane-based separation

This technique are commonly used for purification control in Active Pharmaceutical Ingredients (APIs). Here are some of the main membrane-based separation methods.

1. Ultrafiltration (UF)

Principle: Ultrafiltration uses semi-permeable membranes with pore sizes typically ranging from 1 to 100 nm to separate components based on size.

Application: UF is often used to remove macromolecules, colloids, and particulate matter from API solutions. It can also be employed for concentrating and desalting solutions.

2. Microfiltration (MF)

Principle: Microfiltration utilizes membranes with larger pore sizes compared to UF, typically ranging from 0.1 to 10 mm. It separates particles based on size.

Application: MF is commonly used for the removal of bacteria, yeast, and other microorganisms from API solutions. It can also be used for clarification and pre-filtration.

3. Nanofiltration (NF)

Principle: Nanofiltration membranes have smaller pore sizes than UF but larger than reverse osmosis membranes, typically ranging from 1 to 10 nm. NF separates components based on size and charge.

Application: NF is employed for the removal of small organic molecules, ions, and certain contaminants from API solutions. It can be used for purification and concentration^[34]

4. Reverse Osmosis (RO)

Principle: Reverse osmosis utilizes membranes with very small pore sizes, typically less than 1 nm to separate solutes from solvents based on their size and charge.

Application: RO is used for the removal of ions, salts, and other dissolved impurities from API solutions. It is commonly employed for water purification and concentration^[35]

5. Membrane Chromatography: Principle: Membrane chromatography employs membranes functionalized with ligands or affinity groups that selectively interact with target molecules based on specific binding interactions.

Application: Proteins, peptides, and different biomolecules in API solutions can be purified using membrane chromatography. It offers advantages such as high binding capacity, rapid kinetics, and scalability^[36]

Solvent of extraction

1. Principle: Liquid-liquid extraction exploits differences in solubility between the API and impurities in two immiscible liquid phases. By selectively partitioning the API and impurities between these phases, impurities can be effectively separated from the API.
2. Selection of Solvent Systems: Choose two immiscible solvents, typically one polar and one non-polar, that have different affinities for the API and impurities. Common solvent pairs include water and organic solvents like ether, chloroform, or dichloromethane.
3. Extraction Process: Mix the API solution containing impurities with the selected solvents in a separating funnel or extraction vessel. Shake or agitate the mixture to facilitate the transfer of the API and impurities between the two solvent phases. Allow the phases to

separate based on their densities, forming distinct layers. Collect the organic (non-polar) phase containing the impurities, leaving behind the aqueous (polar) phase containing the purified API. Optionally, repeat the extraction process to further purify the API or remove additional impurities.^[37]

4. **Washing and Drying:** After extraction, wash the organic phase with a suitable solvent to remove any remaining traces of impurities or water. Then, dry the organic phase using an appropriate drying agent to remove any residual moisture.
5. **Concentration and Recovery:** Concentrate the purified API by evaporating the solvent under reduced pressure or using other suitable techniques. Recover the API in its desired form, such as a solid or liquid, ready for further processing or analysis.
6. **Validation and Control:** Validate the extraction method to ensure its effectiveness in removing impurities and preserving the quality of the API. Monitor process parameters such as solvent ratio, extraction time, and agitation speed to maintain consistency and control impurity levels.^[38]

Chromatography is a widely used method for purification control in API research. Here's how chromatography works for purification control.

1. **Principle:** Chromatography separates components in a mixture based on their differential interactions with a stationary phase and a mobile phase. The sample mixture is passed through a chromatographic column, where the components are selectively retained and eluted at different rates.^[39]
2. **Selection of Chromatographic Technique**
 - a) **Flash Chromatography:** Used for rapid purification of APIs on a medium to large scale. It employs a column packed with a stationary phase (e.g., silica gel) and a mobile phase (solvent or solvent mixture).
 - b) **Preparative High-Performance Liquid Chromatography (Prep-HPLC):** Suitable for high-resolution purification of APIs. It utilizes a column packed with a stationary phase (e.g., silica-based material) and a high-pressure pump to deliver the mobile phase.
 - c) **Thin-Layer Chromatography (TLC):** used for small-scale purification and rapid qualitative analysis. The sample combination is applied to a thin layer of stationary phase on a plate, and a solvent system is used to develop it.
3. **Sample Loading:** The sample mixture containing the API and impurities is loaded onto the chromatographic column or plate.

4. Separation: As the mobile phase is passed through the chromatographic column or over the stationary phase, the components in the sample mixture interact differently with the stationary phase based on factors such as polarity, size, and charge. This results in differential retention and separation of the components.^[40]
5. Elution: The components are eluted from the chromatographic column or plate by varying the composition or gradient of the mobile phase. The API and impurities are eluted at different times or under different conditions, allowing for their separation.
6. Collection and Fractionation: Fractions containing the purified API and impurities are collected separately as they elute from the chromatographic system.
7. Analysis and Validation: Analyse the collected fractions using analytical techniques such as spectroscopy, mass spectrometry, or chromatography to assess purity and identify the API peak. Validate the chromatographic method to ensure its effectiveness in purifying the API and removing impurities.^[41]

CONCLUSION

This article provides important information regarding Impurities and purification classification, their method, extraction methods and their regulatory guidelines. Impurities and purification control are critical aspects of API research to ensure the safety, efficacy, and quality of pharmaceutical products. Through various extraction, chromatographic, and membrane-based separation techniques, researchers can effectively remove impurities and purify APIs to meet regulatory standards and industry requirements. The type of the contaminants, the API's physicochemical characteristics, and the required purity level are some of the variables that must be taken into consideration while choosing the best purification techniques. Validation of purification methods is essential to ensure their reliability and reproducibility. Additionally, continuous monitoring and optimization of purification processes are necessary to maintain consistency and control impurity levels throughout API research and development. By implementing robust impurity control and purification strategies, researchers can produce high-quality APIs with minimal impurities ultimately contributing.

REFERENCE

1. Joshua ofoeda, Richard Boateng, John Effah Application programming interface (API) Reasearch A Review of the past to inform the future .international journal of enterprises information system, 15(3): 76-95.

2. Bondigalla Ramachandra Development of impurity profiling methods using modern analytical techniques, Critical Reviews in Analytical Chemistry, DOI: 10.1080/10408347.2016.1169913, 2016.
3. Ludmila Peeva, Joao da Silva Bural, Irina Valtcheva, Andrew G. Livingston. Continuous purification of active pharmaceutical ingredients using multistage organic solvent nanofiltration membrane cascade Chemical Engineering Science, 2014; 116(6): 183-194.
4. A Chanda, N Ramalakshi, CN Nalini, S Mahabubi: Impurity profiling an emerging trend in pharmaceutical: A Review Pharma Tutor, 2015; 3(11): 29-35.
5. Parnali Chatterjee and Mohammed M. Alvi Excipients and Active Pharmaceutical Ingredients D. Bar-Shalom and K. Rose (eds.), Pediatric Formulations: A Roadmap, AAPS Advances 347 in the Pharmaceutical Sciences Series 11, DOI 10.1007/978-1-4899-8011-3-24
6. Kavita Pilaniya, Harish K. Chandrawanshi, Urmila Pilaniya, Pooja Manchandani, 2 Pratishtha Jain, and Nitin Singh Recent trends in the impurity profile of pharmaceuticals J Adv Pharm Technol Res., 2010 Jul-Sep; 1(3): 302-310. doi: 10.4103/0110-5558.72422.
7. Roxana Cristina Popescu, Alina Maria Holban, Nanostructured membranes for the microbiological purification of drinking water in Water Purification, 2017.
8. Wioletta Parys ID Małgorzata Dołowy and Alina Pyka-Pająk Significance of Chromatographic Techniques in Pharmaceutical Analysis Department of Analytical Chemistry, Faculty of Pharmaceutical Sciences in Sosnowiec, Medical University of Silesia in Katowice, Jagiellońska 4, 41-200.
9. Altaf Cheng Process optimization and scale-up in pharmaceutical manufacturing Manuscript No. AJPTI-23-98687.
10. J. Chil. Chem. Journal of the Chilean Chemical Society Soc. vol.62 no.2 Concepción jun, 2017.
11. Prajesh Prajapati and Yadvendra K. Agrawal Analysis and impurity identification in pharmaceuticals From the journal Reviews in Analytical Chemistry.
12. Palve Rohit, Gaikwad Annasaheb, Mandale Komal, Pawar Reshma A Review of Impurity Profile in Pharmaceutical Substance Human Journals Review Article August, 2018; 13: 1.
13. ICH Topic Q 6 A, Specifications: Test Procedures and Acceptance Criteria for New Drug Substances and New Drug Products: Chemical Substances.
14. S. V. Saibaba*, M. Sathish Kumar and B. Ramu Pharmaceutical Impurities And Their Characterization: A Review ejpmr, 2016; 3(5): 190-196.

15. Separation techniques: Chromatography Ozlem Coskun Department of Biophysics, Canakkale Onsekiz Mart University, Canakkale, Turkey Correspondence: Dr. Ozlem COSKUN. Canakkale Onsekiz Mart Universitesi Tip Fakultesi, Terzioglu Yerleskesi, Dekanlik Binasi, Biyofizik Anabilim Dalı, Canakkale, Turkey.
16. Qiu, F.; Norwood, D. L. Identification of pharmaceutical impurities. *Journal of liquid chromatography & related technologies*, 2007; 30(5-7): 877-935.
17. Kavita Pilaniya, Harish K. Chandrawanshi, Urmila Pilaniya, Pooja Manchandani, Pratishtha Jain, and Nitin Singh Recent trends in the impurity profile of pharmaceuticals *J Adv Pharm Technol Res*, 2010 Jul-Sep; 1(3): 302–310. doi: 10.4103/0110-5558.72422
18. Neeraj Kumar, Subba Rao Devineni, Prasad Reddy Gajjala, Shailendra Kumar Dubey, and Pramod Kumar Synthesis, isolation, identification and characterization of new process-related impurity in isoproterenol hydrochloride by HPLC, LC/ESI-MS and NMR *J Pharm Anal*, 2017 Dec; 7(6): 394–400. Published online 2017 May 10. doi: 10.1016/j.jpha.2017.05.002
19. Prajesh Prajapati, Yadvendra K Agrawal *May Reviews in Analytical Chemistry*, 2014; 33(2): 123-133. Analysis and impurity identification in pharmaceuticals.
20. Vinod Kumar, Vasudha Bansal, Aravind Madhavan, Manoj Kumar, Raveendran Sindhu, Mukesh Kumar Awasthi, Parameswaran Binod, and Saurabh Saran Active pharmaceutical ingredient (API) chemicals: a critical review of current biotechnological approaches *Bioengineered*, 2022; 13(2): 4309–4327.
21. Peter Koklitis API Purification pharmaceutical Technology Europe-09-01-2011, 23(9).
22. Michael W. Stocker RCID, Matthew J. Harding ,Valerio Todaro ,Anne Marie Healy ORCID and Steven Ferguson Integrated Purification and Formulation of an Active Pharmaceutical Ingredient via Agitated Bed Crystallization and Fluidized Bed Processing *Pharmaceutics*, 2022; 14(5): 1058.
23. Qing-Wen Zhang Li-Gen Lin & Wen-Cai Ye Techniques for extraction and isolation of natural products: a comprehensive review *Chinese Medicine* 13, Article number, 2018; 20.
24. Ayre A. Varpe, D. Nayak R., & Vasa N. Impurity profiling of pharmaceuticals. *Adv Res Pharm Biol*, 2011; 1(2): 76-90.
25. Bari S.B., Kadam B.R., Jaiswal Y.S. and Shirkhedkar A.A., Impurity profile: significance in active pharmaceutical ingredient. *Eurasian journal of analytical chemistry*, 2007; 2(1): 32-53.

26. Radhika Rajagopalan Review of regulatory guidance on impurities. Separation Science and Technology, 5.
27. Swati Patole, Amit Gosar, Tabrez Shaikh Impurities Characterization in Pharmaceuticals: A Review Human Journals Review Article, July 2019; 15(4).
28. Abhimanyu Thakur, Bishal Mishra, Partha Pratim Mahata Pharmaceutical Impurities: A Review. International Journal of Pharmaceutical Chemistry ISSN: 2249-734X (Online) Journal DOI: 10.7439/ijpc
29. ICH Harmonized Triplicate Guideline: Impurities in New Drug Substances Q3A (R2), ICH Steering Committee, Step 4 of ICH process, 25th Oct. 2006.
30. Prajesh Prajapati and Yadvendra K. Agrawal Analysis and impurity identification in pharmaceuticals. From the journal Reviews in Analytical Chemistry.
31. Skoog, et al. Principles of Instrumental Analysis. 6th ed. Thomson Brooks/Cole, 2007; 349-351.
32. Fiori, J.; Bragieri, M.; Zanotti, M.C.; Liverani, A.; Borzatta, V.; Mancini, F.; Cavrini, V.; Andrisano, V. LC-TMS for the identification of impurities in d-allethrine samples. J. Chromatogr, 2005; 1099: 149.
33. Anita Singh, Sadaf Afreen¹, Dharendra Pratap Singh and Rajeev Kumar. A REVIEW ON PHARMACEUTICAL IMPURITIES AND THEIR IMPORTANCE, 6(10): 1337-1354.
34. Suhas P. Dharupaneedi, Sanna Kotrappanavar Nataraj, Mallikarjuna Nadagouda, Kakarla Raghava Reddy, Shyam S. Shukla, and Tejraj M. Aminabhavi. Membrane-based separation of potential emerging pollutants Sep Purif Technol, 2019 Feb 8; 210: 850–866.
35. Arunima Saxena, Bijay P Tripathi, Mahendra Kumar, Vinod K. Shahi, Membrane-based techniques for the separation and purification of proteins: An overview .Advances in Colloid and Interface Science, 145(1-2): 1-22.
36. Jing Chen, Bing Yu, Youqing Shen Recent development and application of membrane chromatography Chemistry, 2023; 415: 45–65.
37. Yan Xu, Maria Aparecida de Souza, Marcela Zanella Ribeiro-Pontes, Michele Vitolo Liquid-liquid extraction of pharmaceuticals by aqueous two-phase systems Revista Brasileira de Ciências Farmacêuticas, 37(3).
38. Sakthivel Lakshmana Prabu, Suriyaprakash T N K Impurities and its importance in pharmacy, July – August 2010; 3(2): 01237.
39. Prabu, S. L., Suriyaprakash, T. N. K., International Journal of Pharmaceutical Sciences Review and Research, 2010; 3(2): 66- 71.

40. Raymond, S.; Chromatographic detectors design: Function and operation, Chromatographic science series, 1995; 73: 201-204.
41. Narasimha S. Lakka and Chandrasekar Kuppan Principles of Chromatography Method Development Chapter Metrics Overview DOI: 10.5772/intechopen.89501