

**EXTRACTION OF DYES FROM PARTS OF THE PLANTS AND THEIR
PHYTOCHEMICAL SCREENING****S. Sashikala^{1*}, S. Sharmila² and A. Nousheen Iffath³**

¹Assistant Professor, Department of Chemistry, D.K.M College for Women, Vellore, Tamil Nadu, India.

^{2,3}UG students, D.K.M College for Women, Vellore, Tamil Nadu, India.

Article Received on
14 May 2024,

Revised on 04 June 2024,
Accepted on 25 June 2024

DOI: 10.20959/wjpr202413-32915



***Corresponding Author**

Dr. S. Sashikala

Assistant Professor,
Department of Chemistry,
D.K.M College for Women,
Vellore, Tamil Nadu, India.

ABSTRACT

The primary goal of obtaining dyes from natural plant sources is to prevent pollution of the environment. Any colour, pigment or material originating from organic materials plants, animals or minerals are considered as a natural dye, since it is a sustainable bioresource with no negative influence on the environment. The oldest known kind of dyeing is arguably the use of natural dyes.

Nowadays, natural dyes are widely used in the textile sector because synthetic colours are harmful to human skin. Much study is being done globally on the use of natural dyes in the textile industry in light of the current worldwide concern about the usage of eco-friendly and biodegradable products. They have been utilized for colouring food items, textiles, natural protein fibres like wool, silk, cotton and leather. Natural dye was initially used by humans as a thought tool to create

artwork that reflected both their environment and themselves. Synthetic dyes have effluent issues not only when they are used in the textile industry but also when they are made and may even arise while synthesising other raw materials and intermediates. After synthetic dyes were discovered in 1856, the usage of natural dyes for textile dyeing was greatly reduced. Natural dyes have been used to color fabrics since ancient times, the discovery of mauve colorant in the 19th century led to the replacement of natural dyes with synthetic ones. Naturally occurring dyes have become a significant substitute for artificial dyes. Furthermore, the plant dyes might serve as a substitute for synthetic dyes when it comes to dyeing natural cotton fibre. In order to verify the plant dyes' qualities and introduce them into textiles more quickly and efficiently, this process is used to first identify plants

from which dyes can be derived. In the present study, the phytochemical Screening was carried from natural dyes which were obtained from different plant parts, namely *Ixora Coccinea* Flower (pink), *Nerium Oleander* Flower (red), *Tradescantia Pallida* flower, *Portulaca Oleracea*, *Cissus Qudrangularis* stem and *Celosia Cristata* stem. The extracted dyes were categorized for different chemical groups like alkaloids, flavonoids, tannins, saponins, carbohydrates, amino acids etc., using various chemical reactions.

KEYWORDS: Natural dyes, Biodegradable, Phytochemical Screening, *Ixora Coccinea* Flower (pink), *Nerium Oleander* Flower (red), *Tradescantia Pallida* flower, *Portulaca Oleracea*, *Cissus Quadrangularis* stem and *Celosia Cristata* stem.

INTRODUCTION

The utilization of plants for dyes is considered crucial. Lately, there has been a significant increase in interest towards natural dyes, mainly due to the strict environmental regulations imposed by numerous countries in response to the harmful effects and allergic reactions caused by synthetic dyes. As a result, with a distinct lowering in synthetic dyestuff costs, the natural dyes were virtually unused at the beginning of twentieth century (Kumaresan et. al., 2011). Currently, in the majority of nations, natural dyeing is predominantly carried out as a traditional craft, while synthetic dyes have become the norm in all industrial dyeing procedures. However, with the worldwide concern over the use of eco-friendly and biodegradable materials, the use of natural dyes has once again gained interest (Agarwal A, Goel A & Gupta K C, 1992).

Dyeing can be conducted in an alkaline, acidic, or neutral bath. Numerous studies have been published on various techniques of mordanting for different types of fibres, including cellulosic, protenic, and synthetic fibers, when dyeing with natural dyes. By utilizing different mordants, a wide range of shades can be achieved, from one colour to another. Dyeing of cotton and silk with henna, indigo, marigold etc is reported. (Gulrajani et al, 1992). There is a growing interest in the revival of natural dyes in textile colouration (Mehanta z. & Osman et al, 2003 & 2004). In contrast, natural dyes are environmental friendly, exhibit better biodegradability and generally have a higher compatibility with the environment than synthetic dyes. (Ahlstrom et al, 2005). The process is economically feasible since the raw materials are readily available at a low cost, resulting in a significantly low production cost as well.

MATERIALS AND METHODS

The dye was extracted from the fresh plant parts of *Ixora Coccinea* Flower (pink), *Nerium Oleander* Flower (red), *Tradescantia Pallida* flower, *Portulaca Oleracea*, *Cissus Quadrangularis* stem and *Celosia Cristata* stem, which were collected from the nearby villages of Vellore district, Vellore in the month of December, 2023.

Materials Required

- Plant materials
- Cotton cloths
- Scissor
- Beakers
- Bowls
- Glass rods
- Test tubes
- Test tube stand
- Funnel
- Filter paper
- Tripod stand
- Conical flask
- Mordants
- Knife
- Filter paper
- Mesh

Plants Used

- *Ixora Coccinea* Flower (pink)
- *Nerium Oleander* Flower (red)
- *Tradescantia Pallida* (flower)
- *Portulaca Oleracea* (flower and leaves)
- *Cissus Quadrangularis* stem and
- *Celosia Cristata* stem.

Extraction of Dyes from the Plants

The flowers of various plant materials were collected from the locality. The plant material was weighted using chemical balance (10 gram). It was thoroughly washed with distilled

water and allowed to dry. The chopped flower material was dissolved in 100 ml of distilled water in beaker and was heated for 30 minutes at a temperature range of 80-85°C, until the dye was released. The dye from aqueous extraction was filtered through a funnel and filter paper. After the dye was extracted, it was stored in a closed flask, in the refrigerator for further use.

PREPARATION OF MORDANT

Mordant

The creation of a bond between the colouring matter and fibre is called mordanting.

Copper Sulphate

5g of Copper Sulphate was dissolved in 100 ml of distilled water.

Ferrous Sulphate

5g of Ferrous Sulphate was dissolved in 100 ml of distilled water.

Potassium Chromate

5g of Potassium Chromate was dissolved in 100 ml of distilled water.

Preparation of the cloth

The cloth material (cotton) is cut into small pieces (10*10 cm) and it is dissolved in sodium hydroxide and refluxed for 15 minutes to remove the starch; cellulose and other dirt particles from it.

The cloth treated with sodium hydroxide was put in the mordant solution and simmered for 15 minutes and then it is taken out.

Application of dye to fabrics: The dye is applied to fabric by two methods

- **Without Mordant**
- **With mordant**

Without mordant: The fabric which is treated with sodium hydroxide is directly immersed at the dye bath and the fabric is simmered for half an hour. After the dye enters through the cloth. The cloth is taken out and dried for further studies.

With mordant: The mordanted fabric is immersed in dye bath for half an hour. After that the dye enters through the cloth. The cloth is taken out and dried.

Phytochemical Test**(a) Detection of Phenols****❖ Ferric Chloride Test**

To 3ml of the extracts 3-4 drops of ferric chloride solution was added. Formation of bluish black colour, indicates the presence of Phenols.

(b) Detection of tannins**❖ Gelatine test**

To 3ml of the extracts 1% gelatine solution containing sodium chloride was added. Formation of white precipitate, indicates the presence of tannins.

(c) Detection of Flavonoids**❖ Alkaline Reagent**

To 3ml of the extracts sodium hydroxide solution was added. Formation of intense yellow colour, which becomes colourless on the addition of dilute acids, indicates the presence of Flavonoid.

❖ Lead acetate Test

To 3ml of the few drops of lead acetate solution was added. Formation of yellow precipitate, indicates the presence of Flavonoid.

(d) Detection of Saponins**❖ Foam test**

1ml of the extracts was diluted with distilled water to 20 ml and shaken in a graduated cylinder for 15 minutes. Development of stable foam indicates the presence of saponins.

❖ Froth Test

1ml of the extracts taken in various test-tubes was stoppered and shaken vigorously for about 5 min, it was allowed to stand for 30 min and observed for honeycomb froth, which was indicative of the presence of saponins.

❖ Lead acetate Test

To 1ml of the extracts, 1% lead acetate solution was added. Formation of white precipitates indicates the presence of saponins

(e) Detection of Steroids

A few drops of acetic anhydride are added to the extracts and the formation of violet to blue to green in some samples indicates the presence of steroids.

(f) Quinine

1ml of the extracts, was mixed with few drops of concentrated hydrochloric acid; appearance of green colour indicates the presence of quinine.

(g) Cellulose

1ml of the extracts, was mixed with iodine crystals and then a few drops of conc. sulphuric acid are added. Appearance of brown colour indicates the presence of cellulose.

(h) Detection of Terpenoids**❖ Salkowski's Test**

Extract of 5 mg of the selected plant part is mixed with 2 mL chloroform and 3 mL concentrated sulfuric acid added carefully to form a layer. A reddish-brown colour indicates the presence of terpenoids.

(i) Detection of Glycosides

The leaf extracts were individually hydrolysed with dilute HCl and filtered.

❖ Legal's Test

To 3ml of the extracts Sodium Nitroprusside in Pyridine and Sodium Hydroxide was added. Formation of red colour, indicates the presence of glycosides.

❖ Borntrager's Test

To 3mL of the extracts, 5ml dilute HCl and 5ml FeCl₃ was added. The mixture was cooled and extracted with equal volumes of benzene. The benzene layer was separated and treated with ammonia solution. Formation of red/pink colour, indicates the presence of glycosides.

(j) Fat and fixed oil

1ml of the extracts, was mixed with Sudan III which results in shining orange which indicates the presence of fat and fixed oil.

Table 1: Phytochemical Test.

		E1	E2	E3	E4	E5	E6	
(a)	Phenol	+	+	+	+	+	+	Dye from aqueous extraction E1-Ixora Coccinea Flower (pink) E2-Nerium Oleander Flower (red) E3-Tradescantia Pallida (flower) E4-Portulaca Oleracea (flower and leaves) E5-Cisus Quadrangularis stem and E6-Celosia Cristata stem.
(b)	Tannin	+	+	—	+	+	+	
(c)	Flavonoids	+	+	+	+	+	+	
(d)	Saponin	+	—	+	+	+	+	
(e)	Steroids	+	+	—	+	—	+	
(f)	Quinine	—	—	—	—	—	—	
(g)	Cellulose	+	+	—	+	+	+	
(h)	Terpenoids	+	+	—	+	+	+	
(i)	Glycosides	+	—	—	+	+	—	
(j)	Fat and fixed oil	—	—	—	+	—	—	

+: Present; -: Absent

RESULTS AND DISCUSSION

The color of the dye extracted from the plant is determined by the compounds present in that particular plant. The colorant is subsequently applied to the cotton fabric to ensure color permanence. The fabric showcased above has successfully bonded with the appropriate dye through the assistance of a mordant. By incorporating these mordants into the dye, various hues of color were achieved, allowing us to create diverse shades from a single plant with the aid of the mordant.

Preliminary phytochemical screening of the all the extracts showed the presence of Phenols, Tannins, Flavonoids, Saponins and Glycosides. This screening test on the extracts showed the absence of Quinines, Fat and Fixed oils in maximum number of plant parts.



Fig 1a: Ixora Coccinea Flower (pink)



Fig 1b: Cloth with and without mordants



**Fig 1c:
Phytochemical
Screening of Ixora
Coccinea Flower**



**Fig 2a: Nerium Oleander
Flower (red)**



**Fig 2b: Cloth with and without
mordants**



**Fig 2c:
Phytochemical
Screening of Nerium
Oleander Flower**



**Fig 3a: Tradescantia
Pallida (Flower)**



**Fig 3b: Cloth with and without
mordants**



**Fig 3c:
Phytochemical
Screening of
Tradescantia Pallida
Flower**



**Fig 4a: Portulaca Oleracea
(leaves)**



**Fig 4b: Cloth with and without
mordants**



**Fig 4c:
Phytochemical
Screening of
Potulaca Oleracea
leaves**



**Fig 5a: Cissus
Quandrangularis (stem)**



**Fig 5b: Cloth with and without
mordants**



**Fig 5c:
Phytochemical
Screening of Cissus
Quandrangularis
Stem**



**Fig 6a: Celosia Cristata
(stem)**



**Fig 6e: Cloth with and without
mordants**



**Fig 6c:
Phytochemical
Screening of Celosia
Cristata Stem**

REFERENCE

1. Agarwal A, Goel A & Gupta K C: Textile Dyers and Printer, 1992; 25(10): 28.
2. Ahlström L., Eskilsson C. S., and Björklund E: Determination of banned azo dyes in consumer goods, Trends Anal. Chem., 2005; 24(1): 49-56.
3. Gulrajani M L & Gupta D: Natural dyes and application to textiles, Department of textile technology, Indian Institute of Technology, New Delhi, India, 1992.
4. Kumaresan M, Palanisamy P N and Kumar P E: Application of Eco-friendly Natural dye obtained from flower of Spathodea campanulata on silk using combination of mordants, Eur J Sci Res., 2011; 52(3): 306- 312.
5. Mehanta Z. A, and Tiwari I. A: Natural dye-yielding plants and indigenous knowledge on dye preparation in Arunachal Pradesh Northeast India, Curr. Sci., 2003; 88(4): 1474-1480.
6. Olymers, 9: 334-340, Jadhao, N.U., Rathod, S.P. The extraction process and antioxidant properties of patuletin dye from wasted temple french marigold flower. Asian Journal of Plant Science and Research, 2013; 3: 127-132.
7. Khan, M.I., Pharmacological potentials of phenolic compounds from Prosopis spp-a review. Hans-Uwe Dahmer Ahmad, A., Khan, S.A., Yusuf, M., Shahid, M., Manzoora, N., Mohammad, F., Assessment of antimicrobial activity of catechu and its dyed substrate. Journal of Clean Production, 2011; 29: 1385-1394.
8. Kulkarni, S.S., Gokhale, A.V., Bodake, U.M., Pathade, G.R. Cotton dyeing with natural dye extracted from Pomogranate (punica granatum) peel. Universal Journal of Environment Research & Technology, 2011; 1: 135-139.
9. Sara Kadolph, Natural Dyes: A.N.M. Aljiamali, Biochem. Anal. Biochem, 2015; 4: 1-4. [Crossref], [Google Scholar], [publisher]
10. Traditional Craft Experiencing New Attention, The Delta Kappa Gamma Bulletin.
11. In vivo antimalarial extracts and constituents of Prosopis Juliflora (Fabaceae).

12. M.L. Cardozo, R.M. Ordonez, I.C. Zampini, A.S. Cuello, G. Dibenedetti, and M.I. Isla, "Evaluation of antioxidant capacity, genotoxicity and polyphenol content of non conventional foods: propolis flour", Food Research International, 2010; 43(5): 1500-1510.