# WORLD JOURNAL OF PHARMACEUTICAL RESEARCH

SJIF Impact Factor 8.084

Volume 12, Issue 6, 573-594.

**Review Article** 

ISSN 2277-7105

# MICROSPHERES AS DRUG DELIVERY SYSTEM

# Prachi Jagdish Salwatkar\*, Arun M. Mahale, Vrushabh Dipak Boralkar

Department of Industrial Pharmacy, Sudhakarrao Naik Institute of Pharmacy, Pusad Nagpur Road Dist. Yavtamal Pusad, 445204.

Article Received on 01 March 2023,

Revised on 22 March 2023, Accepted on 12 April 2023

DOI: 10.20959/wjpr20236-27897

\*Corresponding Author Prachi Jagdish Salwatkar

Department of Industrial Pharmacy, Sudhakarrao Naik Institute of Pharmacy, Pusad Nagpur Road Dist. Yavtamal Pusad, 445204.

#### **ABSTRACT**

"Oral modified-release multiple-unit dosage forms have been found to be more effective than conventional or immediate release single-unit dosage forms. In order to create the final dosage form, the multiparticulates are typically formulated into microspheres and then filled into hard gelatin capsules. Microspheres have been widely studied not only for their ability to provide prolonged release, but also for their potential to target specific drugs. In the future, microspheres are expected to play a central role in novel drug delivery, particularly in areas such as diseased cell sorting, diagnostics, genetic materials, and targeted drug delivery. The purpose of this review is to examine various aspects of the microparticulates drug delivery system,

including formulation methods, evaluation, and characterization."

**KEYWORDS:** Microspheres, Controlled release, Novel Drug Delivery, Therapeutic Efficacy.

# INTRODUCTION

"Microspheres are small spherical particles with diameters ranging from 10 µm to 1000 µm. They have an important role in improving the bioavailability of conventional drugs and minimizing side effects. One of the main advantages of microspheres as a drug delivery system is the ability to control the release of the drug content. Microencapsulation is a technique used to slow down drug release from dosage forms, reduce adverse effects, and increase patient compliance. This technique involves coating an aqueous insoluble core (the drug) with an aqueous insoluble coat (a polymer) using the emulsion solvent diffusion evaporation technique to create a sustained-release drug delivery system. There are various methods for preparing microspheres, including the emulsification technique with single or double solvent evaporation system, spray-dry technique, or phase separation technique. To produce microspheres, starting materials are dissolved in volatile solvents and then dispersed in another solvent that is not miscible with the previous solvent. Upon complete evaporation of the last solvent, a fine powder called microspheres is produced, which is soluble in water. There are two types of microspheres: microcapsules, where the entrapped substance is distinctly surrounded by a distinct capsule wall, and micrometrics, where the entrapped substance is dispersed throughout the microsphere matrix. Solid biodegradable microspheres that incorporate a drug dispersed or dissolved throughout the particle matrix have the potential for controlled drug release. These microspheres are made up of polymeric, waxy, or other protective materials, such as biodegradable synthetic polymers and modified natural products. [1-3]"

# Materials Used<sup>[4-10]</sup>

"Microspheres are commonly made of polymers, which can be classified into two types. synthetic polymers and natural polymers. Synthetic polymers are further divided into non-biodegradable polymers, such as poly methyl methacrylate (PMMA), acrolein, glycidyl methacrylate, and epoxy polymers, and biodegradable polymers, such as lactides, glycolides, their copolymers, poly alkyl cyano acrylates, and poly anhydrides.

Natural polymers are obtained from different sources, including proteins and carbohydrates. Examples of protein-based natural polymers include albumin, gelatin, and collagen, while examples of carbohydrate-based natural polymers include agarose, carrageenan, chitosan, and starch. Chemically modified carbohydrates, such as poly dextran and poly starch, are also used for microsphere preparation."

# Types of microspheres

# 1. Bioadhesive microspheres

Adhesion is the attaching of a substance to a membrane using the adhesive properties of water soluble polymers. Bio adhesion can be defined as the attachment of a medication delivery device to a mucosal membrane, such as the buccal, ocular, rectal, nasal, etc. These types of microspheres have a longer residence duration at the application site, which results in close contact with the site of absorption and improves therapeutic activity. By attaching the drug to a carrier particle, such as microspheres, nanospheres, liposomes, or nanoparticles, carrier technology provides an intelligent method for drug administration by modulating the

drug's release and absorption. These particulate drug delivery techniques rely heavily on microspheres because of their small size and effective carrier capacity.<sup>[11-13]</sup>

# 2. Magnetic Microspheres<sup>[14]</sup>

"Localized drug delivery systems are crucial for targeted treatment of diseases. Magnetic microspheres, which can be composed of materials such as chitosan and dextran, can be used to deliver smaller amounts of drugs specifically to disease sites in response to a magnetic field.

There are two types of magnetic microspheres.

# I. therapeutic magnetic microspheres

Therapeutic magnetic microspheres are used to deliver chemotherapeutic agents to liver tumors, and can also target proteins and peptides.

# II. diagnostic microspheres.

Diagnostic microspheres, on the other hand, can be used to image liver metastases and distinguish bowel loops from other abdominal structures using nano-sized particles of supramagnetic iron oxides."

# 3. Floating microspheres<sup>[15, 16]</sup>

Because the bulk density of floating kinds is lower than that of gastric fluid, they float unaffected by the rate at which the stomach empties. If the system is floating on stomach content and increases gastric residence and increases plasma concentration fluctuation, the medicine is released slowly at the desired rate. Additionally, it lessens the likelihood of striking and dose dumping. It also results in a sustained therapeutic impact, which lowers the frequency of dose.

# 4. Polymeric microspheres<sup>[17]</sup>

The many kinds of polymeric microspheres can be divided into two categories: synthetic polymeric microspheres and biodegradable polymeric microspheres.

# Biodegradable polymeric microspheres

Starch is an example of a natural polymer that is biodegradable, biocompatible, and bioadhesive. Biodegradable polymers can prolong the time that microspheres remain in contact with mucous membranes due to their high degree of swelling in an aqueous medium, resulting in gel formation. The concentration of polymer can control the rate and extent of

drug release in a sustained manner. However, one challenge of using biodegradable microspheres in clinical settings is that controlling the drug loading efficiency and release pattern can be difficult.

# **Synthetic polymeric microspheres**

In addition to being employed as bulking agents, fillers, embolic particles, drug delivery vehicles, etc., synthetic polymeric microspheres are also frequently used in clinical applications and have proven to be both safe and biocompatible. The main drawback of these microspheres is that they have a propensity to migrate away from the injection site, increasing the risk of embolism and subsequent organ damage.

# 5) Radioactive microspheres

The subset of microspheres that interact radioactively is often handled similarly to nonradioactive microspheres. But in addition to the matrix material that characterises the microsphere and gives it its targeting properties in a certain tissue or organ, the radioactive microsphere always contains one, and occasionally more radio-nuclides. Radioactive microspheres can also deliver high radiation doses to a particular area in small quantities while obstructing the surrounding healthy tissue. [17, 18]

# **Advantages of Microspheres**

- A. Increased surface area leading to increased potency of poorly soluble material.
- B. Improved patient compliance by providing a steady quantity of medication in the body.
- C. Reduced dose and risk.
- D. Polymer packaging protects drugs from enzymatic cleavage and allows for delivery systems.
- E. Higher patient compliance due to shorter duration of dosing.
- F. Enhanced bioavailability and decreased occurrence/severity of harmful effects.
- G. Protection of gastrointestinal tract from opioid irritants.
- H. Transformation of liquid drugs into solid shape, blocking unpleasant taste.
- I. Reliable means of delivering medication to target location with precision and sustained concentration.
- J. Reduction in central reactivity related to external world.
- K. Benefits of degradable microspheres over large polymer implants, including not requiring surgical implantation.

L. Controlled release of drugs and reduction in toxicity/discomfort of repeated injection with degradable microspheres.<sup>[20]</sup>

# LIMITATION<sup>[21]</sup>

The following list of drawbacks was discovered.

- 1. The altered formulation release.
- 2. A range of circumstances, including meals and the speed at which the medication travels through the gut, can affect the controlled release dosage form's release rate.
- 3. Variations in the rate of release between doses.
- 4. Because controlled release formulations often have a larger drug load, any degradation of the dosage form's release properties could potentially be hazardous.
- 5. Crushing or chewing these dosage forms is not advised.

# Method of preparation of microspheres

Table 1: Microsphere property. [22]

S. No	Property	Consideration
1	Size Diameter	Uniformity/distribution
2	Composition	Density, Refractive Index, Hydrophobicity/hydrophilicity Nonspecific binding Autofluorescence
3	Surface Chemistry	Reactive groups Level of functionalization Charge
4	Special Properties	Visible dye/fluorophore Superparamagnetic

# Method of preparation of microspheres

The characteristics of the polymer being used, the drug, factors equivocally determined by many formulations, technological factors, such as the requirement for particle size, and the drug or protein should not be significantly impacted by the process, the reproducibility of the release profile and the method, there should be no stability issue, in relation to the finished product, are the main factors that influence the method selection. the various methods for preparing the microspheres with hydrophilic and hydrophobic polymers as the matrix components.<sup>[23]</sup>

- 1. The ability to incorporate small medication doses.
- 2. Stability of the preparation after synthesis, with a clinically acceptable shelf life.
- 3. Precise control over particle size and dispersibility for injection in aqueous vehicles.
- 4. Reliable release of the reagent with precise control over an extended time period.
- 5. Biocompatibility with controllable biodegradability and chemical alteration response.

# **Emulsion Solvent Evaporation Technique**

This method involves dissolving the drug in a polymer solution that was also previously dissolved in chloroform. The resulting solution is then added to an aqueous phase containing 0.2% sodium PVP, which acts as an emulsifying agent. The mixture is agitated at 500 rpm to form fine droplets of the drug and polymer (eudragit), which then solidify into rigid microspheres through solvent evaporation. These microspheres are collected by filtration, washed with demineralized water, and dried at room temperature for 24 hours.<sup>[24]</sup>

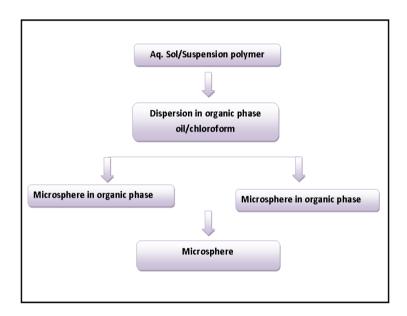


Fig. 1: Single emulsion technique.

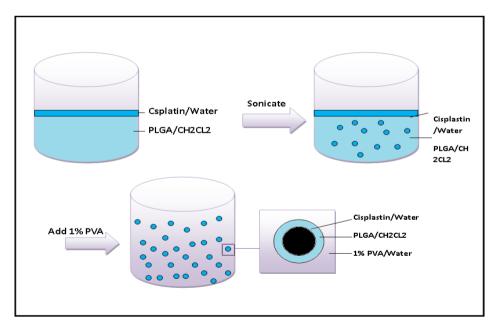


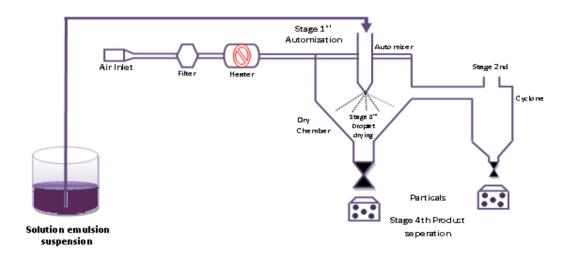
Fig. 2: Double emulsion technique.

## Wax coating and hot melt

One method for encapsulating components involves dissolving or dispersing the product in melted wax. A waxy paste is formed and then blended with cold water to release it. After heating the mixture for at least an hour and stirring it for another hour, the external layer (liquid paraffin) is separated from the microspheres. The microspheres are then immersed in a non-miscible solvent and dried using dry air. Carnauba wax and beeswax are suitable for surface ingredients and can be combined to achieve desired characteristics. [25, 26]

# Spray drying technique

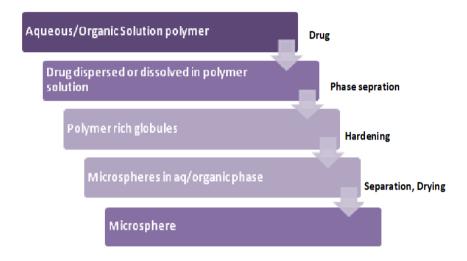
This method involves dissolving the polymer in a volatile organic solvent, such as dichloromethane or acetone, then dispersing the medication (in solid form) in the polymer solution using high-speed homogenization. After that, the dispersion is atomized in the hot air stream, and atomization causes the development of tiny droplets from which the solvent instantly evaporates, resulting in the formation of microspheres with a size range of 1–100 m. With the aid of a cyclone separator, prepared microparticles are separated by hot air, and solvent remnants are eliminated by vacuum drying. [27, 28]



#### Coacervation

The basic coacervation method involves separating a macromolecular fluid into two types of materials that do not mix: a condensed coacervate layer and a clear equilibrium layer. This method is used when there is only one type of macromolecule involved. On the other hand, complex coacervation occurs when two or more macromolecules with opposite charges are present. Basic coacervation can be induced by various factors, such as changes in

temperature, use of non-solvents, or the presence of micro-ions that cause dehydration in macromolecules. These factors promote interactions between polymers and lead to the formation of microspheres with different properties.<sup>[29]</sup>



# **Solvent evaporation**

The process of solvent evaporation has also been widely employed to create PLA and PLGA microspheres that contain a wide range of medications. There are a number of factors that have been found to have a substantial impact on microspheric properties, including drug solubility, internal morphology, solvent type, diffusion rate, temperature, polymer composition, viscosity, and drug loading. Because the effectiveness of the solvent evaporation system relies on the effective entanglement of the active ingredient into the particles, this process is especially effective with medications that are either insoluble or only partially soluble in the liquid medium that makes up the constant phase. [30]

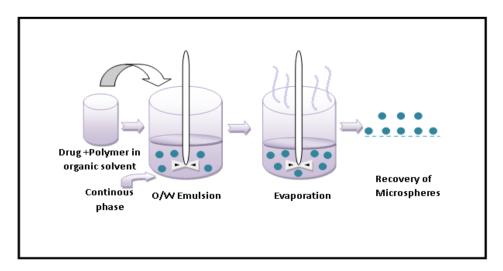


Fig: Solvent evaporation technique.

# **Ionic Gelation Method**

This method was used to create an alginate/chitosan particulate system for the release of nateglinide. A 2% (w/v) aqueous solution of sodium alginate was mixed with various percentages (w/v) of nateglinide. Stirring is continued until the mixture is complete, and then it is dropped gradually into a Ca2+ and chitosan solution in acetic acid. After forming, the microspheres were left in the original solution for 6 and 24 hours to allow for internal gellification before being filtered to separate them. While the medication did not release at an acidic pH, the full release was obtained at 7.4<sup>[31]</sup>

# Quassi Emulsion Solvent Diffusion<sup>[32]</sup>

The literature has described a unique quasi-emulsion solvent diffusion process for creating drug-filled acrylic polymer controlled release microspheres. By adopting a quasi-emulsion solvent diffusion process with an exterior phase made of polyvinyl alcohol and distilled water, microsponges can be produced. To increase flexibility, 20% of the polymer is introduced to the internal phase, which is made up of the medication, ethanol, and polymer. The external phase is added to the internal phase after the internal phase has first been created at 60 oC and is at room temperature. The mixture is continually swirled for two hours following emulsification. To separate the microsponges, the mixture can then be filtered. The product is subsequently cleaned and dried for a day in a vacuum oven at 40°C.

# **Freeze Drying**

In the preparation of protein API microspheres, freeze-drying works well. The process involves sublimation, freezing, primary drying, and secondary drying. The eutectic point of the constituents is taken into consideration during the freezing process. By eliminating water, establishing a glass matrix, decreasing intermolecular interactions by forming hydrogen bonds between the molecules, or by forming dipole-dipole interactions, lyoprotectants or cryoprotectants will stabilise API molecules during the process. Given its high cost, the cycle is advantageous for molecules that can withstand heat. Particles in an aqueous medium can then be reconstituted after freeze-drying solidifies them.<sup>[33]</sup>

#### **Emulsion-Solvent Diffusion Technique**

Ketoprofen floating microperticles were made utilising the emulsion solvent diffusion approach to increase the residence period in the colon. Dropwise additions of the dissolved drug polymer mixture to sodium lauryl sulphate (SLS) solution were made after it had been first dissolved in an ethanol and dichloromethane (1:1) mixture. At room temperature, the

solution was agitated for 1 hour at 150 rpm using a propeller-style agitator. As a result, the produced floating microspheres were cleaned before being dried at room temperature in a dessicator. The microparticles listed below were sieved and gathered. [34]

# **Multiple Emulsion Method**

This method was used to manufacture oral controlled release medication delivery for a variety of medicines. The powdered medication was first dissolved in methyl cellulose solution, then it was emulsified in ethyl cellulose solution in ethyl acetate. Reemulsification of the main emulsion in aqueous media followed. During this phase, discrete microspheres were formed under optimal conditions.<sup>[34]</sup>

# **Physicochemical Evaluation**

#### Characterization

A crucial phenomenon that aids in the development of an effective carrier for the transport of proteins, drugs, or antigens is the characterisation of the microparticulate carrier. Different microstructures can be seen in these microspheres. These microstructures control the carrier's release and stability.<sup>[35]</sup>

#### Particle size and shape

Using a calibrated ocular micrometer, optical microscopy can measure particle size. One hundred microspheres are sized and their average particle size is determined.

D mean =  $\sum n d/\sum n$ 

Where, n = number of microspheres checked; d = Mean size

# Scanning electron microscopy (SEM) study

The analysis of the samples was conducted using scanning electron microscopy (SEM) and energy dispersive X-ray spectroscopy (EDXA) to determine their elemental structure. A back-scattered electron sensor was utilized for image analysis, while a centered electron beam was used to scan the sample in parallel lines. To prepare for SEM characterization, microspheres were coated with conductive metals like platinum or zirconium using a sputter coater. The sample was then scanned using a guided, fine electron beam, and the surface properties were derived from the secondary electrons emitted from the sample surface. [36]

## Flow properties

By calculating the Carr's compressibility index, Hausner ratio, and resting angle of repose, the flow parameters can be studied. To evaluate the bulk and tapped densities, a volumetric cylinder was employed.<sup>[37]</sup>

#### Thermal analysis

Thermal analysis techniques are commonly used to study changes in materials by subjecting them to scheduled variations in temperature for heating and cooling, as well as defined specimen atmospheres and pressures. These techniques can measure various properties such as heat and enthalpy changes, weight loss or gain, Young's modulus, thermal expansion or shrinkage, and gas evolution.<sup>[38]</sup>

#### **Determination of percentage yield**

Calculating the product's measured quantity, the polymers utilised in the microspheres' formulation, and the total number of microspheres produced will yield the percentage yield.<sup>[39]</sup>

# **Density Determination**

The density of microspheres can be determined using a multi-volume pycnometer. First, a precisely weighed sample is placed into a cup and introduced into the pycnometer chamber. Helium is then introduced into the chamber at a constant pressure and allowed to expand, resulting in a decrease in pressure within the chamber. Two consecutive readings of pressure reduction at different initial pressures are taken. By analyzing these readings, the volume and density of the microsphere carrier can be accurately determined.

#### **Isoelectric Point**

The micro electrophoresis is a device that measures the electrophoretic mobility of microspheres and uses that information to calculate the isoelectric point. The time of particle travel over a distance of 1 mm is used to compute the mean velocity at various Ph values ranging from 3 to 10. This information can be used to estimate the particle's electrical mobility. The microspheres' ion-absorbing properties, ionisable behavior, or surface-contained charge can all influence their electrophoretic mobility.

# **Surface Carboxylic Acid Residue**

Radioactive glycine is used to measure the surface carboxylic acid residue. The c14-glycine ethyl ester hydro chloride reacts with the microspheres to produce the radioactive glycine conjugates. The water-soluble condensing compound 1- ethyl-3 (3-dimethyl amino propyl) carbidiimide is used to connect the glycine residue. (EDAC). Then, a liquid scintillation counter is used to determine the conjugate's radioactivity. Thus, it is possible to compare and correlate the carboxylic acid residue. For either hydrophobic or hydrophilic microspheres, as well as any other derivatized kind, the free carboxylic acid residue can be determined.

### **Drug entrapment efficiency**

Crushed microspheres are obtained and weighed in quantity. Afterward, it was stirred into a buffer solution before being dissolved and filtered. Using a calibration curve, the UV spectrophotometer measures the filtrate at a specific wavelength.

# **Swelling index**

It is calculated by measuring how much microspheres swell when placed in a specific solvent. The equilibrium swelling degree of microspheres is assessed by overnight swelling in a measuring cylinder of 5 mg of dried microspheres in 5 ml of buffer solution. It is computed using the provided formula.

#### **Electron Spectroscopy for Chemical Analysis**

The electron spectroscopy for chemical analysis can be used to determine the microspheres' surface chemistry. (ESCA). The ESCA method offers a way to ascertain the surface's atomic make-up. You can assess the surfacial degradation of the biodegradable microspheres using the spectra produced using ECSA.

### Attenuated total reflectance Fourier TransfomInfrared Spectroscopy

The carrier system's polymeric matrix degradation is assessed using FT-IR. The microspheres' surface is examined using alternated total reflectance measurements. (ATR). The sample received many reflections from the IR beam travelling through the ATR cell, yielding IR spectra primarily of surface materials. Depending on the conditions and methods of manufacture, the ATRFTIR provides information about the surface composition of the microspheres.

#### **APPLICATIONS**

# 1. Microspheres in Vaccine Delivery<sup>[40,41]</sup>

Protection against the microorganism or its harmful product is a need for a vaccination. An ideal vaccination must meet the criteria for effectiveness, safety, ease of use, and cost. Safety and minimising negative reactions is a complicated topic.64. The degree of antibody formation and the safety factor are directly correlated with the application method. The shortcomings of conventional vaccines may be overcome by biodegradable delivery technologies for vaccines administered parenterally65. Parenteral (subcutaneous, intramuscular, and intradermal) carriers are appealing because they provide a number of benefits, including.

- 1. Improved antigenicity by adjuvant action
- 2. modifying the release of an antigen.
- 3. Antigen stabilisation.

### 2. Targeting using Microparticulate Carriers

Targeting, or site-specific medication delivery, is a well-established paradigm that is now receiving a lot of attention. The drug's ability to reach and engage specifically with its candidate receptors is essential to its therapeutic effectiveness. The main pharmacological action is mediated by the employment of a carrier system, which allows the drug to exit the pool in a repeatable, effective, and precise manner. When particles are placed in a certain anatomical compartment, either the environment's physical characteristics or the particles' biophysical interactions with the target tissue's cellular composition allow the particles to be retained.

# 3. Microspheres in Gene delivery

Viral vectors, nonionic liposomes, polycation complexes, and microcapsule technologies are all used in genotype medication delivery. Although they are quite effective and have a wide range of cell objectives, viral vectors are helpful for genotype delivery. However, when utilised in vivo, they produce harmful consequences and immunological responses. Nonviral delivery techniques for gene therapy have been considered as a solution to the limitations of viral vectors. Benefits of nonviral delivery systems include ease of preparation, cell and tissue targeting, less immune response, unrestricted plasmid size, and highly reproducible manufacture on a wide scale. For applications involving the transfer of genes, polymer will be employed as a DNA transporter. [42,43]

## 4. Monoclonal Antibodies Mediated Microspheres Targeting

Microspheres that are immune to mAbs are called microspheres. This targeting is a technique for achieving particular site-specific targeting. Monoclonal antibodies are highly specialised substances. Monoclonal antibodies (Mabs) can be used to target microspheres loaded with bioactive compounds to specific regions due to their great selectivity. Mabs can be covalently coupled to the microspheres directly to form an attachment. The antibodies may be coupled to the free aldehyde, amino, or hydroxyl groups on the surface of the microspheres. There are a variety of ways to attach the Mabs to microspheres.

- 1. Non specific adsorption
- 2. Specific adsorption
- 3. Direct coupling
- 4. Coupling via reagents

#### 5. Oral drug delivery

Rabbits have been used to assess the potential of polymer matrix, which typically comprises diazepam as an oral medication delivery. Its results demonstrated that even a film made of a drug-polymer mixture in a 1:0.5 ratio would have been a useful dosage form that is equivalent to existing tablet formulations. The ability of polymers to form films may make it possible to construct film dosage forms as an alternative to medication tablets. When the major amine groups react in both directions, the polymer begins to stand out as a special polymer for applications involving oral medication delivery.<sup>[44]</sup>

# 6. Transdermal drug delivery

Polymer has effective film-forming properties. The membrane thickness and crosslinking of a film both have an effect on the release profile from the devices. Additionally, in-situ preparation of the chitosan-alginate polyelectrolyte structure in beads and microspheres has been done in preparation for prospective uses in packaging, controlled release systems, and surgical instruments. For chemotherapy of inflammatory cytokines for drugs like prednisolone that also showed extended release action boosting treatment efficiency, polymer gel beads are an amazing extremely biocompatible delivery system. It was discovered that the features of the cell wall being used also affected how much medication was released. A local anaesthetic made of chitosan hydrogel and membrane that is known to contain lidocaine hydrochloride is a fantastic all-encompassing method for managing drug release kinetics.<sup>[45]</sup>

### 7. Other applications

For membrane technology created for mass spectrometry, cell biology, and fluorescently coupled immuno-sorbent assay, microspheres are used. Yttrium has the potential to be employed in the routine therapy of hepatocellular carcinoma and even in conjunction with pre-transplant management of HCC. There are other uses for microencapsulation in other business sectors. The most well-known microencapsulated products include carbonless copying paper, photosensitive paper, "scent-strips" (sometimes called "snap-n-burst"), and "scratch-n-sniff" microencapsulated scents. These other items are typically made using a complex of gelatin and acacia. Children's literature, as well as the advancement of nutrition and fragrance advertising for cosmetics, have all used scratch-and-sniff techniques. Additionally, the use of microcapsules in diagnostic procedures is widespread. One such use is the temperature-sensitive microcapsules for temperature-dependent visual cancer diagnosis. Microcapsules containing microorganisms are utilised in the biotech sector to produce recombinant proteins. [46]

#### 8. Surface Modified Microspheres

Different methods have been used to modify carriers' surface characteristics, shield them against phagocytic clearance, and modify their body distribution patterns. The polystyrene, polyester, or poly methyl methacrylate microspheres become more hydrophilic as a result of the poloxamer's adsorption on their surface, which lowers their MPS uptake. PEG derivative-covalently modified protein microspheres exhibit decreased immunogenicity and clearance.

The surface modifications that have received the most research are.

- 1. Antibodies and their fragments
- 2. Proteins
- 3. Poly-, oligo-, and monosaccharides
- 4. Compounds that corrode (EDTA, DTPA or Desferroxamine)
- 5. Artificial soluble polymers In order to target certain organs and prevent quick evacuation from the body, these changes are added to the surface of microspheres.

#### RECENT ADVANCEMENT IN MICROSPHERE

#### 1. Significant uses for chitosan polymer

#### **Effects of reducing cholesterol**

Examples of fibres with high, moderate, and poor bile acid-binding capabilities were given using chitosan and cellulose, respectively. The inclusion of any of these fibres at 75% of the

diet avoided the 2-fold increase in blood cholesterol levels that occurred in a control group of mice fed a high fat/high cholesterol diet for 3 weeks. Treatment with these fibres also helped to lessen the amount of cholesterol that the HFHC diet caused to accumulate in hepatic storage. Similar hypocholesterolaemic effects were seen with all three types of fibre; however, cholestyramine significantly reduced the amount of cholesterol in liver tissue. The processes underpinning cholestyramine's ability to decrease cholesterol included,

- 1) Less dietary cholesterol intake;
- 2) Less effective cholesterol absorption;
- 3) More bile acid and cholesterol excretion in the faeces.

Due to cholestyramine's strong bile acid binding ability, the latter effects can be explained. In contrast, adding chitosan or cellulose to the meal decreased the amount of cholesterol consumed, but had no impact on how much cholesterol was absorbed or excreted in the faeces. The current study offers compelling evidence that the decrease of LDL is mostly caused by satiation and satiety effects. [47]

# 2. Increase Stability of Drug

When a medicine is complexed with chitosan and made into a slurry and dough mass, kneading for 45 minutes, chitosan polymer is employed to boost the stability of the drug. This dough mass is sent through filter number 16 to create granules that are utterly stable under various conditions.

# 3. Orthopaedic Patients

Chitosan is a biopolymer that has advantages for osteointegration of orthopaedic and craniofacial implant devices, including osteoconductive, improved wound healing, and antibacterial qualities. It has been shown to be effective in increasing bone regeneration, wound healing, and tissue growth in tissue repair.

#### 4. Cosmetics industry

The disclosure of cosmetic formulations for the treatment of hair or skin that contain brandnew quaternary chitosan derivatives of the formula. The chitosan derivatives demonstrate hair strengthening and conditioning properties and have a good substantial, particularly to hair keratin. for instance, hair setting lotion and oxidation Skin Cream, Hair Treatment Composition, Hair Coloring Composition, and Hair Toning Composition in Gel form.

#### 5. Dental Medicine

Chitosan has been shown to hasten wound healing, achieve an aesthetically pleasing skin surface, and avoid the production of excessive scar tissue. Chitosan is also used in dental medicine as a tampon after radical therapy for maxillary sinusitis and as a bandage for oral mucosal wounds. It is also being looked into as a potential absorbent membrane for periodontal surgery. Chitosan is promoted as a nutritious meal that can treat or improve a variety of diseases, including diabetes, cancer, hepatitis, and arthritis. It has a wide range of biological activity.

#### 6. Chitosan as Permeation Enhancer

According to certain reports, chitosan can open tight connections in a cell membrane because of its cationic character. This characteristic has prompted several research to examine the potential of chitosan as a permeation enhancer for hydrophilic medicines, such as peptides, which may otherwise have poor oral bioavailability. The phenomenon is pH and concentration dependent because the absorption amplification results from interactions between the cell membrane and positive charges on the polymer. Additionally, a polymer with a higher charge density would have a higher permeability.

#### 7. Chitosan as Mucoadhesive Excipient

Bioadhesivity is frequently employed as a strategy to lengthen a drug's duration in the GI system, hence enhancing the oral bioavailability. Chitosan has a better bioadhesivity than other common polymeric excipients such cellulose, Xantham gum, and starch, according to a comparison with other widely used polymeric excipients.

#### 8. Effect of chitosan

Ratio of citric acid to medication release It has been proven that polymers with the right viscosity and expansion properties can be employed as osmotic agents to release drugs that are not water soluble. Chitosan has an excellent gelation characteristic and is completely biodegradable, toxicologically safe, and inexpensive due to its high molecular weight and linear, unbranched structure. Therefore, it is clear that chitosan has the potential to be exploited as a polymeric osmotic agent in osmotic pumps. The pH of the environment has a big impact on how chitosan hydrates and forms gels. It is insoluble at neutral and alkaline pH levels but soluble in acid. The polymer's amine groups protonate during dissolution, resulting in the formation of a viscous polysaccharide that is soluble. Citric acid was added as a pH-regulating excipient to the formulations in order to lower the microenvironmental pH of the

core to a level where chitosan could form a suitable viscous gelling solution, increasing the osmotic pressure of the core tablets in the process.

#### 9. Chitosan as Permeation Enhancer

According to certain reports, chitosan can open tight connections in a cell membrane because of its cationic character. This characteristic has prompted several research to examine the potential of chitosan as a permeation enhancer for hydrophilic medicines, such as peptides, which may otherwise have poor oral bioavailability. The phenomenon is pH and concentration dependent because the absorption amplification results from interactions between the cell membrane and positive charges on the polymer. Additionally, a polymer with a higher charge density would have a higher permeability.

#### 10. Enhanced Bone Formation by transforming growth factor (TGF-pl)

In order to achieve high bone-forming efficacy, chitosan composite microgranules were created as bone substitutes. In order to create the chitosan microgranules, a mixed solution was dropped into a NaOH/ethanol solution. The chitosan microgranules were soaked in a TGF-pl solution to load them with TGF-pl.

# 11. Direct Compressible Excipients and as Binder

When used as an excipient in direct compression of tablets, chitosan has the good property of causing quick disintigration when added at a concentration of 50%. The amount of moisture absorption is determined by the degree of deacetylation. Corn starch and microcrystalline cellulose were outperformed as a disintigrant by chitosan at concentrations greater than 5%. Chitosan crystalinity, degree of deacetylation, molecular weight, and particle size all had an impact on the effectiveness. With the rank order correlation for binder efficiency, chitosan is discovered to be an excellent tablet binder when compared to other excipients. Chitosan, methyl cellulose, and sodium carboxy methyl cellulose are the order of preference.

#### CONCLUSION

As opposed to other cutting-edge drug delivery systems, it has been observed. In comparison to traditional drug delivery methods, the idea of microsphere drug delivery systems has some benefits, such as regulated and sustained delivery. In addition, microspheres enable medication targeting to a variety of systems, including the ocular, oral, intranasal, and intravenous routes. For medication delivery, microspheres are a preferable option, especially

in disease-related cell sorting and diagnostic procedures effective, targeted, in vivo delivery of a gene. Microspheres will therefore be crucial to the growth of science in the future.

#### REFERENCES

- 1. Freitas S, Merkle HP, Gander B. Microencapsulation by solvent Extraction/Evaporation: reviewing the state of the art of microsphere preparation process technology. J Controlled Release, 2004; 102: 313–32.
- 2. Sahil K, Akanksha M, Premjeet S, Bilandi A, Kapoor B, Microsphere: a review. Int J Res Pharm Chem, 2011; 1: 2231-781.
- 3. Rajput S, Agrawal P, Pathak A, Shriyasataya N, Baghe SS, Baghe RS. A review on microspheres: methods of preparation and evaluation. World J Pharm Pharm Sci, 2012; 1: 422-38.
- 4. P.M. Dandagi, VS. Mastiholimath, M.B. Patil, M.K. Gupta, Biodegradable microparticulate system of captopril. International Journal of Pharmaceutics, 2006; 307: 83-88.
- 5. Chinna Gangadhar B, Shyam Sunder R., Vimal Kumar Varma. M., Sleeva Raju M., Sai Kiran M, Formulation and Evaluation of Indomethacin Microspheres using natural and synthetic polymers as Controlled Release Dosage Forms. International Journal of Drug Discovery, 2010; 2(1): 8-16.
- 6. Rana mazumder, lila K. Nath, Anwarul, Haque, Tarasankar Maity, Prasant K. Choudhary, Bhupendra Shreshta, Formulation and in vitro evaluation of natural polymers based microsphere for colonic drug delivery, International journal of pharmacy and pharmaceutical sciences, 2010; 2(1): 211-219.
- 7. Kavitha K, Chintagunta Pavanveena, Anil Kumar S. N., Tamizh Mani T, Formulation and evaluation of trimetazine hydrochloride loaded gelatin microsphere. International Journal of Pharmacy and Pharmaceutical Sciences, 2010; 2(3): 67-70.
- 8. Lorenzo-Lamosa ML. Design of microencapsulated chitosan microspheres forcolon drug delivery. J. Control. Release, 1998; 52(1-2): 109-118.
- 9. Sudha Mani T and Naveen Kumar K, At preparation and evaluation of ethyl cellulose microspheres of ibuprofen for sustained drug delivery. International Journal of Pharma Research and Development, 2010; 2(8): 120-121.
- 10. Bunty Chanu Irom, K. Kavitha, M. Rupeshkumar1, SD. Jagadeesh Singh, Natural Polymeric Microsphere for Drug Delivery: A Review. International Journal of Pharmaceutical Research And Development, 2012; 4(07): 31-37.

- 11. Meghna KS, Krishna MP, Giridas S, Sreelakshmi C, Vijayakumar B. Microsphere a drug delivery system—a review. Int J Novel Trends Pharm Sci, 2017; 7: 109-18.
- 12. Kumar A, Mahajan S, Bhandari N, Microspheres: a review. World J Pharm Pharm Sci, 2017; 6: 724-40.
- 13. Vikrant KN, Gudsoorkar VR, Hiremath SN, Dolas RT, Kashid VA. Microspheres-a novel drug delivery system: an overview. Int J Pharm Chem Sci, 2012; 1: 113-28.
- 14. Guojun Liu, Husheng Yang, Jiayun Zhou, Preparation of magnetic microsphere from waterin-oil emulsion stabilized by block copolymer dispersant,. Biomacromolecules, 2005; 6: 1280-1288.
- 15. P. Dutta, J.Struti, Ch. Niranajan patra, M.E. Bhaoji rao, Floating Microsphere: Recents Trends in the Development of Gastroretentive Floating Drug Delivery System. International Journal of Pharmaceutical Science and nanotechnology, 2011; 4(1): 1293-1306.
- 16. Y. Kawashima, T. Niwa, H. Takeuchi, T. Hino, Y. Ito, Preparation of multiple unit hollow microspheres (microbal loons) with acrylic resin containing tranilast and their drug release characteristics (in vitro) and floating behavior (in vivo). J. Control. Release, 1991; 16: 279-290.
- 17. DAS, M. K., A. B. AHMED, and D. SAHA. "MICROSPHERE A DRUG DELIVERY SYSTEM-A REVIEW". International Journal of Current Pharmaceutical Research, July 2019; 11(4): 34-41, doi:10.22159/ijcpr.2019v11i4.34941.
- 18. Urs Häfeli, et. al., Review: Radioactive Microspheres for Medical Applicationsm, Cleveland Clinic Foundation, Radiation Oncology Department T28, page 1-29.
- 19. Lachman LA, Liberman HA, Kanig JL. The Theory and Practice of Industrial Pharmacy. Varghese Publishing House, Mumbai, India, 1991; 3rd edition; P-414-415.
- 20. Tarun Virmani, et. al., Pharmaceutical Application of Microspheres: An Approach for The Treatment of Various Diseases, International Journal of Pharmaceutical Sciences and Research, 2017; 7: 3252-3260.
- 21. Vyas SP, Khar RK. Targeted and Controlled drug delivery., 7th Edition; Vallabh Prakashan, New Delhi India, 420-445.
- 22. Dr. Fishers Microsphere Selection Bangs laboratories inc, Tech Notes 201A, 1-4 Available from **URL** http://www.bangslabs.com/sites/default/files/bang s/do cs/pdf/201A.pdf.
- 23. E Veena Rani, et. al., Preparation and Evaluation of Aspirin Loaded Microspheres by Solvent Evaporation Technique, Journal of Medicine and Biology, 2019; 1(1): 27-32.

- 24. Trivedi P., Verma A.M.L., Garud N., Preperation and Charecterization of Acclofenac Microspheres, Asian Journal of pharmaceutics, 2008; 2(2): 110-115.
- 25. Saravana Kumar K, et. al., A Review on Microsphere for Novel drug delivery System, Journal of Pharmacy Research, 2012; 5(1): 420-424.
- 26. Harsh Bansal, et. al., Microsphere: Methods of Prepration and Applications; a Comparative Study, International Journal of Pharmaceutical Sciences Review and Research, 2011; 10(1): Article-012.
- 27. Orienti I, Aiedeh K, Gianasi E, Bertasi V, Zecchi V. Indomethacin loaded chitosan microspheres correlation between the erosionprocess and release kinetics. J Microencapsul, 1996; 13: 463-72.
- 28. Nair R, Reddy B. Application of chitosan microspheres as drug carriers: a review. J Pharm Sci Res, 2009; 1: 1-12.
- 29. Saravana Kumar K, et. al., A Review on Microsphere for Novel drug delivery System, Journal of Pharmacy Research, 2012; 5(1): 420-424.
- 30. Patrick B. O'Donnell, et. al., Preparation of microspheres by the solvent evaporation technique, Advanced Drug Delivery Reviews, 1997 28: 25–42.
- 31. Mathew Sam T., Devi Gayathri S., Prasanth V.V., Vinod B; NSAIDs as microspheres, The Internet Journal of Pharmacology, 2008; 6(1): 67–73.
- 32. Davis S.S. and Illum L. (1989). Microspheres as drug carrier in drug carrier system, F.H Roerdink and A.M.Kron (Eds), John Wiley and sons Ltd, 1-6.
- 33. Xinghang Ma, et. al., Stability Study of Drugloaded Proteinoid Microsphere Formulations during Freeze-drying, Iournal of Drug Targeting, 1994; 2: 9-21.
- 34. Mathew Sam T., Devi Gayathri S., Prasanth V.V., Vinod B; NSAIDs as microspheres, The Internet Journal of Pharmacology, 2008; 6(1): 67–73.
- 35. Schugens C., Larvelle. N., Nihantn., Grandfils C., Jerome R. and Teyssie. P. J. Control. Rel, 1994; 32: 161-167.
- 36. Patitapabana Parida, et. al., Development and characterization of ethylcellulose based microsphere for sustained release of nifedipine, journal of pharmaceutical analysis, 2016; 6(5): 341-344.
- 37. Navid Jubaer Ayon, et. al., Preparation and Characterization of Gliclazide Incorporated Cellulosic Microspheres: Studies on Drug Release, Compatibility and Micromeritics, Dhaka Univ. J. Pharm. Sci, 2014; 13(2): 149-166.

- 38. Rakesh Gupta, et. al., Characterization of Captopril-Ethyl Cellulose Microspheres by Thermal Analysis, International Journal of Drug Development and Research, April-June 2010; 2(2): 394-398.
- 39. Abhay M.L, et. al., Formulationa and Characterization of Microspheres of Artemether, Literati Journal of Pharmaceutical Drug Delivery Technologies, 2015; 1(2): 65-69.
- 40. Funden berg H.H., Stites D.P., Caldwel J.L. and Wells J.V. (1978) In: Basic and clinical immunology, 2 ed., Lange Medical, Los Altosca.
- 41. Capron A.C., Locht C. and Fracchia G.N (1994) Vaccine.12, 667; Edelman R. vaccine 11, 1361; Drews J. (1984) Immunostimulantien, Klin. Wochenscher.62, 254; Spier K.E. (1993) vaccine 11, 1993, 1450.
- 42. Abbaraju Krishna shailaja, et. al., Biomedical applications of microspheres, Journal of Modern Drug Discovery and Drug Delivery Research, 2015; 4(2): 1-5.
- 43. Kazi M. Zakir Hossain, et. al., Development of microspheres for biomedical applications: a review, Prog Biomater, 2014; 1-19.
- 44. Kataria Sahil, et. al., Microsphere: A Review, International Journal of Research in Pharmacy and Chemistry, 2011; 1(4): 1184-1198.
- 45. Reza Arshady, et. al., Microspheres for biomedical applications: preparation of reactive and labeled microsphere, biomaterials, 1993; 14(1): 5-15.
- 46. Manoj Kumar Das, Microsphere a Drug Delivery System–A Review, International Journal of Current Pharmaceutical Research, 2019; 11(4): 34-41.
- 47. Kalyan Shweta, Sharma Parmod Kumar et al. Recent Advancement In Chitosan Best Formulation And Its Pharmaceutical Application. Pelagia Research Library, 2010; 1(3): 195-210.